APPENDICES

40 Year Water Quality Report (1984-2024)

Environmental Commission Township of Stillwater, New Jersey

Appendix No.	Description
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Appendix A

Prior Stream Names and Stations Excerpted from the 2004 Surface Water Report*

*S.Grodsky, 2004. Stream Surface Water Sampling – Chemical Data and Biological Analysis through Macroinvertebrates. Prepared for the Stillwater Township Environmental Commission. 2004.

Stillwater Township

Stream Surface Water Sampling – Chemical Data and Biological Analysis through Macroinvertebrates

Prepared For The Stillwater Township Environmental Commission

> By Steve Grodsky

Sampling Stations and Dates

Sampling stations were selected in four watersheds within Stillwater Township at the following locations to evaluate trends in water quality:

Station Number	Sampling Station	Trout	Location
·		Identification	a a
1	Blair Creek	Trout Maintenance	Off of Old Schoolhouse Rd - along dirt road below confluence of North and South branch of Blair Creek
· 2	Trout Brook	Trout Production	Owassa Rd. at the junction with Fairview lake Rd.
3	Trout Brook	Trout Brook Trout Production	
4	Spring Brook (Quick Pond)	Trout Maintenance	Mount Benevolence Rd. near Crandon Lodge
5	Spring Brook (Crandon)	Trout Maintenance	Hampton Rd. Bridge
6	Spring Brook	Trout Maintenance	Swartswood Rd. Bridge
7	Paulinskill River	Trout Maintenance	Stillwater Rd. – Main St. Bridge
8	Keen's Mill	Trout Maintenance	Off of Rt. 521 - next to Keen's Mill

Dates of sampling range from the beginning of the project in the spring of 1984 to present, the most recent sampling being the fall of 2003. All water samples were collected in accordance with the New Jersey Department of Environmental Protection (NJDEP) "Field Procedures Manual for Water Data Acquisition" (NJDEP Division of Water Resources) and were remanded to a licensed environmental laboratory (QC Laboratories - Southampton, PA) for analysis.

^{*}No sampling took place during the years of 1991, 1992, 1994, and 1995. As a result, the graphical representation of the data will exclude these time periods.

Appendix B NJDEP SWQS Excerpts, Screening Levels and References

40 Year Water Quality Report (1984-2024)
Environmental Commission - Township of Stillwater, NJ

Summary of Surface Water Qulaity Criteria (SWQC) and Screenineg Levels (SLs) used for 2024 SWMP Report

Parameter	Units	General	Acute	Chronic	НН	TP	TM	NT	Notes	Source
Alkalinity	mg/L	20 - 200							(1) (1A)	USEPA; NMSU
Aluminum	mg/L		0.75	0.087					(2)	CDC_ASTDR
E. coli	Counts/100 ml	100 GM; 320 STV			100				(3)	NJAC 7:9B
Fecal Coliforms	Counts/100 ml	100 GM; 320 STV							(3A)	NJAC 7:9B
Lead (ug/L)	mg/L		0.038(d)(s)	0.0054(d)(s)	0.005(h)(T)					NJAC 7:9B
Nitrogen, Ammonia	mg/L		2.4	0.54					(4)	NJAC 7:9B
Nitrate (as N)	mg/L				10(h)					NJAC 7:9B
Nitrite (as N)	mg/L				1				(5)	NJAC 7:10
Nitrogen,Total Kjeldhal (TKN)	mg/L	6							(6)	USEPA
Ortho Phosphate as P	mg/L	0.1							(7)	
Phosphorus (Total)	mg/L	0.1								NJAC 7:9B
рН	su	6.5-8.5								NJAC 7:9B
Specific Conductance	umhos/cm	500							(8)	USEPA
Total Dissolved Solids (TDS)	mg/L								(9)	NJAC 7:9B; NJAC 7:10
Total Suspended Solids (TSS)	mg/L					25	25	40		NJAC 7:9B

Notes

- (d) Criterion is expressed as a function of the Water Effect Ratio (WER). For criterion in the table, WER equates to the default value of 1.0
- (h) Human health noncarcinogen
- (s) Dissolved criterion
- (T) Total recoverable criterion
- (1) https://www.epa.gov/wqc/national-recommended-water-quality-criteria-aquatic-life-criteria-table (acessed 03/22/2024)
- (1A) https://aces.nmsu.edu/pubs Cooperative Extension Servise Guide W-104 (acessed 09/01/2024)
- (2) CDC_ASTDR Toxilogical Profiles; www.atsdr.cdc.gov/ToxProfiles/tp22-c8.pdf
- (3) See NJAC7:9B. Primary Contact Recreation;

Max E. coli geometric mean of 100/100 ml per 90-day period and STV of 320/100 ml with no greater than 10 percent excursion frequency over 90-days.

- (3A) SWQC-based screening level for E.coli used for Fecal coliform SWMP data (see report for basis and background).
- (4) Calculated per NJAC 7:9B using field measured pH (8.45) and temp (14.5 c) from October 16, 2024 sampling event.
- (5) Screening Level (SL) based on NJDEP Safe Drinking Water MCL per NJAC 7:10.
- (6) USEPA "Total Nitrogen" https://nepis.epa.gov (acessed 09/01/2024)
- (7) Phosphorus SWQC adopted as SL.
- (8) https://archive.epa.gov/water/archive/web/html/vms59.html (acessed 09/01/2024)
- (9) Screening Level (SL) based on NJDEP Safe Drinking Water Secondary Standards per NJAC 7:10; Background evaluation and WET test data unavailable for SWQC.

7:9B-1.14(d) General Surface Water Quality Criteria for FW2, SE and SC Waters: (Expressed as Maximum concentrations unless otherwise noted)

Substance		luxiiiiui	Criteria	Classifications
1. <mark>Bacteri</mark> ml)	<mark>al quality</mark> (Counts/100	i.	Shellfish Harvesting: Bacterial Indicators shall not exceed, in all shellfish waters, the standard for approved shellfish waters as established by the National Shellfish Sanitation Program as set forth in its current manual of operations.	Shellfish Waters
	İ	ii.	Primary Contact Recreation:	
		(1) Enterococci levels shall not exceed a geometric mean of 30/100 ml over a 90-day period and a Statistical Threshold Value of 110/100 ml, which shall not be greater than 10 percent excursion frequency over a 90-day period.	SE1 and SC
		(2	E. coli levels shall not exceed a geometric mean of 100/100 ml over a 90-day period and a Statistical Threshold Value of 320/100 ml, which shall not be greater than 10 percent excursion frequency over a 90-day period.	All FW2
	i	iii. <mark>S</mark>	econdary Contact Recreation:	
		(1) Fecal coliform levels shall not exceed a geometric mean of 770/100 ml.	SE2
		(2) Fecal coliform levels shall not exceed a geometric mean of 1500/100ml.	SE3
2. Dissolve	ed oxygen (mg/L)	i.	Not less than 7.0 at any time;	FW2-TP
	i		24 hour average not less than 6.0. Not less than 5.0 at any time (see paragraph viii below);	FW2-TM

7:9B-1.14(d) General Surface Water Quality Criteria for FW2, SE and SC Waters: (Expressed as Maximum concentrations unless otherwise noted) e Criteria Classification

	Substance		Criteria	Classifications
		iii.	24 hour average not less than 5.0, but not less than 4.0 at any time (see paragraph viii below);	FW2-NT (except as in iv below), SE1
		iv.	Not less than 4.0 at any time;	Tidal portions of FW2-NT tributaries to the Delaware River, between Rancocas Creek and Big Timber Creek inclusive.
		v.	Not less than 5.0 at any time;	SC
		vi.	Not less than 4.0 at any time;	SE2
		vii.	Not less than 3.0 at any time; and	SE3
		viii.	Supersaturated dissolved oxygen values shall be expressed as their corresponding 100 percent saturation values for purposes of calculating 24 hour averages.	FW2-TM, FW2-NT, SE1
3.	Floating, colloidal, color and settleable solids; petroleum hydrocarbons and other oils and grease	i.	None noticeable in the water or deposited along the shore or on the aquatic substrata in quantities detrimental to the natural biota. None which would render the waters unsuitable for the designated uses.	All Classifications

7:9B-1.14(d) General Surface Water Quality Criteria for FW2, SE and SC Waters:

(Expressed as Maximum concentrations unless otherwise noted)

Substance

Criteria

Classifications

4. Nutrients

- i. Except as due to natural conditions, nutrients shall not be allowed in concentrations that render the waters unsuitable for the existing or designated uses due to objectionable algal densities, nuisance aquatic vegetation, diurnal fluctuations in dissolved oxygen or pH indicative of excessive photosynthetic activity, detrimental changes to the composition of aquatic ecosystems, or other indicators of use impairment caused by nutrients.
- ii. Phosphorus (mg/L)*
 - (1) Non Tidal Streams: Concentrations of total P shall not exceed 0.1 in any stream, unless site-specific criteria or watershed-specific translators are established pursuant to N.J.A.C. 7:9B-1.5(g)2 or if the Department determines that concentrations do not render the waters unsuitable in accordance with (d)4i. above.

All Classifications

(2) Lakes: Concentrations of total P shall not exceed 0.05 in any lake, pond or reservoir, or in a tributary at the point where it enters such bodies of water, unless site-specific criteria or watershed-specific translators are developed pursuant to N.J.A.C. 7:9B-1.5(g)2 or if the Department determines that concentrations do not render the waters unsuitable in accordance with (d)4i. above.

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7:9B-1.14(d) General Surface Water Quality Criteria for FW2, SE and SC Waters: (Expressed as Maximum concentrations unless otherwise noted)

	Substance (Expressed as	IVIUXIIII	Criteria Criteria	Classifications
5.	pH (Standard Units)	i.	6.5-8.5	FW2 waters listed at 1.15(d), (f), (g) and (i), All SE
		ii.	4.5 - 7.5	FW2 waters listed at 1.15(c), (e) and (h)
		iii.	Natural pH conditions shall prevail.	SC
6.	Radioactivity	i.	Prevailing regulations including all amendments and future supplements thereto adopted by the U.S. Environmental Protection Agency pursuant to Sections 1412, 1445, and 1450 of the Public Health Services Act, as amended by the Safe Drinking Water Act (PL 93-523)	All Classifications
7.	Solids, Suspended (mg/L) (Non-filterable residue)	i.	25.0	FW2-TP, FW2-TM
		ii.	40.0	FW2-NT
		iii.	None of which would render the water unsuitable for the designated uses.	All SE, SC
8.	Solids, Total Dissolved (mg/L) (Filterable Residue)	i.	No increase in background which may adversely affect the survival, growth or propagation of the aquatic biota. Compliance with water quality-based WET limitations or $LC_{50} \ge 50$ percent, whichever is more stringent, shall be deemed to meet this requirement.	FW2

7:9B-1.14(d) General Surface Water Quality Criteria for FW2, SE and SC Waters: (Expressed as Maximum concentrations unless otherwise noted)

	Substance		Criteria Criteria	Classifications
		ii.	No increase in background which would interfere with the designated or existing uses, or 500 mg/L, whichever is more stringent.	FW2
		iii.	None which would render the water unsuitable for the designated uses.	All SE
9.	Sulfate (mg/L)	i.	250	FW2
10.	Taste and odor producing substances	i.	None offensive to humans or which would produce offensive taste or odors in water supplies and biota used for human consumption. None which would render the water unsuitable for the designated uses.	All Classifications
11.	Temperature	i.	Temperatures shall not exceed a daily maximum of 22 degrees Celsius or rolling seven-day average of the daily maximum of 19 degrees Celsius, unless due to natural conditions	FW2-TP
		ii.	Temperatures shall not exceed a daily maximum of 25 degrees Celsius or rolling seven-day average of the daily maximum of 23 degrees Celsius, unless due to natural conditions	FW2-TM
		iii.	Temperatures shall not exceed a daily maximum of 31 degrees Celsius or rolling seven-day average of the daily maximum of 28 degrees Celsius, unless due to natural conditions	FW2-NT
		iv.	No thermal alterations which would cause temperatures to exceed 29.4 degrees Celsius (85 degree Fahrenheit) Summer seasonal average	SE

7:9B-1.14(d) General Surface Water Quality Criteria for FW2, SE and SC Waters: (Expressed as Maximum concentrations unless otherwise noted)

	Substance (Expressed a	as Maxim	um concentrations unless otherwise noted) Criteria	Classifications
		V.	No thermal alterations which would cause temperatures to exceed 26.7 degrees Celsius (80 degree Fahrenheit) Summer seasonal average	SC
12.	Toxic Substances (general)	i.	None, either alone or in combination with other substances, in such concentrations as to affect humans or be detrimental to the natural aquatic biota, produce undesirable aquatic life, or which would render the waters unsuitable for the designated uses.	All Classifications
		ii.	None which would cause standards for drinking water to be exceeded after appropriate treatment.	FW2
		iii.	Toxic substances shall not be present in concentrations that cause acute or chronic toxicity to aquatic biota, or bioaccumulate within an organism to concentrations that exert a toxic effect on that organism or render it unfit for consumption.	All Classifications
		iv.	The concentrations of nonpersistent toxic substances in the State's waters shall not exceed one-twentieth (0.05) of the acute definitive LC_{50} or EC_{50} value, as determined by appropriate bioassays conducted in accordance with N.J.A.C. 7:18.	All Classifications
		V.	The concentration of persistent toxic substances in the State's waters shall not exceed one-hundredeth (0.01) of the acute definitive LC_{50} or EC_{50} value, as determined by appropriate bioassays conducted in accordance with N.J.A.C. 7:18.	All Classifications

7:9B-1.14(d) General Surface Water Quality Criteria for FW2, SE and SC Waters: (Expressed as Maximum concentrations unless otherwise noted)

	(Expressed Substance	as Maxim	um concentrations unless otherwise noted) Criteria	Classifications
13.	Turbidity (Nephelometric Turbidity Unit-NTU)	i.	Maximum 30-day average of 15 NTU, a maximum of 50 NTU at any time.	FW2, SE3
		ii.	Maximum 30-day average of 10 NTU, a maximum of 30 NTU at any time.	SE1, SE2
		iii.	Levels shall not exceed 10.0 NTU.	SC

^{*} See N.J.A.C. 7:9B-1.14(g) for site-specific criteria.

(e) Surface Water Quality Criteria for Ammonia are derived in accordance with the formulas set forth below. Acute criteria are expressed as three-hour average using MA1CD10 flow and chronic criteria are expressed as 30-day average using MA30CD10 flow. No exceedance of criteria shall be permitted at or above the design flows specified.

<u>Cas Number</u> <u>Criteria</u> <u>Classification</u>

- 1. Ammonia, unionized (mg NH₃-N/L)
- i. at pH < 8.30

PL

 $0.238*10^{0.026(\text{Temp-20}) + 0.41 \text{ (pH-7.80)}} \text{ (acute)}$

 $0.061*10^{0.026(\text{Temp-20}) + 0.41 \text{ (pH-7.80)}} \text{ (chronic)}$

ii. 0.115 (acute); 0.030 (chronic)

All SE

iii. 0.094 (acute); 0.024 (chronic)

SC

2. Ammonia, Total (mg TAN/L) - Acute criteria are expressed as one-hour average using MA1CD10 flow, chronic criteria are expressed as 30-day rolling average using MA30CD10 flow and the highest four-day average within the 30-day averaging period should not be more than 2.5 times of chronic criteria. No exceedance of criteria shall be permitted at or above the design flows specified.

CAS Number

Criteria applicable in FW2 waters

Classifications

3. Ammonia, total (mg TAN/L)

7664-41-7

FW2-TP.

FW2-TM

1.

$$\operatorname{MIN} \left(\frac{\left(\frac{0.275}{1 + 10^{7.204 - pH}} + \frac{39.0}{1 + 10^{pH - 7.204}} \right),}{\left(0.7249 \times \left(\frac{0.0114}{1 + 10^{7.204 - pH}} + \frac{1.6181}{1 + 10^{pH - 7.204}} \right) \times \left(23.12 \times 10^{0.036 \times (20 - T)} \right) \right) \right) \text{ (acute)}$$

ii. $0.7249 \times \left(\frac{0.0114}{1 + 10^{7.204 - pH}} + \frac{1.6181}{1 + 10^{pH - 7.204}}\right) \times MIN(51.93,23.12 \times 10^{0.036 \times (20 - T)}) \text{ (acute)}$

iii. All FW2 $0.8876 \times \left(\frac{0.0278}{1 + 10^{7.688 - pH}} + \frac{1.1994}{1 + 10^{pH - 7.688}}\right) \times \left(2.126 \times 10^{0.028 \times (20 - MAX(T,7))}\right) \text{ (chronic)}$

- (f) Surface Water Quality Criteria for Toxic Substances are as follows:
 - 1. Acute aquatic life protection criteria are determined with no exceedance at or above the MA1CD10 flow and expressed as one-hour average except,
 - i. for copper the criteria are expressed as 24-hour average, and
 - ii. for cadmium, chromium, lead, mercury, nickel, silver, and zinc the criteria are expressed as 6-hour average.
 - 2. Chronic aquatic life protection criteria are determined with no exceedance at or above the MA7CD10 flow and expressed as four-day average.
 - 3. Freshwater aquatic criteria for cadmium, chromium III, copper, nickel, silver, and zinc are expressed as a function of water hardness. Criteria can be calculated at any hardness using these equations as listed below. Criteria thus calculated are multiplied by appropriate conversion factor (CF) to convert total recoverable metal into dissolved metal and by the default Water Effect Ratio (WER) of 1.0.

General formula $WER [e^{(V[ln (hardness)] + ln A - V[ln Z])}] CF$

where:

V = pooled slope

A = FAV at given hardness

Z = selected value of hardness

Cadmium:

Acute dissolved criterion WER $[e^{(1.0166 (ln [hardness])-3.924)}] 0.651$

Chronic dissolved criterion WER $[e^{(0.7409 (ln [hardness])-4.719)}] 0.651$

Chromium III:

Acute dissolved criterion WER $[e^{(0.819 (ln [hardness])+3.7256)}] 0.277$

Chronic dissolved criterion WER $[e^{(0.819 (ln [hardness])+0.6848)}] 0.277$

Copper:

Acute dissolved criterion WER $[e^{(0.9422 (ln [hardness])-1.7)}]$ 0.908

Chronic dissolved criterion WER $[e^{(0.8545 (ln [hardness])-1.702)}]$ 0.908

Nickel:

Acute dissolved criterion WER $[e^{(0.846 (ln [hardness]) + 2.255)}] 0.846$

Chronic dissolved criterion WER $[e^{(0.846 (ln [hardness])+0.0584)}] 0.846$

Silver:

Acute dissolved criterion WER $[e^{(1.72 (ln [hardness])-6.59)}] 0.85$

Zinc:

Acute or dissolved criterion WER $[e^{(0.8473 (ln [hardness])+0.884)}] 0.950$

Chronic dissolved criterion WER $[e^{(0.8473 (ln [hardness])+0.884)}] 0.950$

4. Freshwater criteria for pentachlorophenol are expressed as a function of pH. Criteria are derived in accordance with the formula set forth below:

Acute criterion = $e^{(1.005[pH]-4.869)}$

Chronic criterion = $e^{(1.005[pH]-5.134)}$

- 5. Human health noncarcinogenic effect-based criteria are expressed as a 30-day average with no frequency of exceedance at or above the MA7CD10 flow.
- 6. Human health carcinogenic effect-based criteria are based on a risk level of one-in-one-million and are expressed as a 70-year average with no frequency of exceedance at or above the design flow as specified at N.J.A.C. 7:9B-1.5(c)2iii.

7. SURFACE WATER QUALITY CRITERIA FOR TOXIC SUBSTANCES: $(\mu g/L)$

$(\mu g L)$									
Toxic Substance	CAS		esh Water (F	W2) Criteria	Saline Water (SE & SC) Criteria				
TOXIC Substance	Number	Aqui	atic Chronic	Human Health	Aqu	atic Chronic	Human Health		
Acenaphthene	83-32-9	Acute	Chronic	670(h)	Acute	Chronic	990(h)		
Acrolein	107-02-8			6.1(h)			9.3(h)		
Acrylonitrile	107-13-1			0.051(hc)			0.25(hc)		
Aldrin	309-00-2	3.0		0.000049(hc)	1.3		0.000050(hc)		
Ammonia, un-ionized	<mark>7664-41-7</mark>	See N.J.A 1.14			See N.J.A 1.14				
Anthracene	120-12-7			8,300(h)			40,000(h)		
Antimony	7440-36-0			5.6(h)(T)			640(h)(T)		
Arsenic	7440-38-2	340(d)(s)	150(d)(s)	0.017(hc)(T)	69(d)(s)	36(d)(s)	0.061(hc)(T)		
Asbestos	1332-21-4			7x10 ⁶ fibers/L >10μm(h)					
Barium	7440-39-3			2,000(h)(T)					
Benz(a)anthracene	56-55-3			0.038(hc)			0.18(hc)		
Benzene	71-43-2			0.15(hc)			3.3(hc)		
Benzidine	92-87-5			0.000086(hc)			0.00020(hc)		
3,4-Benzofluoranthene (Benzo(b)fluoranthene)	205-99-2			0.038(hc)			0.18(hc)		
Benzo(k)fluoranthene	207-08-9			0.38(hc)			1.8(hc)		
Benzo(a)pyrene (BaP)	50-32-8			0.0038(hc)			0.018(hc)		
Beryllium	7440-41-7			6.0(h)(T)			42(h)(T)		
alpha-BHC (alpha-HCH)	319-84-6			0.0026(hc)			0.0049(hc)		
beta-BHC (beta-HCH)	319-85-7			0.0091(hc)			0.017(hc)		
gamma-BHC (gamma- HCH/Lindane)	58-89-9	0.95		0.98(h)	0.16		1.8(h)		
Bis(2-chloroethyl) ether	111-44-4			0.030(hc)			0.53(hc)		
Bis(2-chloroisopropyl) ether	108-60-1			1,400(h)			65,000(h)		
Bis(2-ethylhexyl) phthalate	117-81-7			1.2(hc)			2.2(hc)		
Bromodichloromethane (Dichlorobromomethane)	75-27-4			0.55(hc)			17(hc)		
Bromoform	75-25-2			4.3(hc)			140(hc)		
Butyl benzyl phthalate	85-68-7			150(h)			190(h)		
Cadmium	7440-43-9	(a)	(a)	3.4(h)(T)	40(d)(s)	8.8(d)(s)	16(h)(T)		
Carbon tetrachloride	56-23-5			0.33(hc)			2.3(hc)		
Chlordane	57-74-9	2.4	0.0043	0.00010(hc)	0.09	0.0040	0.00011(hc)		

	CAS	Fre	esh Water (FV	V2) Criteria	Salin	e Water (SE &	& SC) Criteria
Toxic Substance	Number	Aqu		Human Health	Aqua		Human Health
		Acute	Chronic		Acute	Chronic	
Chloride	16887-00-6	860,000	230,000	250,000(ol)			
Chlorine Produced Oxidants (CPO)	7782-50-5	19	11		13	7.5	
Chlorobenzene	108-90-7			210(h)			2,500(h)
Chloroform	67-66-3			68(h)			2,100(h)
2-Chloronaphthalene	91-58-7			1,000(h)			1,600(h)
2-Chlorophenol	95-57-8			81(h)			150(h)
Chlorpyrifos	2921-88-2	0.083	0.041		0.011	0.0056	
Chromium	7440-47-3			92(h)(T)			750(h)(T)
Chromium ⁺³	16065-83-1	(a)	(a)				
Chromium ⁺⁶	18540-29-9	15(d)(s)	10(d)(s)		1,100(d)(s)	50(d)(s)	
Chrysene	218-01-9			3.8(hc)			18(hc)
Copper*	7440-50-8	(a)	(a)	1,300(h)(T)	4.8(d)(s)	3.1(d)(s)	
Cyanide (Total)	57-12-5	22(fc)	5.2(fc)	140(h)	2.7(fc)	2.7(fc)	140(h)
4,4'-DDD (p,p'-TDE)	72-54-8			0.00031(hc)			0.00031(hc)
4,4'-DDE	72-55-9			0.00022(hc)			0.00022(hc)
4,4'-DDT	50-29-3	1.1	0.0010	0.00022(hc)	0.13	0.0010	0.00022(hc)
Demeton	8065-48-3		0.1			0.1	
Dibenz(a,h)anthracene	53-70-3			0.0038(hc)			0.018(hc)
Dibromochloromethane (Chlorodibromomethane)	124-48-1			0.40(hc)			13(hc)
Di-n-butyl phthalate	84-74-2			2,000(h)			4,500(h)
1,2-Dichlorobenzene	95-50-1			2,000(h)			6,200(h)
1,3-Dichlorobenzene	541-73-1			2,200(h)			8,300(h)
1,4-Dichlorobenzene	106-46-7			550(h)			2,200(h)
3,3'-Dichlorobenzidine	91-94-1			0.021(hc)			0.028(hc)
1,2-Dichloroethane	107-06-2			0.29(hc)			28(hc)
1,1-Dichloroethylene	75-35-4			4.7(h)			100(h)
trans-1,2-Dichloroethylene	156-60-5			590(h)			43,000(h)
2,4-Dichlorophenol	120-83-2			77(h)			290(h)
1,2-Dichloropropane	78-87-5			0.50(hc)			15(hc)
1,3-Dichloropropene (cis and trans)	542-75-6			0.34(hc)			21(hc)

	CAS	Fı	resh Water (F	W2) Criteria	Salir	ne Water (SE	& SC) Criteria
Toxic Substance	Number	Aqu		Human Health	Aqu		Human Health
Dieldrin	60-57-1	Acute 0.24	Chronic 0.056	0.000052(hc)	0.71	0.0019	0.000054(hc)
Diethyl phthalate	84-66-2	0.24	0.000	17,000(h)	0.71	0.0013	44,000(h)
2,4-Dimethyl phenol	105-67-9			380(h)			850(h)
4,6-Dinitro-o-cresol	534-52-1			• • • • • • • • • • • • • • • • • • • •			. ,
*				13(h)			280(h)
2,4-Dinitrophenol	51-28-5			69(h)			5,300(h)
2,4-Dinitrotoluene	121-14-2			0.11(hc)	-		3.4(hc)
1,2-Diphenylhydrazine	122-66-7			0.036(hc)			0.20(hc)
Endosulfans (alpha and beta)	115-29-7	0.22	0.056	62(h)	0.034	0.0087	89(h)
Endosulfan sulfate	1031-07-8			62(h)			89(h)
Endrin	72-20-8	0.086	0.036	0.059(h)	0.037	0.0023	0.060(h)
Endrin aldehyde	7421-93-4			0.059(h)			0.060(h)
Ethylbenzene	100-41-4			530(h)			2,100(h)
Fluoranthene	206-44-0			130(h)			140(h)
Fluorene	86-73-7			1,100(h)			5,300(h)
Guthion	86-50-0		0.01			0.01	
Heptachlor	76-44-8	0.52	0.0038	0.000079(hc)	0.053	0.0036	0.000079(hc)
Heptachlor epoxide	1024-57-3	0.52	0.0038	0.000039(hc)	0.053	0.0036	0.000039(hc)
Hexachlorobenzene	118-74-1			0.00028(hc)			0.00029(hc)
Hexachlorobutadiene	87-68-3			0.44(hc)			18(hc)
Hexachlorocyclopenta- diene	77-47-4			40(h)			1,100(h)
Hexachloroethane	67-72-1			1.4(hc)			3.3(hc)
Indeno(1,2,3-cd)pyrene	193-39-5			0.038(hc)			0.18(hc)
Isophorone	78-59-1			35(hc)			960(hc)
Lead	7439-92-1	38(d)(s)	5.4(d)(s)	5.0(h)(T)	210(d)(s)	24(d)(s)	
Malathion	121-75-5		0.1			0.1	
Manganese	7439-96-5						100(h)(T)
Mercury	7439-97-6	1.4(d)(s)	0.77(d)(s)	0.050(h)(T)	1.8(d)(s)	0.94(d)(s)	0.051(h)(T)
Methoxychlor	72-43-5		0.03	40(h)		0.03	
Methyl bromide (bromomethane)	74-83-9			47(h)			1,500(h)
Methyl t-butyl ether (MTBE)	1634-04-4			70(h)			

Federal and NJ State Primary and Secondary Drinking Water Standards as of June 2020

	•					•	
Volatile On	Volatile Organic Compounds	Inorganic (Inorganic Chemicals	Synthe	Synthetic Organic Compounds	Sec	Secondary Standards
Contaminants M	Maximum Contaminant	Contaminants	Maximum Contaminant	Contaminants	Maximum Contaminant	Physical Characteristics	Recommended Upper Limit or Optimum
Benzene	1*	Antimony	Q	Alachlor	2	Color	10 color units (standard cobalt scale)
Carbon Tetrachloride	2*	Arsenic	5 *	Aldicarb	+	PH	6.5 to 8.5 (optimum range)
1,2-Dichlorobenzene	600	Asbestos	7×10^6 fibers/l >10 μ m	Aldicarb Sulfone	+	Odor	3 Threshold odor number
1,3- Dichlorobenzene	600*	Barium	2,000	Aldicarb Sulfoxide	+	Taste	No objectionable taste
1,4- Dichlorobenzene	75	Beryllium	4	Atrazine	ω		
1,1-Dichloroethane	50*	Cadmium	5	Benzo[a]pyrene	0.2		
1,2-Dichloroethane	2*	Chromium	100	Carbofuran	40	Chemical Characteristics	Recommended Upper Limit [mg/l or ppm]
1,1-Dichloroethylene	2*	Copper	1,300**[AL]	Chlordane	0.5*		
cis- 1,2-Dichloroethylene	70	Cyanide	200	Dalapon	200	ABS/L.A.S.	0.5
trans- 1,2-Dichloroethylene	100	Fluoride	4,000	Dibromochloropropane [DBCP]	[DBCP] 0.2	Aluminum	0.2
1,2-Dichloropropane	σ	Lead	15**[AL]	Di[2-ethylhexyl]adipate	400	Chloride	250
Ethylbenzene	700	Mercury	2	Di[2-ethylhexyl]phthalate		Fluoride	2
Methyl tertiary Butyl Ether	70*	Nickel	+	Dinoseb	7	Hardness (as CaCO ₃)	250
Methylene Chloride	₩ *	Nitrate [as nitrogen]	10,000	Diquat	20	Iron	0.3
Monochlorobenzene	50*	Nitrite	1,000	Endothall	100	Manganese	0.05
Naphthalene	300*	[combined nitrate/nitrite]	10,000	Endrin	2	Silver	0.1
Styrene	100	Selenium	50	Ethylene dibromide [EDB]		Sodium	50
1, 1,2,2-Tetrachloroethane	1*	Thallium	2	Glyphosate	700	Sulfate	250
Tetrachloroethylene	1*			Heptachlor	0.4	Total Dissolved Solids (TDS)	500
Toluene	1,000	Disinfection	Disinfection Byproducts	Heptachlor Epoxide	0.2	Zinc	5
1,2,4-Trichlorobenzene	9*			Hexachlorobenzene	1		
1,1,1-Trichloroethane	30*	Contaminants Ma	Maximum Contaminant Levels	Hexachloroclyclopentadiene	iene 50	* N J MCI [A-280]	
1,1,2-Trichloroethane	3* *		[MCL] µg/L or ppb (as running	Lindane	0.2	** An [AL] action level is not an i	** An [AL] action level is not an MCL. It is a trigger point at which remedial action
Trichloroethylene	+ *		ailliuai avei ages pei gioup)	Methoxychlor	40	is to take place	-
Vinyl Chloride	2	Dichlorobromomethane	80 (TTHM)	Oxamyl	200	+ No MCL – Monitoring Required	
Xylenes [Total]	1,000*	Chlorodibromomethane	80 (TTHM)	PCBs	0.5	One milligram per liter [mg/l] = one part per m	one part per million = one cent in \$10,000 or one
		Bromofrom	80 (TTHM)	Pentachlorophenol		second in 12 days.	One microgram per liter film //II – one part per billion – one cent in ¢10 000 000
		2	8 /1	7	D17-A\		* 000 THE TOTAL TO

Radionuclide

-		
2	Dichlorobromomethane	80 (TTHM)
1,000*	Chlorodibromomethane	80 (TTHM)
	Bromofrom	80 (TTHM)
11Clides	Chloroform	80 (TTHM)
	Monochloroacetic acid	60 (HAA5)
Maximum Contaminant	Dichloroacetic acid	60 (HAA5)
Levels [MCL]	Trichloroacetic acid	60 (HAA5)
5 pCi/L	Bromoacetic acid	60 (HAA5)
15 pCi/L	Dibromoacetic acid	60 (HAA5)
4 mrem/year	Bromate	10
30 µg/L	Chlorite	1,000
ıtaminants	TTHM- Trihalomethanes HAA5- Haloacetic Acids	
the samples may exceed 0.3 NTU.	Chlorite (only for treatment plants using ozone) Chlorite (only for treatment plants using chlorine dioxide), requries daily/follow-up monitoring, not annual	nts using ozone) nts using chlorine dioxide), ng, not annual

Other Contamin

Beta/photon emitters Gross alpha particles

Combined radium 226/228

Contaminants

Turbidity No more than 5% of the sample NTU, nor any sample exceed 1 NTU.

approach to address fecal contamination that could enter into **Coliform bacteria** standards are based on an MCL for E. coli, and uses E. coli and total coliforms to initiate a "find and fix" subsequently take action to correct them perform assessments to identify sanitary defects and the distribution system. It requires public water systems to

For a detailed explanation of the Safe Drinking

Nater Program, refer to the Federal Safe Drinking

143] and the New Jersey Safe Drinking Water Water Act regulations [40 CFR Parts 141, 142,

> their regulatory framework follows that of Volatile Compounds due to their chemical makeup, however, Per- and polyfluoroalkyl substances (PFAS such as PFNA, PFOA & PFOS) are considered to be Synthetic Organic

2,4,5-TP [Silvex]

70 50 0.030*

3x10⁻⁵

1,2,3-Trichloropropane (1,2,3-TCP)

2,3,7,8—TCDD [Dioxin]

Simazine

Toxaphene

Picloram

Perfluorooctane sulfonic acid (PFOS) Perfluorooctanoic acid (PFOA) Perfluorononanoic acid (PFNA)

0.013*

One microgram per liter [µg//l] = one part per billion = one cent in \$10,000,000 or one second in 32 years.



New Jersey Department of Environmental Protection

Division of Water Supply and Geoscience

Bureau of Safe Drinking Water Trenton, New Jersey 08625 401 East State Street Mail Code 401-04Q P.O. Box 420

Tel. # (609) 292-5550

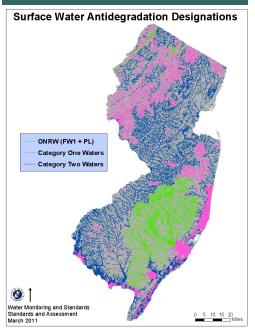
New Jersey Surface Water Quality Standards Antidegradation Designations

July 2017 Antideg. (Page 1 of 2)



Three Tiers of Antidegradation

- Outstanding National Resource Waters (ONRW)
 - -Freshwater 1 (FW1)
 - -Pinelands (PL)
- Category One (C1)-Exceptional Resource waters
- Category Two (C2)-All other waters





The **Surface Water Quality Standards** (SWQS) establish antidegradation policies for all surface waters of the State at N.J.A.C. 7:9B. The SWQS require that all existing and designated uses shall be maintained and protected for all surface waters of the State. Impaired waters must be restored to meet SWQS. Existing water quality shall be maintained. The three tiers of antidegradation designations are described below.

The most protective tier of antidegradation designation is **Outstanding National Resource Waters** (ONRW) which include surface waters classified as FW1 and PL. These waters are set aside for posterity because of their unique significance. The Department cannot approve any activity which might alter existing water quality in these waters.

The second tier of antidegradation designation is **Category One** (C1). C1 waters are designated through rulemaking for protection from measurable changes in water quality because of their Exceptional Ecological Significance, Exceptional Water Supply, Exceptional Recreation, and Exceptional Fisheries to protect and maintain their water quality, aesthetic value, and ecological integrity.

All waters not designated in the SWQS as ONRW (FW1 or PL) or C1 are designated as **Category Two** (C2). The same criteria apply in C1 and C2 waters. In all cases, existing and designated uses must be maintained and protected. Waterbodies that are generally not meeting criteria must be improved to meet water quality criteria.

The SWQS operate in conformance with the Federal Clean Water Act at 33 U.S.C. 1313(c) and the Federal Water Quality Standards Regulations at 40 C.F.R. 131. New Jersey's statutory authority is granted by the Water Pollution Control Act at N.J.S.A. 58:10A and the Water Quality Planning Act at N.J.S.A. 58:11A.





New Jersey Surface Water Quality Standards Antidegradation Designations: Category One

Antideg. (Page 2 of 2)



Category One (C1) designation provides additional protection to waterbodies that help prevent water quality degradation and discourage development where it would impair or destroy natural resources and water quality. The maintenance of water quality is important to all residents, particularly to the many communities that depend upon surface waters for drinking water supplies, recreation, fishing, and shellfish harvesting.



Category One waters were originally designated in 1985 based on parks, wildlife management areas, and trout production waters. After 1985, additional streams upgraded to FW2-trout production were routinely designated as C1. In 2002, the Department began an intensive effort to identify additional waters that warranted enhanced protections afforded by this designation. The Department adopted new definitions of Exceptional Ecological Significance, Exceptional Fisheries Resource(s), and Exceptional Water Supply Significance to clarify data requirements for a waterbody to be designated as C1.



Under the definition of Exceptional Ecological Significance, the Department considers those waterbodies supporting certain aquatic-dependent T&E (bog turtles and mussels) or exceptional aquatic community can qualify for C1. Waterbodies that support unimpaired benthic macroinvertebrates and indicate exceptional value for only two of the four data sources: habitat, physical/chemical water quality data, fish assemblage, and low impervious surface can qualify under exceptional aquatic community.



Under the definition of Exceptional Fisheries Resource(s), trout production waters classified as FW2-TP and approved shellfish harvesting waters can qualify for C1.

Under the definition of Exceptional Water Supply Significance, waterbodies that are part of the water supply system that serves a population greater than 100,000, including any reservoirs and streams that directly flow into those reservoirs can qualify for C1. See http://www.state.nj.us/dep/wms/bears/c1waters.htm.



Implementation of Category One Waters

- New Jersey Pollutant Discharge Elimination System (NJPDES) Rules at N.J.A.C. 7:14A: New or expanded wastewater discharges must maintain the existing water quality of the receiving stream. If the discharge is located above a C1 segment the applicant must meet "no measurable change" at the C1 boundary. See www.nj.gov/dep/dwq/.
- Flood Hazard Area Control Act Rules at N.J.A.C. 7:13: 300 foot riparian zones are imposed through Flood Hazard Area Control Act rule permits to all C1 waters and their upstream tributaries within the same sub-watershed or HUC 14. See www.nj.gov/dep/landuse/.



For additional information, please see http://www.state.nj.us/dep/wms/bears/c1waters.htm.

Parameter Units CAS No. Ammonia mg TAN/L 7664-41-7

Acute	Chronic	Classification
2.35	0.54	FW2-TM,TP
5.80	0.54	FW2-NT
	0.54	FW2

FW2-TM,TP

FW2-TM,TP рН Temp (C) 2.35 0.275 0.26 1.00 0.06 8.45 39.00 2.09 1.00 17.62 8.45 FW2-TM,TP 2.35

14.5 Used field measured pH and temp from October 16, 2024 sampling event

Acute 2.35

FW2-TM,TP		рН	
	2.58		8.45
	0.72	0.72	
	0.01	0.01	
	1.00		
	0.06		8.45
	1.62	0.09	
	1.00		
	17.62		8.45
	23.12		
	36.47	36.47	
FW2-TM,TP		2.58	

14.5

14.5

14.5

14.5

Acute 2.58

FW2-NT	FW2-NT	pН	
	5.80		8.45
		. =-	
	0.72	0.72	
	0.01	0.01	
	1.00		
	0.06		8.45
	1.62	0.09	
	1.00		
	17.62		8.45
	51.93		
	81.93	81.93	
Acute 5.80	FW2-NT	5.80	

FW2		p⊦	ł	
	0.54		8.45	14.5
	0.89	0.89		
	0.03	0.02		
	1.00			
	0.17		8.45	
	1.20	0.18		
	1.00			
	5.78		8.45	
	2.42			
	2.13			
	3.03	3.03		14.5
FW2		0.54		

Chronic

Chronic 0.54 3. Ammonia, total 7664-41-7 (mg TAN/L)

FW2-TP,

i.
$$\text{FW2-TM} \\ \text{MIN} \left(\frac{\left(\frac{0.275}{1+10^{7.204-pH}} + \frac{39.0}{|1+10^{pH-7.204}} \right),}{\left(0.7249 \times \left(\frac{0.0114}{1+10^{7.204-pH}} + \frac{1.6181}{1+10^{pH-7.204}} \right) \times \left(23.12 \times 10^{0.036 \times (20-T)} \right) \right)} \right) \text{ (acute)}$$

$$ii. \\ 0.7249 \times \left(\frac{0.0114}{1+10^{7.204-pH}} + \frac{1.6181}{1+10^{pH-7.204}}\right) \times MIN \left(51.93, 23.12 \times 10^{0.036 \times (20-T)}\right) \text{ (acute)}$$

iii. All FW2
$$0.8876 \times \left(\frac{0.0278}{1+10^{7.688-pH}} + \frac{1.1994}{1+10^{pH-7.688}}\right) \times \left(2.126 \times 10^{0.028 \times \left(20-MAX(\psi,7)\right)}\right) \text{ (chronic)}$$

Parameter Units CAS No. Ammonia mg TAN/L 7664-41-7

Acute	Chronic	Classification
0.98	0.25	FW2-TM,TP
2.21	0.25	FW2-NT
	0.25	FW2

F۷	۷2۰	-TN	1 .1	ГΡ
	v ~		v.,	

FW2-TM,TP		рН		Temp (C)
	2.14		8.5	25
	0.275	0.26		
	1.00	0.20		
	0.05		8.5	
	39.00	1.88		
	1.00			
	19.77		8.5	
FW2-TM,TP		2.14		

Acute 2.14

FW2-TM,TP		рН		
	0.98		8.5	25
	0.72	0.72		
	0.01	0.01		
	1.00			
	0.05		8.5	
	1.62	0.08		
	1.00			
	19.77		8.5	
	23.12			
	15.28	15.28		25
FW2-TM,TP		0.98		

Acute 0.98

FW2-NT	

FW2-NT		рН		
	2.21		8.5	25
	0.72	0.72		
	0.01	0.01		
	1.00			
	0.05		8.5	
	1.62	0.08		
	1.00			
	19.77		8.5	
	51.93			
	34.31	34.31		25
FW2-NT		2.21		

Acute 2.21

FW2 pH

Chronic

[0.25		8.5	25
	0.89	0.89		
	0.03	0.02		
	1.00			
	0.15		8.5	
	1.20	0.16		
	1.00			
	6.49		8.5	
	2.13			
	1.54	1.54		25
FW2		0.25		
		-		

Chronic 0.25

Source: NJAC 7:9B Dec-23

3. Ammonia, total 7664-41-7 (mg TAN/L) FW2-TP, FW2-TM

i.
$$MIN \begin{pmatrix} \frac{0.275}{1+10^{7.204-pH}} + \frac{39.0}{1+10^{pH-7.204}}, \\ \frac{0.0114}{1+10^{7.204-pH}} + \frac{1.6181}{1+10^{pH-7.204}} \times (23.12 \times 10^{0.036 \times (20-T)}) \end{pmatrix}$$
 (acute)

iii. All FW2
$$0.8876 \times \left(\frac{0.0278}{1+10^{7.688-pH}} + \frac{1.1994}{1+10^{pH-7.688}}\right) \times \left(2.126 \times 10^{0.028 \times \left(20-MAX\left(p,7\right)\right)}\right) \text{ (chronic)}$$

Aquatic Life Ambient Water Quality Criteria for Ammonia - Freshwater (2013)

Summary

EPA has published national recommended ambient water quality criteria for the protection of aquatic life from the toxic effects of ammonia, a constituent of nitrogen pollution. These recommended criteria will help States, Territories, and authorized Tribes update their water quality standards with concentration levels for ammonia in surface waters at or below which aquatic organisms will be protected, if not exceeded more frequently than once every three years on average. Acute and chronic criteria were developed to protect organisms from both immediate effects, such as mortality, and longer-term effects on reproduction, growth and survival, respectively.

EPA's final Aquatic Life Ambient Water Quality Criteria for Ammonia – Freshwater (2013) incorporate scientific views received on EPA's 2009 draft updated ammonia criteria and supersede EPA's previously recommended 1999 ammonia criteria.

What are national recommended aquatic life ambient water quality criteria?

Ambient water quality criteria for the protection of aquatic life are numeric concentrations of pollutants, with specific associated duration and frequency information, in surface waters that are protective of aquatic life designated uses. Under Clean Water Act section 304(a), EPA is required to develop and publish water quality criteria that reflect the latest scientific knowledge. Water quality criteria are based solely on data and scientific judgments about the relationship between pollutant concentrations and potential environmental and human health effects. EPA's recommended water quality criteria are not rules, nor do they automatically become part of a state's water quality standards. States must adopt into their standards water quality criteria that protect the designated uses of the water bodies

within their area. These can include scientifically defensible site-specific criteria that are different from EPA's national recommended criteria, as long as the site-specific criteria are protective of the designated use. Water quality criteria are not effective under the Clean Water Act until they have been adopted into a state's water quality standards and approved by EPA.

What is ammonia?

Ammonia is one of several forms of nitrogen that exist in aquatic environments. Unlike other forms of nitrogen, which can cause nutrient over-enrichment of a water body at elevated concentrations and indirect effects on aquatic life, ammonia causes direct toxic effects on aquatic life.

Ammonia is produced for commercial fertilizers and other industrial applications. Natural sources of ammonia include the decomposition or breakdown of organic waste matter, gas exchange with the atmosphere, forest fires, animal and human waste, and nitrogen fixation processes.

How does ammonia enter surface waters?

Ammonia can enter the aquatic environment via direct means such as municipal effluent discharges and the excretion of nitrogenous wastes from animals, and indirect means such as nitrogen fixation, air deposition, and runoff from agricultural lands.

How does ammonia affect aquatic life?

When ammonia is present in water at high enough levels, it is difficult for aquatic organisms to sufficiently excrete the toxicant, leading to toxic buildup in internal tissues and blood, and potentially death. Environmental factors, such as pH and temperature, can affect ammonia toxicity to aquatic animals.

What is the history of EPA's development of ammonia criteria?

EPA first published ammonia criteria for the protection of aquatic life in 1976. The criteria were then updated in 1985 and 1999 to reflect scientific information available at that time. The 1999 recommended aquatic life criteria for ammonia were based on the most sensitive endpoints known at the time: the acute criterion was based on salmonid fish toxicity information, and the chronic criterion was based on bluegill sunfish early life stage toxicity.

In 2003, EPA became aware of new toxicity studies indicating the relative sensitivity of freshwater mussels to ammonia and began to update the 1999 criteria to reflect this new information. In 2009, following external peer review, EPA published draft recommended ammonia criteria, for waters with and without mussels. Since the publication of the draft 2009 ammonia criteria, additional toxicity testing has validated information on the effects of ammonia on sensitive freshwater gill-breathing snail species. In April 2013, EPA finalized the updated ammonia criteria that are applicable nationally, taking into account the latest toxicity information for freshwater species, including unionid mussels and gill-breathing snails. The 2013 criteria incorporate scientific views received on the draft (2009) ammonia criteria and supersede EPA's previously recommended 1999 criteria.

What are the 2013 recommended water quality criteria for ammonia?

EPA recommends an acute criterion magnitude of 17 mg Total Ammonia Nitrogen (TAN) per liter at pH 7 and 20°C for a one-hour average duration, not to be exceeded more than once every three years on average. EPA recommends a chronic criterion magnitude of 1.9 mg TAN/L at pH 7 and 20°C for a 30-day average duration, not to be exceeded more than once every three years on average. In addition, the highest four-day average within a 30-day period should not exceed 2.5 times the chronic criterion magnitude (e.g. 1.9 mg TAN/L x 2.5 = 4.8 mg TAN/L at pH 7 and 20°C) more than once in three years on average.

How do the 2013 criteria compare to the previously recommended 1999 criteria and the draft 2009 criteria?

The 2013 ammonia criteria recommendations take into account the latest freshwater toxicity information for ammonia, including toxicity studies for sensitive unionid mussels and gillbreathing snails. These new criteria are based on robust toxicity data available for 69 genera (acute) and 16 genera (chronic). The updated criteria magnitudes are more stringent than the previously recommended 1999 criteria magnitudes (see Table 1). The duration components of the 1999, 2009 and 2013 criteria remain the same - a one-hour average duration for the acute criterion and 30-day average duration for the chronic criterion. The frequency component for the acute and chronic criteria remains once in three years on average.

Table 1. Comparison of past and current EPA-recommended aquatic life water quality criteria magnitudes for ammonia. Criteria magnitudes are expressed as total ammonia nitrogen (mg TAN/L) at pH 7 and 20°C.

Criterion Duration	1999 Criteria	2009 Draft Updated Criteria	2013 Final Updated Criteria
Acute (1-hour average)	24	19	17
Chronic (30-day rolling average)	4.5*	0.91*	1.9*

^{*}Not to exceed 2.5 times the criterion continuous concentration as a 4-day average within a 30-day period.

Criteria frequency: Not to be exceeded more than once in three years on average.

Additional EPA Resources

EPA has developed three supporting documents to aid states considering adoption of the 2013 recommended ammonia criteria.

Flexibilities for States Applying EPA's Ammonia Criteria Recommendations provides an overview of a number of implementation approaches available for state consideration, including the recalculation procedure for site-specific criteria derivation, variances, revisions to designated uses, dilution allowances, and compliance schedules. The document describes how each of these flexibilities fits within a state's water quality standards adoption and implementation processes.

EPA has also developed a *Revised Deletion Process for the Site-Specific Recalculation Procedure for Aquatic Life Criteria* that describes a recalculation procedure and includes a spreadsheet that may be used to derive site-specific water quality criteria for the protection of aquatic life in order to best reflect the organisms that reside at a specific site.

A third document, which EPA expects to publish in 2013, Technical Support Document for Conducting and Reviewing Freshwater Mussel Studies for the Development of Site-specific Water Quality Criteria for Ammonia, will help states determine if sensitive freshwater mussels are present in their waters. Commonly-used mussel sampling methods will be described and an overview will be provided of various study approaches, considerations, and limitations, including real-life examples.

How to View the Criteria Document and Supporting Information

EPA has established an official public docket for this action under Docket ID No. EPA-HQ-OW-2009-0921, accessed at www.regulations.gov. You may also download the document and supporting information from http://water.epa.gov/scitech/swguidance/standards/criteria/aqlife/ammonia/index.cfm.

For More Information

Contact Lisa Huff by telephone at (202) 566-0787, by email at huff.lisa@epa.gov, or by mail at U.S. EPA, MC: 4304T, 1200 Pennsylvania Ave., N.W., Washington, D.C. 20460.

Understanding Water Quality Parameters to Better Manage Your Pond

Rossana Sallenave¹

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INTRODUCTION

Successful pond management requires an understanding of the role of nutrients and other water quality parameters, as well as regular monitoring of environmental conditions within the pond's ecosystem. Water quality is often overlooked



in pond management, and poor water quality can lead to common problems, such as excessive algal blooms, overgrowth of plants, noxious smells, or dead and dying fish. In order to prevent these problems, an understanding of basic water chemistry and other physical parameters is necessary. This publication describes the most important water quality factors that influence the health of ponds. Some factors that are particularly important in recreational fish ponds to ensure fish health and pond productivity are also discussed. A basic understanding of how these factors interact with one another will help pond owners maintain good water quality and a healthy pond ecosystem. Many companies produce kits and other materials to monitor water quality on your own, or you can send water samples to commercial laboratories for analysis.

DISSOLVED OXYGEN

Dissolved oxygen (DO) is probably the single most important water quality factor for pond owners. Oxygen is needed by fish and other aquatic organisms, and levels of DO will determine the ability of ponds and other water bodies to support aquatic life. Oxygen dissolves in water at very low concentrations measured in parts per million (ppm, which can be used interchangeably with milligrams per liter [mg/L]). Ponds will rarely have more than 10 ppm DO. Most oxygen in water is produced by algae and green plants through photosynthesis, the process whereby green plants use solar energy to convert water and carbon dioxide (CO2) to oxygen and carbohydrates. Oxygen is also naturally incorporated into water from the atmosphere through surface diffusion and turbulence caused by wind.

Daily fluctuations and seasonal changes in DO

Dissolved oxygen levels can vary dramatically in a 24-hour period. During the day, DO concentrations generated by photosynthesis will increase. During the

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night, DO levels will decline as oxygen is removed from water through respiration, the process whereby plants and animals consume oxygen and release carbon dioxide as they convert organic material to energy. For this reason, DO levels are typically highest at dusk and lowest just before dawn. There is also a strong relationship between temperature and DO: the warmer the water, the less oxygen it can hold. For example, water at 52°F (11°C) can hold 40% more oxygen than water at 80°F (27°C). Warm water increases the metabolism of fish and therefore increases their consumption of oxygen. Bacteria also consume oxygen as they decompose organic matter. Therefore, during the summer months, DO levels will be lower because of increased oxygen demands of fish, warmer water that holds less oxygen, and increased bacterial decomposition of dead plant and algal material toward the end of the growing season.

Effects of low DO

Oxygen depletion occurs when the demands for oxygen are greater than what is being produced. Oxygen depletion can occur for different reasons. Situations typically associated with oxygen depletion are

- Hot, cloudy, and still (windless) days;
- Pond stratification followed by turnovers (the mixing of stratified layers, which develop during the summer in ponds 8 ft deep or greater);
- After a sudden algal bloom die-off (from natural causes or after a chemical application); and
- Organic waste decomposition (oxygen depletion will occur in the presence of excessive organic matter from waste products, such as uneaten feed).

Whenever DO levels fall below 3 to 4 ppm, oxygen stress will occur. Lack of adequate dissolved oxygen is the leading cause of fish kills. Normal oxygen content in a healthy pond will range from 5 to 10 ppm. Warmwater fish (e.g., bass, bluegill, catfish) require about 5 ppm and coldwater fish (e.g., trout, salmon) require about 6.5 ppm to maintain good health. Dissolved oxygen levels of less than 3 ppm will kill warmwater fish and levels less than 5 ppm will kill coldwater fish. Fish exposed to low, nonlethal levels of DO over prolonged periods will be chronically stressed, stop eating, and be more susceptible to disease. Low oxygen concentrations also increase the activity of anaerobic bacteria, which create methane and hydrogen sulfide gases during anaerobic decomposition. Ponds with oxygen-poor bottoms and accumulated organic matter can release these gases when the bottom sediment is disturbed. Hydrogen sulfide has a rotten egg smell and is very toxic to fish.

Preventing low DO conditions

To help maintain safe DO levels in ponds, particularly in deeper ponds in which fish are intensively cultured, mechanical aeration is often needed. Aerators help keep pond water mixed so that layering is minimized and the surface water is well-oxygenated. However, aeration should only be thought of as one of many management tools to help maintain healthy oxygen levels. External nutrient loading is still the critical issue that must be addressed because excessive nutrients can lead to an overabundance of aquatic weeds and algae, which can result in oxygen depletions when they die and decompose.

NUTRIENTS (PHOSPHORUS AND NITROGEN)

It is important to understand the sources and basic pathways of nutrients because there is a direct correlation between available nutrients and populations of algae and aquatic weeds. The most important nutrients in aquatic systems are phosphorus (P) and nitrogen (N) in the forms of phosphates (PO₃) and nitrates (NO₃). These nutrients are critical to the growth of plants and animals in aquatic systems. Phosphorus has been identified as the limiting factor for algal growth in most lakes and, as such, is the largest contributor to aquatic plant growth. One gram of phosphorus will produce 100 grams of algal biomass. Excessive amounts of nutrients will lead to over-fertilization, or eutrophic conditions, which can result in an over-abundance of aquatic plants and algal blooms. When the excess plants and/or algae die, they decompose, which leads to a depletion of oxygen that can affect water clarity and smell and can lead to fish kills.

Sources of nutrients

The main sources of nutrients in ponds are bottom silt, dead vegetation, landscape debris, runoff from the surrounding area, poorly functioning septic systems, and wastes from livestock and waterfowl. As aquatic plants and algae grow and die, they sink to the bottom of the pond and provide a source of nutrients for future aquatic growth, a phenomenon known as nutrient cycling. This, along with landscape debris such as grass clippings, leaves, and pine needles, contributes nutrients to ponds, and these nutrients must be managed to prevent eutrophic conditions from developing. Runoff from fertilized fields and lawns in immediate surrounding areas as well as roads, farms, and outlying areas can also be major sources of nutrient enrichment.

Dissolved and particulate phosphorus

Phosphorus in water comes in two forms: dissolved and particulate. Dissolved phosphorus enters the aquatic environment from fertilizers, crop residues, or human or animal wastes, and is the form that is readily available to aquatic plants and algae. Particulate phosphorus is bound to soil particles and minerals that contain aluminum, iron, or calcium, as well as to organic matter, and enters aquatic systems primarily through soil erosion and surface runoff. While it may not be as readily available to aquatic plants, particulate phosphorus can accumulate in sediments and can be a source of slow release of phosphorus into the water for years.

NITROGENOUS WASTE (AMMONIA)

Ammonia is another compound that can affect the health and performance of your pond.

Sources of ammonia

Ammonia is a form of nitrogen found in organic materials and many fertilizers. It is the first form of nitrogen released when organic matter decays and is the main nitrogenous waste excreted by most fish and freshwater invertebrates. It is very unlikely that ammonia levels in your pond will reach levels that are lethal to fish. However, under conditions where fish are cultured intensively and fed proteinrich diets, they can produce high concentrations of ammonia, and fish may be exposed to sub-lethal levels (greater than 0.02 ppm) for extended periods of time. This can lead to reduced growth and increased susceptibility to disease.

Forms of ammonia

Ammonia can exist in two forms: un-ionized ammonia (NH $_3$) and ionized ammonia, also known as ammonium ion (NH $_4$ ⁺). The ratio of un-ionized to ionized ammonia depends on pH and water temperature. Un-ionized ammonia (NH $_3$) is extremely toxic to fish and is the predominant form of ammonia when pH is high. Ionized ammonia (NH $_4$ ⁺) is nontoxic except at extremely high levels and is the predominant form in water when pH is low. As a general rule, less than 10% of the ammonia will be the toxic un-ionized form when water pH values are lower than 8; however, this proportion increases greatly as pH increases. Water temperature will also affect the equilibrium between NH $_3$ and NH $_4$ ⁺. At any given pH, more toxic NH $_3$ will be present in warmer water than in cooler water.

Ammonia removal and transformation processes

There are two processes that remove or transform ammonia released into the water. The first is uptake of ammonia by plants and algae, which readily use the nitrogen in ammonia as a nutrient for growth. For this reason, ammonia levels are usually low in ponds during summer months when algae are most productive, but can increase rapidly after the crash of an algal bloom. This is also one of the reasons why ammonia levels will tend to be higher in ponds during the winter months when algal production is low.

The second process, which transforms ammonia, is a step in the nitrogen cycle known as nitrification, the biological conversion of ammonia and ammonium to nitrate nitrogen. Nitrification is a two-step process. First, *Nitrosomonas* bacteria convert ammonia and ammonium to nitrite (NO₂). Nitrite, which is also highly toxic to fish, is then converted to nitrate (NO₃) by *Nitrobacter* bacteria. These reactions are usually coupled, and nitrite is rapidly converted to nitrate, so nitrite levels are usually low. The rate of nitrification is affected by water temperature. Maximum rates of nitrification occur at water temperatures between 86 and 95°F (30–35°C). At temperatures of 104°F (40°C) and higher, nitrification rates fall to near zero. At temperatures below 68°F (20°C), nitrification proceeds at

a slower rate, but will continue at temperatures of 50°F (10°C) or less. For this reason, ammonia levels tend to be higher in fall and early spring before nitrification rates have increased as a result of increasing temperatures.

pН

The term pH refers to the concentration of hydrogen ions, and is a measure of whether a substance is an acid, a base, or neutral. The "p" in pH stands for "power" and the "H" for hydrogen ions. The scale of pH values ranges from 0 to 14; 7 represents neutral conditions, values less than 7 indicate more acidic conditions, and values above 7 indicate more alkaline or basic conditions.

Daily fluctuations in pH

The pH of freshwater ponds can fluctuate considerably both daily and seasonally; the magnitude of this fluctuation will depend on how well-buffered the freshwater system is. These fluctuations are due to photosynthesis and respiration by plants and animals, which results in the highest pH typically occurring at dusk and the lowest at dawn. This is because during the night respiration increases concentrations of carbon, which interacts with water to produce carbonic acid (H₂CO₃), lowering the pH. During the day, carbon dioxide concentrations decrease because of photosynthesis, driving pH values up.

How pH affects animals and other water quality variables

Optimum pH for fish growth and health is between 6 and 9. If pH is outside this range, fish growth will be reduced. Mortalities will occur when pH values are less than 4.5 or greater than 10. In addition to the direct effects pH can have on fish and other aquatic animals, pH interacts with other water quality variables such as ammonia, hydrogen sulfide, and dissolved metals, affecting their aqueous equilibria and toxicity as well. For example, as previously mentioned, high pH increases the toxicity of ammonia to fish, whereas low pH increases toxicity of aluminum and copper. Hydrogen sulfide (H2S) is a toxic, colorless gas that can form in pond sediments when bacteria feed on organic debris in areas that are low or depleted of oxygen, giving off a rotten egg smell when the sediments are stirred up. When dissolved in water, H₂S can undergo two chemical steps, which go back and forth depending on the pH. At pH less than 6, most of the hydrogen sulfide will be in the toxic H₂S form, whereas at higher pH (8–12), most of the hydrogen sulfide will be in the less toxic HS⁻ form.

ALKALINITY

Alkalinity refers to the water's buffering capacity, or its ability to withstand changes in pH. It is a measure of the total concentration of bases in pond water, including carbonates, bicarbonates, hydroxides, phosphates, and borates, and is expressed in ppm calcium carbonate. All these

bases react with and neutralize acids, which in turn buffers changes in pH. The pH of well-buffered water will normally fluctuate between 6.5 and 9. Carbonates and bicarbonates are the most common and important components of alkalinity. In an established pond, the ideal alkalinity measurement should be around 100 ppm, but readings from 50 to 200 ppm are acceptable. If the alkalinity is low, even a small amount of acid can cause a large change in pH. Alkalinity values greater than 300 ppm will not adversely affect fish, but such high values will render some commonly used chemicals, such as copper sulfate, ineffective. Alkalinity can be increased by adding agricultural limestone [CaCO₃ and CaMg(CO₃)₂] to ponds.

HARDNESS

Hardness is a measure of divalent salts, or positively charged ions, particularly calcium (Ca²⁺) and magnesium (Mg²⁺), in water. Total hardness is the sum of the concentrations of Ca^{2+} and Mg^{2+} , expressed in ppm calcium carbonate. Calcium carbonate hardness is a general term that indicates the total amount of divalent salts present, but it does not specify which salts are causing water hardness. Hardness and alkalinity are often confused because both are expressed using the same term (ppm calcium carbonate), and sometimes both parameters have similar values in a given water body. However, alkalinity measures negative ions (carbonate and bicarbonate) and hardness measures positive ions (calcium and magnesium), and sometimes these values can differ greatly. If limestone (calcium carbonate) is the cause of hardness and alkalinity, these values will be similar or identical. However, if sodium bicarbonate (NaHCO₂) is responsible for high alkalinity, it is possible for water to have high alkalinity and low hardness and calcium. Calcium and magnesium are essential to fish for biological processes such as bone and scale formation. If your pond is used to culture fish, water hardness should be above 50 ppm and can be adjusted by adding agricultural limestone.

SUMMARY

A basic understanding of the chemical components of aquatic ecosystems is important to successfully manage any pond or lake. The interaction between temperature, nutrients, and oxygen plays a critical role in many common problems encountered by pond owners, such as excessive algal growth, oxygen depletion, and fish kills. A healthy pond ecosystem is easier to achieve by understanding this interaction and managing excessive nutrient loading to the pond system. Other parameters, such as pH, alkalinity, and

hardness, can also affect fish growth and survival and can influence toxicity of other compounds, such as ammonia and metals. Water quality testing should be considered if your pond is to be used for intensive fish culture. A variety of methods are available to monitor water quality. Several companies produce kits and materials to monitor water quality, or water samples can be sent off to commercial laboratories for testing.

REFERENCES

Buttner, J.K., R.W. Soderberg, and D.E. Terlizzi. 1993. An introduction to water chemistry in freshwater aquaculture
 [Publication No. 170-1993]. Dartmouth: University of Massachusetts, Northeastern Regional Aquaculture Center.

Dietrich, D., and C. Schlatter. 1989. Aluminum toxicity to rainbow trout at low pH. *Aquatic Toxicology*, 15, 197–212.

Laurén, D.J., and D.G. McDonald. 1986. Influence of water hardness, pH, and alkalinity on the mechanisms of Cu toxicity in the juvenile rainbow trout, *Salmo gairdneri. Canadian Journal of Fisheries and Aquatic Sciences*, 43, 1,488–1,496.

Hargreaves, J.A., and C.S. Tucker. 2004. *Managing ammonia in fish ponds* [Publication No. 4603]. Stoneville, MS: Southern Regional Aquaculture Center.

Lewis, G.W. 1993. Oxygen depletion in ponds [Leaflet 233]. Athens: University of Georgia Cooperative Extension Service.

Wetzel, R.G. 2001. *Limnology: Lake and river ecosystems*, 3rd ed. San Diego, CA: Academic Press.

Wurts, W.A. 1993. Dealing with oxygen depletion in ponds. *World Aquaculture*, 24, 108–109.

Wurts, W.A. 2002. Alkalinity and hardness in production ponds. *World Aquaculture*, 33, 16–17.

Wurts, W.A., and R.M. Durborow. 1992. *Interactions of* pH, carbon dioxide, alkalinity and hardness in fish ponds [Publication No. 464]. Stoneville, MS: Southern Regional Aquaculture Center.



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Environmental Assessment and Risk Analysis Element



Research Project Summary



May, 2003

Ambient Levels of Metals in New Jersey Soils

Paul F. Sanders, Ph.D.

Abstract

Between 1996 and 2001, three studies were conducted to determine the ambient levels of extractable metals in New Jersey soils. These studies were conducted to gather information to support the development of soil cleanup criteria, which cannot be set below ambient levels. A total of 248 soil samples were taken from the urban Piedmont region, the urban Coastal Plain region, and rural regions of the Valley and Ridge, Highlands, and Coastal Plain provinces. Local or point sources of contamination were avoided by the use of Geographic Information System databases and by following sample location guidelines in the field. Surface soil samples (0-6") were analyzed for acid-extractable Target Analyte List (TAL) metals using USEPA SW-846 methods that are normally used to conduct initial investigations at hazardous waste sites. With one exception, median and 90th percentile concentrations of all metals were below current soil cleanup criteria. The exception was the 90th percentile arsenic concentration from the urban Piedmont study, which slightly exceeded the arsenic criterion. Otherwise, only certain individual samples contained metal concentrations above current criteria. A single rural soil sample yielded a beryllium concentration slightly above the corresponding criterion. For the urban Coastal Plain study, three of the 91 samples contained levels of arsenic above the current criterion. The urban Piedmont study yielded eight samples out of 67 where levels of arsenic or lead exceeded the criteria.

Introduction

Current New Jersey law requires that the NJDEP determine background levels of contaminants in soils and that "Remediation [of contaminated areas] shall not be required below regional natural background levels for any particular contaminant" [N.J.S.A. 58:10B-12(g)(4)]. "Natural background level" is further defined as "...the concentration of a contaminant consistently present in the environment of the region of the site and which has not been influenced by localized human activities...." Therefore, naturally occurring constituents in soil and those resulting from regional deposition are included, but not those from point contamination sources. The concentrations of contaminants included in this definition are referred to as "ambient concentrations." To support the above directive, three studies were conducted to determine ambient levels of metals in several regions of New Jersey (BEM Systems, Inc., 1997, 1998, 2002). The first two studies investigated metal concentrations in the urban Piedmont and urban Coastal Plain regions of New Jersey. These two areas contain a majority of the hazardous waste sites in the state. Furthermore, the high population density and significant industrial activity in these

regions yields an upper estimate of ambient metal concentrations, due to a larger impact of regional atmospheric deposition. The third study focused on rural areas of the Ridge and Valley, Highlands and Coastal Plain provinces, and provided an indication of metals concentrations in areas less impacted by atmospheric deposition.

Methods

A total of 248 soil samples were taken in the three studies: 67 from the urban Piedmont region, 91 from the urban Coastal Plain region, and 90 from the three rural provinces (Figure. 1). The rural soil samples were distributed among the predominant soil types in each of the regions on a rural acreage basis. The sample locations for the urban studies were generally distributed among municipalities classified as "urban" by population density criteria. For the urban Piedmont studies, the criterion usually applied was a population density of 7,500 or more people per square mile, as per the State Development and Redevelopment Plan. For the urban Coastal Plain study, the population density criteria was reduced to 4,000 or more people per square mile because a

higher population density would have resulted in only eight municipalities being sampled. In addition, municipalities were added to both urban studies that did not meet the density criteria because they contained high population density regions or substantial industrial activity. For the urban studies, samples were located in public parks because of ease of access, a likelihood that these soils have been undisturbed for some time, and because they usually met a specified 50% open space (lawn) requirement. This latter requirement was specified because forest cover or other obstructions could reduce the impact of atmospheric deposition. Sports fields or other manicured areas were not acceptable locations because of turf maintenance issues often associated with them, including chemical applications. For the rural study, sampling in forested areas was usually unavoidable since this is the natural vegetation cover of much of New Jersey, and it was desired to avoid areas impacted by human activity. For all studies, sample locations were also required to be specified minimum distances from known hazardous waste sites, roadways, and railroads.

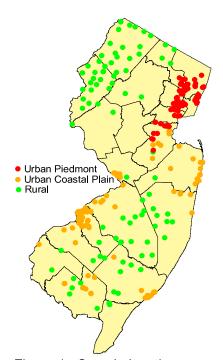


Figure 1. Sample locations

Soil samples were taken from a depth of 0-6 inches after removing surface litter. Samples were analyzed for acid-extractable Target Analyte List (TAL) metals using New Jersey certified laboratories. The methods used were those used to conduct site investigations at hazardous waste sites, specifically

the U.S. Environmental Protection Agency Office of Solid Waste SW-846 methods. The extraction method used was Method 3050. Many metals were analyzed using inductively coupled plasma – atomic emission spectrometry (Method 6010). For certain metals, lower detection limits were desired because of low ambient concentrations or low soil ingestion criteria based on toxicity concerns. Atomic absorption furnace methods were used for arsenic, lead, selenium and thallium (Methods 7060, 7421, 7740 and 7841, respectively). Mercury was analyzed via cold vapor atomic adsorption (Method 7471).

Results

Acid-extractable median and 90th percentile concentrations, and the corresponding method detection limits for the twenty-three target analyte metals in 248 surface soil samples from the three studies were calculated (Table 1). The rural study results are separately presented for each of the three physiographic provinces.

Aluminum, iron, calcium, sodium, potassium, magnesium and manganese are abundant in soils and were frequently measured at several hundred to several thousand mg/kg. (Sodium concentrations are lower because it is largely leached from soils in humid climates.) Barium, chromium, vanadium and zinc are also relatively common in soils and were frequently measured at concentrations between 10 and 100 mg/kg. Zinc showed some indication of anthropogenic contribution in urban areas in that a few samples yielded concentrations in the 150-350 mg/kg range. (Naturally occurring total zinc concentrations in soil do not commonly exceed 100 mg/kg [Kabata-Pendias and Pendias, 1984].)

Nickel, copper, cobalt and lead are somewhat less common. The first three of these metals were generally measured at levels less than 50 mg/kg. Some samples in urban areas had copper concentrations in the 50-150 mg/kg range, while rural samples never exceeded 30 mg/kg. This suggests urban contribution above natural levels. Lead is well known to be elevated in surface soils due to industrial activities and the historical use of leaded gasoline (Kabata-Pendias and Pendias, 1984). Lead concentrations were highest in the urban Piedmont region (several samples in the 300 mg/kg range), somewhat lower in the urban Coastal plain (usually less than 200 mg/kg), and lowest in the rural study (only two samples greater than 125 mg/kg).

Mean total arsenic concentrations in United States soils have been reported to be 5-8 mg/kg (Kabata-Pendias and Pendias, 1984). In this study, median

Table 1. Ambient Concentration of Extractable Metals Measured in 248 New Jersey soil samples.

		Urbai	n Piedmont		Urban Coastal Plain			
	Method	No. of	Median	90th Percentile	Method	No. of	Median	90th Percentile
	Detection	detects	Concentration	Concentration	Detection	detects	Concentration	Concentration
	Limit (mg/kg)	n=67	(mg/kg)	(mg/kg)	Limit (mg/kg)	n=91	(mg/kg)	(mg/kg)
Aluminum	1.5	67	10500	14400	20	91	6800	10800
Antimony	1.7	17	<dl< td=""><td>3.48</td><td>6</td><td>0</td><td><dl< td=""><td><dl< td=""></dl<></td></dl<></td></dl<>	3.48	6	0	<dl< td=""><td><dl< td=""></dl<></td></dl<>	<dl< td=""></dl<>
•	0.13	67	5.2	24.2	0	82	5.2	13.6
Arsenic					1			
Barium	0.22	67	80.6	168	20	60	28.3	65.8
Beryllium	0.14	65	0.51	0.82	0.5	15	<dl< td=""><td>0.68</td></dl<>	0.68
Cadmium	0.4	21	<dl< td=""><td>0.67</td><td>0.5</td><td>5</td><td><dl< td=""><td><dl< td=""></dl<></td></dl<></td></dl<>	0.67	0.5	5	<dl< td=""><td><dl< td=""></dl<></td></dl<>	<dl< td=""></dl<>
Calcium	11.2	67	1425	3010	500	59	995	2000
Chromium	0.9	67	18.5	29.9	1	91	11.8	34.7
Cobalt	0.5	67	6.3	10.4	5	7	<dl< td=""><td><dl< td=""></dl<></td></dl<>	<dl< td=""></dl<>
Copper	0.52	67	29.5	75.5	2.5	82	9.3	33.3
Iron	2.2	67	14600	20000	10	91	8830	21100
Lead	0.063	67	111	297	10	82	37.6	144
Magnesium	5	67	2190	4614	500	54	673	1870
Manganese	0.21	67	311	859	1.5	91	62.4	206
Mercury	0.1	50	0.18	0.63	0.1	39	<dl< td=""><td>0.21</td></dl<>	0.21
Nickel	0.9	67	12.4	24.6	4	43	<dl< td=""><td>12.3</td></dl<>	12.3
Potassium	32	67	693	1524	500	45	<dl< td=""><td>1750</td></dl<>	1750
Selenium	0.5	61	0.41	0.71	1	0	<dl< td=""><td><dl< td=""></dl<></td></dl<>	<dl< td=""></dl<>
Silver	0.22	28	<dl< td=""><td>0.86</td><td>1</td><td>3</td><td><dl< td=""><td><dl< td=""></dl<></td></dl<></td></dl<>	0.86	1	3	<dl< td=""><td><dl< td=""></dl<></td></dl<>	<dl< td=""></dl<>
Sodium	16.4	60	90.1	141	500	0	<dl< td=""><td><dl< td=""></dl<></td></dl<>	<dl< td=""></dl<>
Thallium	0.2	28	<dl< td=""><td>0.25</td><td>1</td><td>2</td><td><dl< td=""><td><dl< td=""></dl<></td></dl<></td></dl<>	0.25	1	2	<dl< td=""><td><dl< td=""></dl<></td></dl<>	<dl< td=""></dl<>
Vanadium	0.95	67	29.6	41.7	5	86	16	35.5
Zinc	2	67	75.3	162	2	88	39.9	106

Rural Areas of New Jersey

	Rural studies	Ridge and Valley Province			F	Highlands Province			Coastal Plain Province		
	Method	No. of	Median	90th Percentile	No. of	Median	90th Percentile	No. of	Median	90th Percentile	
	Detection	detects	Concentration	Concentration	detects	Concentration	Concentration	detects	Concentration	Concentration	
	Limit (mg/kg)	n=23	(mg/kg)	(mg/kg)	n=23	(mg/kg)	(mg/kg)	n=44	(mg/kg)	(mg/kg)	
Aluminum	3.23	23	15300	21080	23	16800	28980	44	1375	6760	
Antimony	0.42	0	<dl< td=""><td><dl< td=""><td>0</td><td><dl< td=""><td><dl< td=""><td>11</td><td><dl< td=""><td>0.56</td></dl<></td></dl<></td></dl<></td></dl<></td></dl<>	<dl< td=""><td>0</td><td><dl< td=""><td><dl< td=""><td>11</td><td><dl< td=""><td>0.56</td></dl<></td></dl<></td></dl<></td></dl<>	0	<dl< td=""><td><dl< td=""><td>11</td><td><dl< td=""><td>0.56</td></dl<></td></dl<></td></dl<>	<dl< td=""><td>11</td><td><dl< td=""><td>0.56</td></dl<></td></dl<>	11	<dl< td=""><td>0.56</td></dl<>	0.56	
Arsenic	0.28	23	4.9	7.32	23	4.8	9.96	36	1.15	6.15	
Barium	0.09	19	60.2	101.16	22	69.6	96.64	34	7.25	55.31	
Beryllium	0.01	8	<dl< td=""><td>0.91</td><td>19</td><td>0.73</td><td>1.08</td><td>9</td><td><dl< td=""><td>0.14</td></dl<></td></dl<>	0.91	19	0.73	1.08	9	<dl< td=""><td>0.14</td></dl<>	0.14	
Cadmium	0.03	0	<dl< td=""><td><dl< td=""><td>11</td><td><dl< td=""><td>0.32</td><td>9</td><td><dl< td=""><td>0.13</td></dl<></td></dl<></td></dl<></td></dl<>	<dl< td=""><td>11</td><td><dl< td=""><td>0.32</td><td>9</td><td><dl< td=""><td>0.13</td></dl<></td></dl<></td></dl<>	11	<dl< td=""><td>0.32</td><td>9</td><td><dl< td=""><td>0.13</td></dl<></td></dl<>	0.32	9	<dl< td=""><td>0.13</td></dl<>	0.13	
Calcium	41.7	10	<dl< td=""><td>2272</td><td>20</td><td>1160</td><td>4518</td><td>33</td><td>76.4</td><td>341.7</td></dl<>	2272	20	1160	4518	33	76.4	341.7	
Chromium	0.17	23	14.3	21.2	23	17.7	26.64	44	2.9	11.76	
Cobalt	0.18	12	7.3	11.4	18	6.8	12.44	32	0.37	1.18	
Copper	0.33	23	17.2	26.04	23	16	28.96	44	4.2	11.43	
Iron	4	23	14800	28540	23	18700	27860	44	1795	10587	
Lead	0.22	23	31.6	54	23	26.6	59.02	44	17.5	54.05	
Magnesium	14.4	19	2600	7182	23	2340	4024	34	79.65	513.2	
Manganese	0.27	23	470	1192	23	407	836.8	44	11.65	35.39	
Mercury	0.016	22	0.1	0.15	23	0.09	0.18	28	0.04	0.14	
Nickel	0.21	20	15.7	22.5	23	11.6	19.04	30	0.84	3.87	
Potassium	6.63	17	961	1660	21	955	1456	30	76	328	
Selenium	0.49	0	<dl< td=""><td><dl< td=""><td>10</td><td><dl< td=""><td>0.99</td><td>7</td><td><dl< td=""><td>0.68</td></dl<></td></dl<></td></dl<></td></dl<>	<dl< td=""><td>10</td><td><dl< td=""><td>0.99</td><td>7</td><td><dl< td=""><td>0.68</td></dl<></td></dl<></td></dl<>	10	<dl< td=""><td>0.99</td><td>7</td><td><dl< td=""><td>0.68</td></dl<></td></dl<>	0.99	7	<dl< td=""><td>0.68</td></dl<>	0.68	
Silver	0.1	0	<dl< td=""><td><dl< td=""><td>5</td><td><dl< td=""><td>0.21</td><td>0</td><td><dl< td=""><td><dl< td=""></dl<></td></dl<></td></dl<></td></dl<></td></dl<>	<dl< td=""><td>5</td><td><dl< td=""><td>0.21</td><td>0</td><td><dl< td=""><td><dl< td=""></dl<></td></dl<></td></dl<></td></dl<>	5	<dl< td=""><td>0.21</td><td>0</td><td><dl< td=""><td><dl< td=""></dl<></td></dl<></td></dl<>	0.21	0	<dl< td=""><td><dl< td=""></dl<></td></dl<>	<dl< td=""></dl<>	
Sodium	6.78	0	<dl< td=""><td><dl< td=""><td>8</td><td><dl< td=""><td>85.1</td><td>28</td><td>54.65</td><td>91.9</td></dl<></td></dl<></td></dl<>	<dl< td=""><td>8</td><td><dl< td=""><td>85.1</td><td>28</td><td>54.65</td><td>91.9</td></dl<></td></dl<>	8	<dl< td=""><td>85.1</td><td>28</td><td>54.65</td><td>91.9</td></dl<>	85.1	28	54.65	91.9	
Thallium	0.41	0	<dl< td=""><td><dl< td=""><td>0</td><td><dl< td=""><td><dl< td=""><td>1</td><td><dl< td=""><td><dl< td=""></dl<></td></dl<></td></dl<></td></dl<></td></dl<></td></dl<>	<dl< td=""><td>0</td><td><dl< td=""><td><dl< td=""><td>1</td><td><dl< td=""><td><dl< td=""></dl<></td></dl<></td></dl<></td></dl<></td></dl<>	0	<dl< td=""><td><dl< td=""><td>1</td><td><dl< td=""><td><dl< td=""></dl<></td></dl<></td></dl<></td></dl<>	<dl< td=""><td>1</td><td><dl< td=""><td><dl< td=""></dl<></td></dl<></td></dl<>	1	<dl< td=""><td><dl< td=""></dl<></td></dl<>	<dl< td=""></dl<>	
Vanadium	0.07	23	20.7	29.68	23	32.3	52.98	40	7.8	20.68	
Zinc	0.18	23	75.8	112.3	23	69.7	111.6	44	6.7	27.64	

extractable arsenic concentrations were typically about 5 mg/kg, and concentrations never exceeded 15 mg/kg in the rural study. In the urban studies, the maximum concentration was 83 mg/kg, and 95th percentile values were below 30 mg/kg. These higher concentrations are within the normal ranges for soils, particularly glauconitic soils naturally high in arsenic (Dooley, 2001). Some soil samples in the urban Coastal Plain study were taken from glauconitic soils, and they yielded some of the higher concentrations measured (including the maximum value measured, 83 mg/kg). However, other samples from this study

with elevated arsenic were not from glauconitic regions. Furthermore, the urban Piedmont study did not include glauconitic soils, where a significant percentage of the samples also exhibited elevated levels of arsenic relative to the rural study. This suggests some anthropogenic contribution of arsenic to urban surface soils.

Mean total beryllium concentrations in United States soils are about 1-2 ppm (Kabata-Pendias and Pendias, 1984). Extractable concentrations of beryllium in this study ranged from non-detectable to

3 mg/kg. Total antimony concentrations in United States soils are generally less than 1 ppm (Kabata-Pendias and Pendias, 1984). This study yielded extractable antimony concentrations that ranged from non-detectable levels to 13 mg/kg, with the higher concentrations being associated with the urban Piedmont samples. Thallium, mercury, cadmium, silver and selenium are naturally present at only trace levels in soils (usually less than 1 ppm) in soils (Kabata-Pendias and Pendias, 1984). In this study, thallium and silver were seldom detected, and selenium, cadmium and mercury were either not detected or measured at concentrations that were usually less than 1 mg/kg. A few samples, mostly from urban areas, contained cadmium at concentrations above 0.5 mg/kg or mercury above 1.0 mg/kg, which suggests contribution from industrial activity or regional atmospheric deposition.

Statistically, several metals were found at lower concentrations in the coastal plain regions of the state relative to the remainder of the state. Coastal Plain soils have high sand contents, lower organic carbon contents and lower pH values than soils in the remainder of the state, which decrease their affinity for metals.

Discussion

The metals concentration data collected in these series of studies are useful for determining typical ambient levels of the various metals in the geographical regions studied. Levels significantly above these concentrations may indicate a spill or discharge has occurred. The metals of greatest environmental concern are those in which ambient levels in surface soils equal or exceed concentrations that are considered hazardous to human health. In such cases, regulatory levels applicable for remedial activities at contaminated sites may be limited by prevailing ambient concentrations. The most important example of this situation occurs with arsenic. The health-based soil ingestion criterion for arsenic (0.4 mg/kg) is significantly below observed ambient levels, even in soils unaffected by human activity. All studies except the rural Coastal Plain study yielded median extractable arsenic concentrations of about 5 mg/kg, more than ten times the health-based number. The rural Coastal Plain median concentration (1 mg/kg) was also above the health-based ingestion criteria.

The current soil cleanup criterion for arsenic in soil is 20 mg/kg, based on an earlier assessment of ambient levels. The 90th percentile concentration of arsenic from the urban Piedmont study (26 mg/kg) is somewhat higher than this value. In the urban Coastal Plain study, three of the ninety-one samples

yielded arsenic levels above 20 mg/kg. The rural areas of the state yielded 95th percentile concentrations ranging from 5-10 mg/kg, and no samples exceeded 20 mg/kg.

Total arsenic levels in glauconite-bearing soils in New Jersey have been reported to range from 13-131 mg/kg, with a median of 30 mg/kg (Dooley, 2001). When only extractable arsenic from 0-6" soil samples are considered from the glauconite study, the median, 90th percentile and maximum concentrations are 8.2, 27.5, and 63.3 mg/kg, respectively. These concentrations are only moderately higher than those observed for the urban studies.

The health-based non-residential soil criterion for hexavalent chromium based on soil inhalation is 20 mg/kg, which is below the 90th percentile ambient concentrations reported in this study. However, chromium in soil has been reported to be dominated by the much less toxic trivalent form, so ambient concentrations are not likely to be of concern for this metal. A hexavalent chromium-specific method would be recommended for analysis of soil when this metal is known to be of concern.

With regards to the remaining metals, only one sample yielded a result where current criteria were exceeded. This sample was taken from the urban Piedmont region and yielded a lead concentration of 464 mg/kg, which is slightly greater than the current lead criterion of 400 mg/kg.

Other than arsenic, the only other metal with the current soil criterion set by ambient levels is beryllium (criterion = 2 mg/kg, based on earlier New Jersey data and other literature). In this study, 90th percentile concentrations frequently approached 1 mg/kg (Table 1), 95th percentile concentrations in the rural highlands exceeded 1 mg/kg (1.4 mg/kg), and overall, ten samples out of 248 yielded concentrations between 1 and 2 mg/kg. Only one sample exceeded 2 mg/kg (2.8 mg/kg from the rural Highlands province).

The thallium criterion is currently set by the Practical Quantitation Level (2 mg/kg), since the health-based criteria is lower (zero). Reporting Levels and/or Practical Quantitation Levels for this metal from the two New Jersey certified laboratories used in this study were somewhat lower (1 mg/kg or less), which suggests that analytical capabilities may have improved since the thallium criterion was set.

Metal concentrations reported in this summary were acid-extractable metals, not necessarily total metal concentrations. The extraction method (USEPA

Method 3050) is a vigorous extraction method designed to remove all metals that could become "environmentally available". In practice, the extraction method likely overestimates metals that could become available, since the sample is refluxed with both concentrated nitric acid and hydrogen peroxide. However, by design, the method will not extract chemicals incorporated in silica minerals, as they are usually not mobile in the environment. Thus, concentrations reported in these studies may be lower than those indicated from analyses using methods designed to measure total metal concentrations, such as x-ray fluorescence methods.

References

BEM Systems, Inc. (1997). Characterization of Ambient Levels of Selected Metals and Other Analytes in New Jersey Soils: Year 1, Urban Piedmont Region. Final Report to NJ Dept. of Environmental Protection, Division of Science and Research, Trenton, NJ.

BEM Systems, Inc. (1998). Characterization of Ambient Levels of Selected Metals and Other Analytes in New Jersey Urban Coastal Plain Region Soils. Final Report to NJ Dept. of Environmental Protection, Division of Science and Research, Trenton, NJ.

BEM Systems, Inc. (2002). Characterization of Ambient Levels of Selected Metals and cPAHs in New Jersey Soils: Year III – Rural Areas of New Jersey Highlands, Valley and Ridge, and Coastal Plain Physiographic Provinces. Final Report to NJ Dept. of Environmental Protection, Division of Science, Research and Technology, Trenton, NJ.

Dooley, J. H. (2001). Baseline Concentrations of Arsenic, Beryllium and Associated Elements in Glauconite and Glauconitic Soils in the New Jersey Coastal Plain. New Jersey Geological Survey Investigation Report, NJ Dept. of Environmental Protection, Trenton, NJ.

Kabata-Pendias, A. and Pendias, H. (1984). Trace Elements in Soils and Plants. CRC Press, Inc. Boca Raton, Florida.

Funding

These studies were funded by the Hazardous Waste Research fund and conducted by BEM Systems, Inc.

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Visit the DSRT web site @ www.state.nj.us/dep/dsr



RESEARCH PROJECT SUMMARY

Fact Sheet: Draft Aquatic Life Ambient Water Quality Criteria for Aluminum in Freshwaters

Summary

EPA published a draft update of aluminum aquatic life ambient water quality criteria for freshwaters under Section 304(a)(1) of the Clean Water Act to reflect the latest scientific knowledge. EPA will accept public comment on the draft criteria for 60 days upon publication of the Federal Register notice. Once final, the criteria will serve as recommendations to states and tribes by defining the concentration of aluminum in water that will protect against harmful effects to aquatic life.

Background

EPA first published criteria for aluminum in 1988 to protect aquatic life from harmful effects of aluminum toxicity in freshwaters. Aluminum can inhibit an aquatic organism's ability to regulate salt concentrations and clog fish gills, potentially resulting in death or affecting growth and reproduction. EPA is updating the aluminum criteria to better reflect the latest science. Unfortunately, there are not enough data to support the development of estuarine/marine criteria at this time.

Unlike the fixed acute and chronic values found in the 1988 document, this draft document provides users the flexibility to develop site-specific criteria based on a site's water chemistry using the *Aluminum Criteria Calculator V.1.0.xlsx* or by using the lookup tables provided in the criteria appendix. Studies have shown that three water chemistry parameters, pH, dissolved organic carbon (DOC), and hardness, can affect the toxicity of aluminum by affecting the bioavailability of aluminum in the water to aquatic species.

What is Aluminum and How Does it Enter the Water?

Aluminum is found in most soils and rocks and is the third most abundant element and the most common metal in the earth's crust. Aluminum can enter the water via natural processes, like weathering of rocks. Aluminum is also released to water by mining, industrial processes using aluminum, and waste water treated with alum, an aluminum compound.

How does Aluminum Affect Aquatic Life?

Aluminum is considered a non-essential metal because fish and other aquatic life don't need it to function. Elevated levels of aluminum can affect some species ability to regulate ions, like salts, and inhibit respiratory functions, like breathing.

Aluminum can accumulate on the surface of a fish's gill, leading to respiratory dysfunction, and possibly death. Aquatic plants are generally less sensitive to aluminum than fish and other aquatic life.

What is a Water Quality Parameter and Why is it Important?

Bioavailability is the measure whether a substance in the environment is available to enter living organisms, like fish. The bioavailability of aluminum is dependent on the chemical properties of water, otherwise known as water quality parameters. The more bioavailable the aluminum is, the more likely it is to cause a toxic effect. The water quality parameters that have the greatest impact on aluminum's bioavailability are pH, DOC, and hardness.

pH: a low pH generally makes it easier for aluminum to be dissolved, and therefore more bioavailable. At higher pH, aluminum speciation changes make it more bioavailable.

- DOC: higher dissolved organic carbon reduces the bioavailability of aluminum because it binds to form aluminum complexes.
- Hardness: higher hardness values mean there are more ions present that compete with aluminum.
 This makes aluminum less bioavailable.

What is the Recommended Level of Aluminum in Freshwater for the Protection of Aquatic Life?

The recommended level of aluminum in freshwater depends on a site's water quality parameters. Unlike the fixed values found in the 1988 criteria document, these criteria use a Multiple Linear Regression (MLR) model to normalize the data, and the criteria are based on a site's pH, DOC, and hardness. See Table 1 for a comparison of existing and draft criteria values.

For freshwater criteria, users can put their site's water quality parameters into the *Aluminum Criteria Calculator V.1.0.xlsx* or use the lookup tables in the document's appendix. The resulting acute criterion would have an appropriate level of protection if the one-hour average concentration is not exceeded more than once every three years on average. If the four-day average concentration is not exceeded more than once every three years on average, the chronic criterion is protective.

Where can I find more information?

For more information please visit EPA's website at www.epa.gov/wqc/aquatic-life-criteria-aluminum or contact Diana Eignor at Eignor.Diana@epa.gov.

Table 1: 2017 Draft Aluminum Aquatic Life Criteria Compared to Current 1988 Criteria^a

Version	Freshwater Acute (1 day, total aluminum)	Freshwater Chronic (4-day, total aluminum)
2017 Draft AWQC Criteria (MLR normalized to pH = 7, hardness = 100 mg/L, DOC = 1 mg/L)	1,400 μg/L	390 μg/L
1988 AWQC Criteria (pH 6.5 – 9.0, across all hardness and DOC ranges)	750 μg/L	87 μg/L

^a Values are recommended not to be exceeded more than once every three years on average. Note: Values will be different under differing water chemistry conditions as identified in this document.

2012 Recreational Water Quality Criteria

Summary

EPA has released its 2012 recreational water quality criteria (RWQC) recommendations for protecting human health in all coastal and non-coastal waters designated for primary contact recreation use. EPA provides two sets of recommended criteria. Primary contact recreation is protected if either set of criteria recommendations are adopted into state water quality standards.

These recommendations are intended as guidance to states, territories and authorized tribes in developing water quality standards to protect swimmers from exposure to water that contains organisms that indicate the presence of fecal contamination.

Background

EPA last issued ambient water quality criteria recommendations for recreational waters in 1986. EPA issues such recommendations under the authority of the Clean Water Act (CWA). Amendments to the CWA by the Beaches Environmental Assessment and Coastal Health (BEACH) Act of 2000 direct EPA to conduct studies associated with pathogens and human health, and to publish new or revised criteria recommendations for pathogens and pathogen indicators based on those studies. These 2012 RWQC meet those requirements.

The 2012 RWQC rely on the latest research and science, including studies that show a link between illness and fecal contamination in recreational waters. They are based on the use of two bacterial indicators of fecal contamination, *E. coli* and enterococci. The new criteria are designed to protect primary contact recreation, including swimming, bathing, surfing, water skiing, tubing, water play by children, and similar water contact activities where a high degree of bodily contact with the water, immersion and ingestion are likely.

What are the recommendations?

The 2012 RWQC offer two sets of numeric concentration thresholds, either of which would protect the designated use of primary contact recreation and, therefore, would protect the public from exposure to harmful levels of pathogens. Illness rates upon which these recommendations are based use the National Epidemiological and Environmental Assessment of Recreational Water (NEEAR) definition of gastrointestinal illness, which is not limited to illnesses which exhibit a fever.

The RWQC consist of three components: magnitude, duration and frequency. The magnitude of the bacterial indicators are described by both a geometric mean (GM) and a statistical threshold value (STV) for the bacteria samples. The STV approximates the 90th percentile of the water quality distribution and is intended to be a value that should not be exceeded by more than 10 percent of the samples taken. The table summarizes the magnitude component of the recommendations. All three components are explained in more detail in the sections below.

CRITERIA ELEMENTS		endation 1 ss Rate 36/1,000	Recommendation 2 Estimated Illness Rate 32/1,000		
Indicator	GM (cfu/100 mL)	STV (cfu/100 mL)	GM (cfu/100 mL)	STV (cfu/100 mL)	
Enterococci (marine & fresh)	35	130	30	110	
E. coli (fresh)	126	410	100	320	

Water quality criteria recommendations are intended as guidance in establishing new or revised water quality standards. They are not regulations themselves. States and authorized tribes have the discretion to adopt, where appropriate, other scientifically defensible water quality criteria that differ from EPA's recommended criteria.

RECOMMENDATION 1: MAGNITUDE

Enterococci: Culturable enterococci at a

geometric mean (GM) of 35 colony forming units (CFU per 100 milliliters (mL) and a statistical threshold value (STV) of 130 cfu per 100 mL, measured using *EPA Method 1600*, or any other equivalent method that measures culturable enterococci.

E. coli: Culturable *E. coli* at a GM of 126 cfu per 100 mL and an STV of 410 cfu per 100 mL measured using *EPA Method 1603*, or any other equivalent method that measures culturable *E. coli*.

RECOMMENDATION 2: MAGNITUDE

Enterococi: Culturable enterococci at a GM of 30 cfu per 100 mL and an STV of 110 cfu per 100 mL, measured using *EPA Method 1600*, or any other equivalent method that measures culturable enterococci.

E. coli: Culturable *E. coli* at a GM of 100 cfu per 100 mL and an STV of 320 cfu per 100 mL measured using *EPA Method 1603*, or any other equivalent method that measures culturable *E. coli*.

FOR BOTH RECOMMENDATIONS

Duration and Frequency: The waterbody GM should not be greater than the selected GM magnitude in any 30-day interval. There should not be greater than a ten percent excursion frequency of the selected STV magnitude in the same 30-day interval.

How are these criteria different from the 1986 criteria?

Similar Protection for Fresh and Marine Waters: The EPA used an analysis of NEEAR water quality data to refine the illness rate estimate for the recommended marine criterion for entercocci. The 2012 RWQC values now protect public health similarly in both marine and fresh waters.

A New Measurement Value: EPA is introducing a new term, Statistical Threshold Value (STV), to be used in conjunction with the recommended GM value.

New Early Alert Tool: In addition to recommending criteria values, EPA is now also

providing states with Beach Action Values (BAVs) for use in notification programs. The BAV is provided for states to use as a precautionary tool to provide an early alert to beachgoers, including families with children.

A Single Level of Beach Use: The 1986 bacteria criteria document included four single sample maximum (SSM) values appropriate for different levels of beach usage (use intensities). In the 2012 RWQC, EPA removed those recommendations and instead provided states with optional, precautionary BAVs for use in monitoring and notification programs.

More Tools for Assessing and Managing **Recreational Waters**: EPA is providing information on tools for evaluating and managing recreational waters, such as predictive modeling and sanitary surveys. The Agency is also providing tools for developing site-specific criteria such as epidemiological studies, quantitative microbial risk assessment, and use of alternative indicators or methods. The EPA has developed and validated a molecular testing method using quantitative polymerase chain reaction (qPCR) as a rapid analytical technique for the detection of enterococci in recreational water (EPA Method 1611). For the purposes of beach monitoring, a state may use a qPCR method on a site-specific basis.

Where can I find more information?

EPA has put the 2012 RWQC document, support documents, and the Federal Register Notice, in the docket (Docket identification No. EPA-HQ-OW-2011-0466) which can be accessed via EPA's website at http://water.epa.gov/scitech/swguidance/standards/criteria/health/recreation/index.cfm.

You can also contact Sharon Nappier at mappier.sharon@epa.gov or (202)566-0740, or contact Tracy Bone at bone.tracy@epa.gov or (202) 564-5257 for more information.



Total Nitrogen

Total Nitrogen is an essential nutrient for plants and animals. However, an excess amount of nitrogen in a waterway can lead to low levels of dissolved oxygen and negatively alter various plant life and organisms. Sources of nitrogen include: wastewater treatment plants, runoff from fertilized lawns croplands, failing systems, runoff from animal manure and storage areas, and industrial discharges that contain corrosion inhibitors.



Storm runoff from a cattle operation can increase Total Nitrogen levels in a water body.

<u>Understanding Total Nitrogen:</u> There are three forms of nitrogen that are commonly measured in water bodies: ammonia, nitrates and nitrites. Total nitrogen is the sum of total kjeldahl nitrogen (organic and reduced nitrogen), ammonia, and nitrate-nitrite. It can be derived by monitoring for total kjeldahl nitrogen (TKN), ammonia and nitrate-nitrite individually and adding the components together. An acceptable range of total nitrogen is 2 mg/L to 6 mg/L, though it is recommended to check tribal, state, or federal standards for an adequate comparison of your data.



Trash areas like this may leach chemicals that can increase Total Nitrogen during a storm event into a water body.

Monitoring Equipment: Depending upon monitoring objectives set forth in an environmental program, the following equipment options are commonly used to collect total nitrogen data from the field.

Readily available and economically priced:

• Total Nitrogen Kits

For each component of total nitrogen, the following can be used and are of greater precision and higher cost:

- Meters
- Multiparameter Probes
- Contract Laboratories (if necessary)

For additional information:

www.epa.gov/owow/monitoring/volunteer/stream



www.epa.gov

August 1993

Method 351.2, Revision 2.0: Determination of Total Kjeldahl Nitrogen by Semi-Automated Colorimetry

METHOD 351.2

DETERMINATION OF TOTAL KJELDAHL NITROGEN BY SEMI-AUTOMATEDCOLORIMETRY

Edited by James W. O'Dell Inorganic Chemistry Branch Chemistry Research Division

> Revision 2.0 August 1993

ENVIRONMENTAL MONITORING SYSTEMS LABORATORY OFFICE OF RESEARCH AND DEVELOPMENT U.S. ENVIRONMENTAL PROTECTION AGENCY CINCINNATI, OHIO 45268

METHOD 351.2

DETERMINATION OF TOTAL KJELDAHL NITROGEN BY SEMI-AUTOMATED COLORIMETRY

1.0 SCOPE AND APPLICATION

- 1.1 This method covers the determination of total Kjeldahl nitrogen in drinking, ground, and surface waters, domestic and industrial wastes. The procedure converts nitrogen components of biological origin such as amino acids, proteins and peptides to ammonia, but may not convert the nitrogenous compounds of some industrial wastes such as amines, nitro compounds, hydrazones, oximes, semicarbazones and some refractory tertiary amines.
- 1.2 The applicable range is 0.1-20 mg/L TKN. The range may be extended with sample dilution.

2.0 SUMMARY OF METHOD

- 2.1 The sample is heated in the presence of sulfuric acid, H_2SO_4 for two and one half hours. The residue is cooled, diluted to 25 mL and analyzed for ammonia. This digested sample may also be used for phosphorus determination.
- 2.2 Total Kjeldahl nitrogen is the sum of free-ammonia and organic nitrogen compounds which are converted to ammonium sulfate $(NH_4)_2SO_4$, under the conditions of digestion described.
- 2.3 Organic Kjeldahl nitrogen is the difference obtained by subtracting the freeammonia value from the total Kjeldahl nitrogen value.
- 2.4 Reduced volume versions of this method that use the same reagents and molar ratios are acceptable provided they meet the quality control and performance requirements stated in the method.
- 2.5 Limited performance-based method modifications may be acceptable provided they are fully documented and meet or exceed requirements expressed in Section 9.0, Quality Control.

3.0 **DEFINITIONS**

- 3.1 **Calibration Blank (CB)** -- A volume of reagent water fortified with the same matrix as the calibration standards, but without the analytes, internal standards, or surrogate analytes.
- 3.2 **Calibration Standard (CAL)** -- A solution prepared from the primary dilution standard solution or stock standard solutions and the internal standards and

- surrogate analytes. The CAL solutions are used to calibrate the instrument response with respect to analyte concentration.
- 3.3 **Instrument Performance Check Solution (IPC)** -- A solution of one or more method analytes, surrogates, internal standards, or other test substances used to evaluate the performance of the instrument system with respect to a defined set of criteria.
- 3.4 **Laboratory Fortified Blank (LFB)** -- An aliquot of reagent water or other blank matrices to which known quantities of the method analytes are added in the laboratory. The LFB is analyzed exactly like a sample, and its purpose is to determine whether the methodology is in control, and whether the laboratory is capable of making accurate and precise measurements.
- 3.5 **Laboratory Fortified Sample Matrix (LFM)** -- An aliquot of an environmental sample to which known quantities of the method analytes are added in the laboratory. The LFM is analyzed exactly like a sample, and its purpose is to determine whether the sample matrix contributes bias to the analytical results. The background concentrations of the analytes in the sample matrix must be determined in a separate aliquot and the measured values in the LFM corrected for background concentrations.
- Laboratory Reagent Blank (LRB) -- An aliquot of reagent water or other blank matrices that are treated exactly as a sample including exposure to all glassware, equipment, solvents, reagents, internal standards, and surrogates that are used with other samples. The LRB is used to determine if method analytes or other interferences are present in the laboratory environment, the reagents, or the apparatus.
- 3.7 **Linear Calibration Range (LCR)** -- The concentration range over which the instrument response is linear.
- 3.8 **Material Safety Data Sheet (MSDS)** -- Written information provided by vendors concerning a chemical's toxicity, health hazards, physical properties, fire, and reactivity data including storage, spill, and handling precautions.
- 3.9 **Method Detection Limit (MDL)** -- The minimum concentration of an analyte that can be identified, measured and reported with 99% confidence that the analyte concentration is greater than zero.
- 3.10 **Quality Control Sample (QCS)** -- A solution of method analytes of known concentrations that is used to fortify an aliquot of LRB or sample matrix. The QCS is obtained from a source external to the laboratory and different from the source of calibration standards. It is used to check laboratory performance with externally prepared test materials.

3.11 **Stock Standard Solution (SSS)** -- A concentrated solution containing one or more method analytes prepared in the laboratory using assayed reference materials or purchased from a reputable commercial source.

4.0 INTERFERENCES

- 4.1 High nitrate concentrations (10X or more than the TKN level) result in low TKN values. If interference is suspected, samples should be diluted and reanalyzed.
- 4.2 Method interferences may be caused by contaminants in the reagent water, reagents, glassware, and other sample processing apparatus that bias analyte response.

5.0 SAFETY

- 5.1 The toxicity or carcinogenicity of each reagent used in this method have not been fully established. Each chemical should be regarded as a potential health hazard and exposure should be as low as reasonably achievable. Cautions are included for known extremely hazardous materials or procedures.
- 5.2 Each laboratory is responsible for maintaining a current awareness file of OSHA regulations regarding the safe handling of the chemicals specified in this method. A reference file of Material Safety Data Sheets (MSDS) should be made available to all personnel involved in the chemical analysis. The preparation of a formal safety plan is also advisable.
- 5.3 The following chemicals have the potential to be highly toxic or hazardous, consult MSDS.
 - 5.3.1 Mercury (Sections 7.2 and 7.3)
 - 5.3.2 Sulfuric acid (Sections 7.2, 7.3, and 7.4)
 - 5.3.3 Sodium nitroprusside (Section 7.9)

6.0 EQUIPMENT AND SUPPLIES

- 6.1 Balance Analytical, capable of accurately weighing to the nearest 0.000l g.
- 6.2 Glassware Class A volumetric flasks and pipets as required.
- 6.3 Block digestor with tubes.
- 6.4 Automated continuous flow analysis equipment designed to deliver and react sample and reagents in the required order and ratios.
 - 6.4.1 Sampling device (sampler)

- 6.4.2 Multichannel pump
- 6.4.3 Reaction unit or manifold
- 6.4.4 Colorimetric detector
- 6.4.5 Data recording device

7.0 REAGENTS AND STANDARDS

- 7.1 Reagent water: Ammonia free distilled or deionized water, free of the analyte of interest. ASTM Type II or equivalent.
- 7.2 Mercuric sulfate: Dissolve 8 g red mercuric oxide (HgO) (CASRN 21908-53-2) in 50 mL of 1:4 sulfuric acid (10 mL conc. H_2SO_4 : [CASRN 7664-93-9] 40 mL reagent water) and dilute to 100 mL with reagent water.
- 7.3 Digestion solution: (Sulfuric acid-mercuric sulfate-potassium sulfate solution): Dissolve 133 g of $\rm K_2SO_4$ (CASRN 7778-80-5) in 700 mL of reagent water and 200 mL of conc. $\rm H_2SO_4$. Add 25 mL of mercuric sulfate solution (Section 7.1) and dilute to 1 L.
 - **Note 1:** An alternate mercury-free digestion solution can be prepared by dissolving 134 g K_2SO_4 and 7.3 g CuSQ in 800 mL reagent water and then adding 134 mL conc. H_2SO_4 and diluting to 1 L. Use 10 mL solution per 25 mL of sample.
- 7.4 Sulfuric Acid solution (4%): Add 40 mL of conc. sulfuric acid to 800 mL of reagent water, cool and dilute to 1 L.
 - **Note 2:** If alternate mercury-free digestion solution is used, adjust the above solution to equal the acid concentration of the digested sample (Section 11.6).
- 7.5 Stock Sodium Hydroxide (20%): Dissolve 200 g of sodium hydroxide (CASRN 1310-73-2) in 900 mL of reagent water and dilute to 1 L.
- 7.6 Stock Sodium Potassium Tartrate solution (20%): Dissolve 200 g sodium potassium tartrate (CASRN 6381-59-5) in about 800 mL of reagent water and dilute to 1 L.
- 7.7 Stock Buffer solution: Dissolve 134.0 g of sodium phosphate, dibasic (Na_2HPO_4) (CASRN 7558-79-4) in about 800 mL of reagent water. Add 20 g of sodium hydroxide and dilute to 1 L.
- 7.8 Working Buffer solution: Combine the reagents in the stated order, add 250 mL of stock sodium potassium tartrate solution (Section 7.6) to 200 mL of stock buffer solution (Section 7.7) and mix. Add xx mL sodium hydroxide solution

- (Section 7.5) and dilute to 1 L. See concentration ranges, Table 2, for composition of working buffer.
- 7.9 Sodium Salicylate/Sodium Nitroprusside solution: Dissolve 150 g of sodium salicylate (CASRN 54-21-7) and 0.3 g of sodium nitroprusside (CASRN 13755-38-9 or 14402-89-2) in about 600 mL of reagent water and dilute to 1 L.
- 7.10 Sodium Hypochlorite solution: Dilute 6.0 mL sodium hypochlorite solution (CASRN 7681-52-9) (Clorox) to 100 mL with reagent water.
- 7.11 Ammonium chloride, stock solution: Dissolve 3.819 g NH₄Cl (CASRN 12125-02-9) in reagent water and bring to volume in a 1 L volumetric flask. 1 mL = 1.0 mg NH₃-N.
- 7.12 Teflon boiling chips.

8.0 SAMPLE COLLECTION, PRESERVATION AND STORAGE

- 8.1 Samples should be collected in plastic or glass bottles. All bottles must be thoroughly cleaned and rinsed with reagent water. Volume collected should be sufficient to insure a representative sample, allow for replicate analysis (if required), and minimize waste disposal.
- 8.2 Samples must be preserved with H_2SO_4 to a pH <2 and cooled to 4°C at the time of collection.
- 8.3 Samples should be analyzed as soon as possible after collection. If storage is required, preserved samples are maintained at 4°C and may be held for up to 28 days.

9.0 QUALITY CONTROL

9.1 Each laboratory using this method is required to operate a formal quality control (QC) program. The minimum requirements of this program consist of an initial demonstration of laboratory capability, and the periodic analysis of laboratory reagent blanks, fortified blanks and other laboratory solutions as a continuing check on performance. The laboratory is required to maintain performance records that define the quality of the data that are generated.

9.2 INITIAL DEMONSTRATION OF PERFORMANCE

- 9.2.1 The initial demonstration of performance is used to characterize instrument performance (determination of linear calibration ranges and analysis of QCS) and laboratory performance (determination of MDLs) prior to performing analyses by this method.
- 9.2.2 Linear Calibration Range (LCR) -- The LCR must be determined initially and verified every 6 months or whenever a significant change

in instrument response is observed or expected. The initial demonstration of linearity must use sufficient standards to insure that the resulting curve is linear. The verification of linearity must use a minimum of a blank and three standards. If any verification data exceeds the initial values by $\pm 10\%$, linearity must be reestablished. If any portion of the range is shown to be nonlinear, sufficient standards must be used to clearly define the nonlinear portion.

- 9.2.3 Quality Control Sample (QCS) -- When beginning the use of this method, on a quarterly basis, or as required to meet data-quality needs, verify the calibration standards and acceptable instrument performance with the preparation and analyses of a QCS. If the determined concentrations are not within $\pm 10\%$ of the stated values, performance of the determinative step of the method is unacceptable. The source of the problem must be identified and corrected before either proceeding with the initial determination of MDLs or continuing with on-going analyses.
- 9.2.4 Method Detection Limit (MDL) -- MDLs must be established for all analytes, using reagent water (blank) fortified at a concentration of two to three times the estimated instrument detection limit. (6) To determine MDL values, take seven replicate aliquots of the fortified reagent water and process through the entire analytical method. Perform all calculations defined in the method and report the concentration values in the appropriate units. Calculate the MDL as follows:

$$MDL = (t) x (S)$$

where, t = Student's t value for a 99% confidence level and a standard deviation estimate with n-1 degrees of freedom [t = 3.14 for seven replicates]

S = standard deviation of the replicate analyses

MDLs should be determined every six months, when a new operator begins work, or whenever there is a significant change in the background or instrument response.

9.3 ASSESSING LABORATORY PERFORMANCE

9.3.1 Laboratory Reagent Blank (LRB) -- The laboratory must analyze at least one LRB with each batch of samples. Data produced are used to assess contamination from the laboratory environment. Values that exceed the MDL indicate laboratory or reagent contamination should be suspected and corrective actions must be taken before continuing the analysis.

- 9.3.2 Laboratory Fortified Blank (LFB) -- The laboratory must analyze at least one LFB with each batch of samples. Calculate accuracy as percent recovery (Section 9.4.2). If the recovery of any analyte falls outside the required control limits of 90-110%, that analyte is judged out of control, and the source of the problem should be identified and resolved before continuing analyses.
- 9.3.3 The laboratory must use LFB analyses data to assess laboratory performance against the required control limits of 90-110%. When sufficient internal performance data become available (usually a minimum of 20-30 analyses), optional control limits can be developed from the percent mean recovery (x) and the standard deviation (S) of the mean recovery. These data can be used to establish the upper and lower control limits as follows:

UPPER CONTROL LIMIT = x + 3SLOWER CONTROL LIMIT = x - 3S

The optional control limits must be equal to or better than the required control limits of 90-110%. After each five to ten new recovery measurements, new control limits can be calculated using only the most recent 20-30 data points. Also, the standard deviation (S) data should be used to establish an on-going precision statement for the level of concentrations included in the LFB. These data must be kept on file and be available for review.

9.3.4 Instrument Performance Check Solution (IPC) -- For all determinations the laboratory must analyze the IPC (a mid-range check standard) and a calibration blank immediately following daily calibration, after every 10th sample (or more frequently, if required), and at the end of the sample run. Analysis of the IPC solution and calibration blank immediately following calibration must verify that the instrument is within ±10% of calibration. Subsequent analyses of the IPC solution must verify the calibration is still within $\pm 10\%$. If the calibration cannot be verified within the specified limits, reanalyze the IPC solution. If the second analysis of the IPC solution confirms calibration to be outside the limits, sample analysis must be discontinued, the cause determined and/or in the case of drift the instrument recalibrated. All samples following the last acceptable IPC solution must be reanalyzed. The analysis data of the calibration blank and IPC solution must be kept on file with the sample analyses data.

9.4 ASSESSING ANALYTE RECOVERY AND DATA QUALITY

9.4.1 Laboratory Fortified Sample Matrix (LFM) -- The laboratory must add a known amount of analyte to a minimum of 10% of the routine samples. In each case the LFM aliquot must be a duplicate of the aliquot used for sample analysis. The analyte concentration must be high enough to

be detected above the original sample and should not be less than four times the MDL. The added analyte concentration should be the same as that used in the laboratory fortified blank.

9.4.2 Calculate the percent recovery for each analyte, corrected for concentrations measured in the unfortified sample, and compare these values to the designated LFM recovery range 90-110%. Percent recovery may be calculated using the following equation:

$$R = \frac{C_s - C}{s} \times 100$$

where, R = percent recovery

 C_s = fortified sample concentration

C = sample background concentration

s = concentration equivalent of analyte added to sample

- 9.4.3 If the recovery of any analyte falls outside the designated LFM recovery range and the laboratory performance for that analyte is shown to be in control (Section 9.3), the recovery problem encountered with the LFM is judged to be either matrix or solution related, not system related.
- 9.4.4 Where reference materials are available, they should be analyzed to provide additional performance data. The analysis of reference samples is a valuable tool for demonstrating the ability to perform the method acceptably.

10.0 CALIBRATION AND STANDARDIZATION

- 10.1 Prepare a series of at least three standards, covering the desired range, and a blank by diluting suitable volumes of standard solution (Section 7.11) with reagent water.
- 10.2 Process standards and blanks as described in Section 11.0, Procedure.
- 10.3 Set up manifold as shown in Figure 1 and Table 2.
- 10.4 Prepare flow system as described in Section 11.0, Procedure.
- 10.5 Place appropriate standards in the sampler in order of decreasing concentration and perform analysis.
- 10.6 Prepare standard curve by plotting instrument response against concentration values. A calibration curve may be fitted to the calibration solutions concentration/response data using computer or calculator based regression curve fitting techniques. Acceptance or control limits should be established

- using the difference between the measured value of the calibration solution and the "true value" concentration.
- 10.7 After the calibration has been established, it must be verified by the analysis of a suitable quality control sample (QCS). If measurements exceed $\pm 10\%$ of the established QCS value, the analysis should be terminated and the instrument recalibrated. The new calibration must be verified before continuing analysis. Periodic reanalysis of the QCS is recommended as a continuing calibration check.

11.0 PROCEDURE

- 11.1 Pipet 25.0 mL of sample, standard or blank in the digestor tube.
- 11.2 Add 5 mL of digestion solution (Section 7.3) and mix with a vortex mixer (See Note 1).
- 11.3 Add four to eight Teflon boiling chips (Section 7.12). **CAUTION:** An excess of Teflon chips may cause the sample to boil over.
- 11.4 Place tubes in block digestor preheated to 160°C and maintain temperature for one hour.
- 11.5 Reset temperature to 380°C and continue to heat for one and one half hour.

 (380°C MUST BE MAINTAINED FOR 30 MINUTES)
- 11.6 Remove digestion tubes, cool and dilute to 25 mL with reagent water.
- 11.7 Excluding the salicylate line, place all reagent lines in their respective containers, connect the sample probe to the sampler and start the pump.
- 11.8 Flush the sampler wash receptacle with about 25 mL of 4% sulfuric acid (Section 7.4) (See Note 2).
- 11.9 When reagents have been pumping for at least five minutes, place the salicylate line in its respective container and allow the system to equilibrate. If a precipitate forms after the addition of salicylate, the pH is too low. Immediately stop the proportioning pump and flush the coils with water using a syringe. Before restarting the system, check the concentration of the sulfuric acid solutions and/or the working buffer solution.
- 11.10 To prevent precipitation of sodium salicylate in the waste tray, which can clog the tray outlet, keep the nitrogen flowcell pump tube and the nitrogen Colorimeter "To Waste" tube separate from all other lines or keep tap water flowing in the waste tray.

11.11 After a stable baseline has been obtained, start the sampler and perform analysis.

12.0 DATA ANALYSIS AND CALCULATIONS

- 12.1 Prepare a calibration curve by plotting instrument response against standard concentration. Compute sample concentration by comparing sample response with the standard curve. Multiply answer by appropriate dilution factor.
- 12.2 Report only those values that fall between the lowest and the highest calibration standards. Samples exceeding the highest standard should be diluted and reanalyzed.
- 12.3 Report results in mg N/L.

13.0 METHOD PERFORMANCE

- 13.1 In a single laboratory (EMSL-Cincinnati) using sewage samples at concentrations of 1.2, 2.6, and 1.7 mg N/L, the precision was ± 0.07 , ± 0.03 , and ± 0.15 , respectively.
- 13.2 In a single laboratory (EMSL-Cincinnati) using sewage samples at concentrations 4.7 and 8.74 mg N/L, the recoveries were 99% and 99%, respectively.
- 13.3 The interlaboratory precision and accuracy data in Table 1 were developed using a reagent water matrix. Values are in mg N/L.

14.0 POLLUTION PREVENTION

- 14.1 Pollution prevention encompasses any technique that reduces or eliminates the quantity or toxicity of waste at the point of generation. Numerous opportunities for pollution prevention exist in laboratory operation. The EPA has established a preferred hierarchy of environmental management techniques that places pollution prevention as the management option of first choice. Whenever feasible, laboratory personnel should use pollution prevention techniques to address their waste generation. When wastes cannot be feasibly reduced at the source, the Agency recommends recycling as the next best option.
- 14.2 The quantity of chemicals purchased should be based on expected usage during its shelf life and disposal cost of unused material. Actual reagent preparation volumes should reflect anticipated usage and reagent stability.
- 14.3 For information about pollution prevention that may be applicable to laboratories and research institutions, consult "Less is Better: Laboratory Chemical Management for Waste Reduction", available from the American

Chemical Society's Department of Government Regulations and Science Policy, 1155 16th Street N.W., Washington, D.C. 20036, (202) 872-4477.

15.0 WASTE MANAGEMENT

15.1 The Environmental Protection Agency requires that laboratory waste management practices be conducted consistent with all applicable rules and regulations. Excess Reagents and samples and method process wastes should be characterized and disposed of in an acceptable manner. The Agency urges laboratories to protect the air, water, and land by minimizing and controlling all releases from hoods and bench operations, complying with the letter and spirit of any waste discharge permit and regulations, and by complying with all solid and hazardous waste regulations, particularly the hazardous waste identification rules and land disposal restrictions. For further information on waste management consult "The Waste Management Manual for Laboratory Personnel", available from the American Chemical Society at the address listed in Section 14.3.

16.0 REFERENCES

- 1. McDaniel, W.H., Hemphill, R.N. and Donaldson, W.T., "Automatic Determination of total Kjeldahl Nitrogen in Estuarine Water", Technicon Symposia, pp. 362-367, Vol. 1, 1967.
- 2. Gales, M.E. and Booth, R.L., "Evaluation of Organic Nitrogen Methods", EPA Office of Research and Monitoring, June, 1972.
- Gales, M.E. and Booth, R.L., "Simultaneous and Automated Determination of Total Phosphorus and Total Kjeldahl Nitrogen", Methods Development and Quality Assurance Research Laboratory, May 1974.
- 4. Technicon "Total Kjeldahl Nitrogen and Total Phosphorus BD-40 Digestion Procedure for Water", August 1974.
- 5. Gales, M.E., and Booth, R.L., "Evaluation of the Technicon Block Digestor System for the Measurement of Total Kjeldahl Nitrogen and Total Phosphorus", EPA-600/4-78-015, Environmental Monitoring and Support Laboratory, Cincinnati, Ohio, 1978.
- 6. Code of Federal Regulations 40, Ch. 1, Pt. 136, Appendix B.

17.0 TABLES, DIAGRAMS, FLOWCHARTS, AND VALIDATION DATA

TABLE 1. INTERLABORATORY PRECISION AND ACCURACY DATA

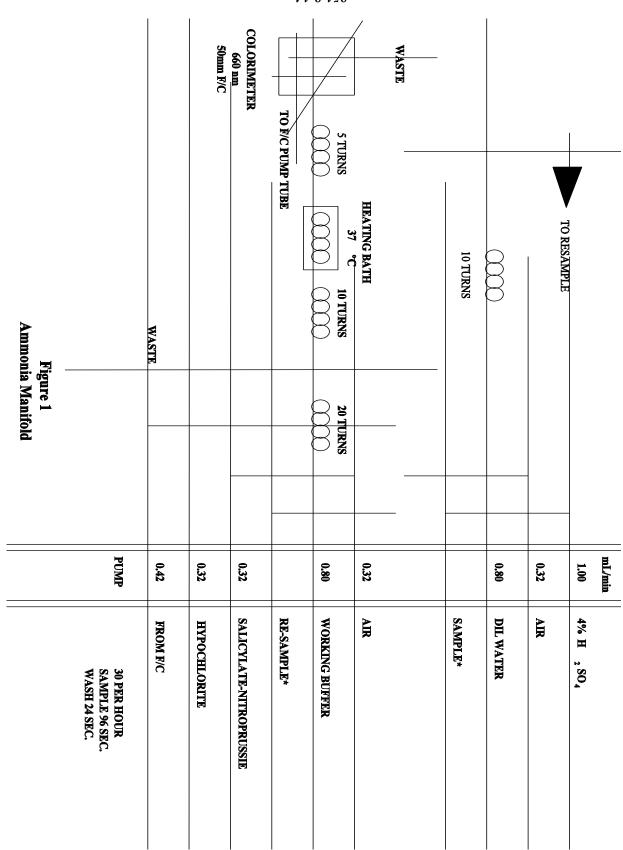
Number of Values Reported	True Value (T)	Mean (X)	Residual for X	Standard Deviation (S)	Residual for S
115	0.380	0.3891	-0.0091	0.0750	-0.0135
134	0.451	0.4807	0.0125	0.1181	0.0238
127	1.00	1.0095	-0.0000	0.1170	-0.0227
164	3.10	3.0992	0.0191	0.2821	-0.0310
138	3.50	3.4765	0.0020	0.3973	0.0512
115	5.71	5.6083	-0.0452	0.4869	-0.0417
175	7.00	6.9246	-0.0008	0.6623	0.0272
121	8.00	7.9991	0.0877	0.6283	-0.0894
120	15.0	15.0213	0.2080	1.2495	-0.0462
127	21.0	20.4355	-0.2937	1.7267	-0.0644
164	25.0	24.7157	0.0426	2.0147	-0.1067
175	26.9	26.1464	-0.4000	2.9743	0.6960

REGRESSIONS: X = 0.986T + 0.024, S = 0.083T + 0.057

TABLE 2. CONCENTRATION RANGES

Pango	Pump	Pump mL/min.			
Range mg/LN	Sample	Resample	Buffer (Section 7.7)		
0-1.5	0.80	0.32	250		
0-5.0	0.16	0.32	120		
0-10.0	0.16	0.16	80		

351.2-14





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Technical Note 14

Total Dissolved Solids from conductivity

Glenn Carlson, Technical Support, In-Situ Inc. May 26, 2005

Total Solids is the term applied to material residue left in a vessel after evaporation of a water sample and subsequent drying of the residue. Total Solids includes Total Suspended Solids (TSS), the portion of total solids in a sample that can be retained by a filter, and Total Dissolved Solids (TDS), the portion that passes through a filter.

The amount of dissolved material in a sample correlates to electrical conductivity. TDS values reported by Win-Situ software are derived from conductivity readings. This calculation, as with other calculations in Win-Situ, is per the 20th edition of *Standard Methods for the Examination of Water and Wastewater*. It should only be used as a rough field check of a sample, though. TDS derived from conductivity is not recommended for critical quantitative reporting purposes. The reason for this is that there is not a relationship between conductivity and TDS that is very repeatable across different locations and different dissolved material. The calculation used is:

TDS = SC * 0.65

where:

TDS = Total Dissolved Solids in mg/LSC = Specific Conductance (temperature corrected) in uS/cm

The constant of 0.65 is only a VERY crude average for natural samples. The actual constant for any particular sample with a specific mix of dissolved materials and measurement temperature can vary widely. The actual multiplier necessary depends on the activity of each specific dissolved species present and the average activity of all species in a sample. These activities are influenced by sample temperature, the relative amounts of each species (they can influence each other) and the total concentration of dissolved

solids in the sample (can be a non-linear relationship).

While the default average value for this calculation can give good results for some samples, this calculation from conductivity only represents a very crude index for other samples and should not be used as the sole method to accurately quantify the actual amount of dissolved material in a sample. If measurements will always be made at the same location, then it would be far better practice to determine the actual constant that would be appropriate for those samples and then manually do the TDS calculation from Specific Conductance. Ideally, measuring TDS of preliminary samples gravimetrically and regressing those results against the measured Specific Conductance of the samples would determine the constant.

The composition of dissolved material in samples will certainly change from one site to another. Even for the same site, however, the type of dissolved material may also change over time. If the composition of dissolved solids changes appreciably, then it will be necessary to again determine a new constant for the site gravimetrically.

For more information contact In-Situ Inc.

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5.9 Conductivity

What is conductivity and why is it important?

Conductivity is a measure of the ability of water to pass an electrical current. Conductivity in water is affected by the presence of inorganic dissolved solids such as chloride, nitrate, sulfate, and phosphate anions (ions that carry a negative charge) or sodium, magnesium, calcium, iron, and aluminum cations (ions that carry a positive charge). Organic compounds like oil, phenol, alcohol, and sugar do not conduct electrical current very well and therefore have a low conductivity when in water. Conductivity is also affected by temperature: the warmer the water, the higher the conductivity. For this reason, conductivity is reported as conductivity at 25 degrees Celsius (25 C).

Conductivity in streams and rivers is affected primarily by the geology of the area through which the water flows. Streams that run through areas with granite bedrock tend to have lower conductivity because granite is composed of more inert materials that do not ionize (dissolve into ionic components) when washed into the water. On the other hand, streams that run through areas with clay soils tend to have higher conductivity because of the presence of materials that ionize when washed into the water. Ground water inflows can have the same effects depending on the bedrock they flow through.

Discharges to streams can change the conductivity depending on their make-up. A failing sewage system would raise the conductivity because of the presence of chloride, phosphate, and nitrate; an oil spill would lower the conductivity.

The basic unit of measurement of conductivity is the mho or siemens. Conductivity is measured in micromhos per centimeter (μ mhos/cm) or microsiemens per centimeter (μ s/cm). Distilled water has a conductivity in the range of 0.5 to 3 μ mhos/cm. The conductivity of rivers in the United States generally ranges from 50 to 1500 μ mhos/cm. Studies of inland fresh waters indicate that streams supporting good mixed fisheries have a range between 150 and 500 μ hos/cm. Conductivity outside this range could indicate that the water is not suitable for certain species of fish or macroinvertebrates. Industrial waters can range as high as 10,000 μ mhos/cm.

Sampling and equipment Considerations

Conductivity is useful as a general measure of stream water quality. Each stream tends to have a relatively constant range of conductivity that, once established, can be used as a baseline for



comparison with regular conductivity measurements. Significant changes in conductivity could then be an indicator that a discharge or some other source of pollution has entered a stream.

Conductivity is measured with a probe and a meter. Voltage is applied between two electrodes in a probe immersed in the sample water. The drop in voltage caused by the resistance of the water is used to calculate the conductivity per centimeter. The meter converts the probe measurement to micromhos per centimeter and displays the result for the user. NOTE: Some conductivity meters can also be used to test for total dissolved solids and salinity. The total dissolved solids concentration in milligrams per liter (mg/L) can also be calculated by multiplying the conductivity result by a factor between 0.55 and 0.9, which is empirically determined (see Standard Methods #2510, APHA 1992).

Suitable conductivity meters cost about \$350. Meters in this price range should also measure temperature and automatically compensate for temperature in the conductivity reading. Conductivity can be measured in the field or the lab. In most cases, it is probably better if the samples are collected in the field and taken to a lab for testing. In this way several teams of volunteers can collect samples simultaneously. If it is important to test in the field, meters designed for field use can be obtained for around the same cost mentioned above.

If samples will be collected in the field for later measurement, the sample bottle should be a glass or polyethylene bottle that has been washed in phosphate-free detergent and rinsed thoroughly with both tap and distilled water. Factory-prepared Whirl-pak® bags may be used.

How to sample

The procedures for collecting samples and analyzing conductivity consist of the following tasks:

TASK 1 Prepare the sample containers

If factory-sealed, disposable Whirl-pak® bags are used for sampling, no preparation is needed. Reused sample containers (and all glassware used in this procedure) must be cleaned before the first run and after each sampling run by following Method A as described in MEthod A in Table 1 in Chapter 5 - Water Quality Conditions.

TASK 2 Prepare before leaving for the sampling site

Refer to <u>section 2.3 – Safety Considerations</u> for details on confirming sampling date and time, safety considerations, checking supplies, and checking weather and directions. In addition to the standard sampling equipment and apparel, when sampling for conductivity, include the following equipment:



- Conductivity meter and probe (if testing conductivity in the field)
- Conductivity standard appropriate for the range typical of the stream
- Data sheet for conductivity to record results

Be sure to let someone know where you are going and when you expect to return.

TASK 3 Collect the sample (if samples will be tested in the lab)

Refer to Task 2 in <u>Chapter 5 – Water Quality Conditions</u> for details on how to collect water samples using screw-cap bottles or Whirl-pak® bags.

TASK 4 Analyze the sample (field or lab)

The following procedure applies to field or lab use of the conductivity meter.

- 1. Prepare the conductivity meter for use according to the manufacturer's directions.
- 2. Use a conductivity standard solution (usually potassium chloride or sodium chloride) to calibrate the meter for the range that you will be measuring. The manufacturer's directions should describe the preparation procedures for the standard solutio n.
- 3. Rinse the probe with distilled or deionized water.
- 4. Select the appropriate range beginning with the highest range and working down. Read the conductivity of the water sample. If the reading is in the lower 10 percent of the range, switch to the next lower range. If the conductivity of the sample ex ceeds the range of the instrument, you may dilute the sample. Be sure to perform the dilution according to the manufacturer's directions because the dilution might not have a simple linear relationship to the conductivity.
- 5. Rinse the probe with distilled or deionized water and repeat step 4 until finished.

TASK 5 Return the samples and the field data sheets to the lab/drop-off point.

Samples that are sent to a lab for conductivity analysis must be tested within 28 days of collection. Keep the samples on ice or refrigerated.

References

APHA. 1992. *Standard methods for the examination of water and wastewater.* 18th ed. American Public Health Association, Washington, DC.



Hach Company. 1992. Hach water analysis handbook. 2nd ed. Loveland, CO.

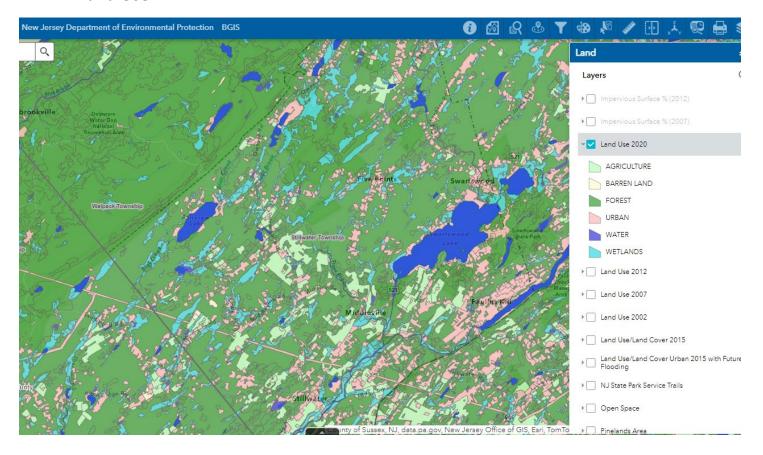
Mississippi Headwaters River Watch. 1991. *Water quality procedures.* Mississippi Headwaters Board. March.

Source: https://archive.epa.gov/water/archive/web/html/vms59.html (accessed March 23, 2024)

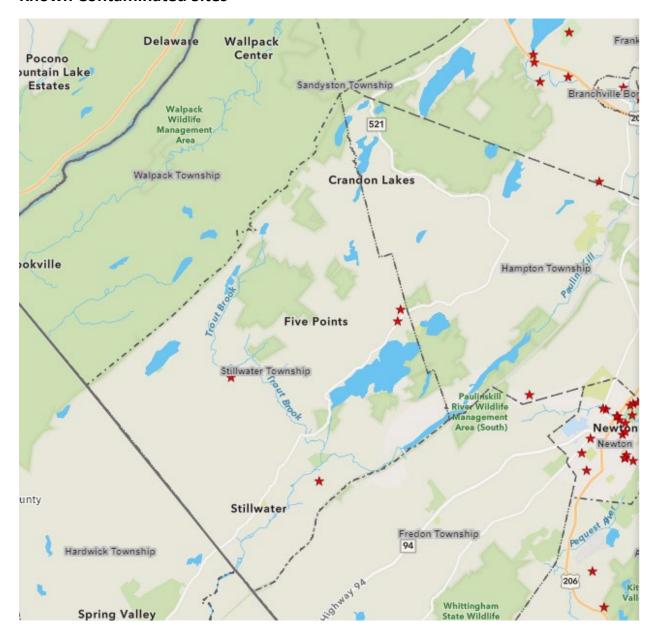
Appendix C

NJDEP GeoWeb Map Information Excerpts

Land Use

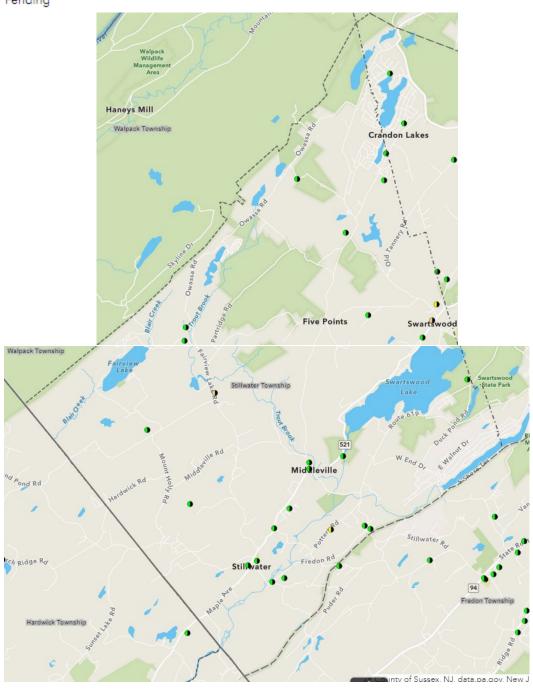


Known Contaminated Sites



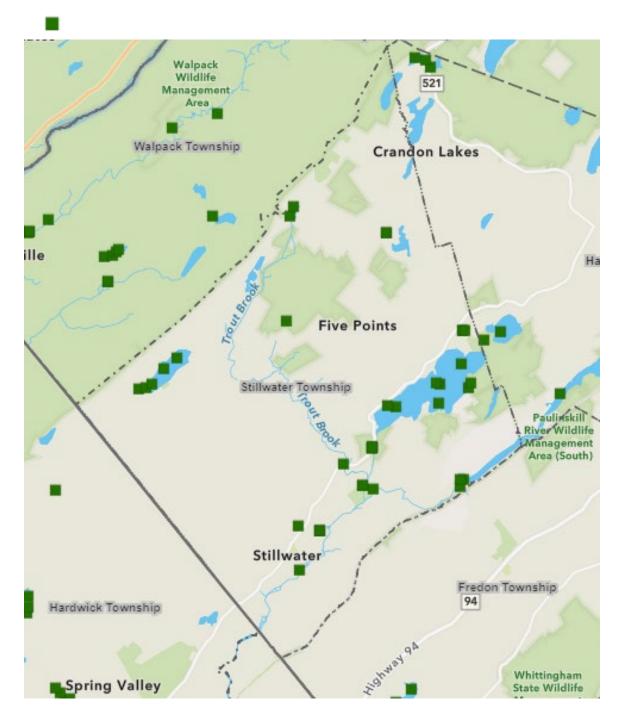
Site Remediation Preferred ID Sites

- ▼ ✓ Site Remediation Program Preferred ID Sites
 - Active
 - Active UHOT
 - RAP\Post Remedial
 - Closed
 - Pending



NJDEP WQDE Sample Stations

▼ ✓ Water Quality Data Exchange (WQDE)



Appendix D

Summary of 95UCL and Statistical Trend Analysis Results

Appendix D Summary of SWMP 95UCL and Trend Analysis Results Surface Water Monitorng Program (1984-2024) - Stillwater, NJ

Parameter	Station #	P-value	Signifcant Trend?	Trend Direction	95 UCL*	Units	Sample Size (n)
Alkalinity	1	0.254	No	No Trend	20.8	mg/L	26
Alkalinity	2	0.474	No	No Trend	14.8	mg/L	26
Alkalinity	3	0.056	No	No Trend	25.4	mg/L	26
Alkalinity	4	0.233	No	No Trend	22.4	mg/L	26
Alkalinity	5	0.005	Yes	Increasing	35.5	mg/L	25
Alkalinity	6	0.034	Yes	No Trend	31.9	mg/L	26
Alkalinity	7	0.362	No	No Trend	137.1	mg/L	26
Alkalinity	8	0.175	No	No Trend	77.3	mg/L	16
Alkalinity	9	0.242	No	No Trend	149.3	mg/L	10
Aluminum	1	0.184	No	No Trend	0.143	mg/L	18
Aluminum	2	0.048	Yes	Decreasing	0.139	mg/L	18
Aluminum	3	0.041	Yes	Decreasing	0.149	mg/L	18
Aluminum	4	0.048	Yes	Decreasing	0.184	mg/L	18
Aluminum	5	0.026	Yes	Decreasing	0.265	mg/L	16
Aluminum	6	0.054	No	No Trend	0.086	mg/L	17
Aluminum	7	0.038	Yes	Decreasing	0.161	mg/L	17
Aluminum	8	0.460	No	No Trend	N/A	mg/L	9
Aluminum	9	N/A		N/A	0.163	mg/L	3
Ammonia	1	0.003	Yes	Decreasing	0.174	mg/L	25
Ammonia	2	0.002	Yes	Decreasing	0.227	mg/L	25
Ammonia	3	0.005	Yes	Decreasing	0.134	mg/L	25
Ammonia	4	0.001	Yes	Decreasing	0.155	mg/L	25
Ammonia	5	0.003	Yes	Decreasing	0.118	mg/L	25
Ammonia	6	0.005	Yes	Decreasing	0.110	mg/L	25
Ammonia	7	0.002	Yes	Decreasing	0.114	mg/L	25
Ammonia	8	0.385	No	No Trend	0.073	mg/L	15
Ammonia	9	0.460	No	No Trend	0.151	mg/L	9
E. Coli	10	0.500	No	No Trend	370.1	col/100ml	
Fecal Coliforms	1	0.121	No	No Trend	59.7	col/100ml	
Fecal Coliforms	2	0.217	No	No Trend	56.8	col/100ml	
Fecal Coliforms	3	0.226	No	No Trend	35.5	col/100ml	
Fecal Coliforms	4	0.007	Yes	Decreasing	19.0	col/100ml	
Fecal Coliforms	5	0.017	Yes	Decreasing	84.2	col/100ml	
Fecal Coliforms	6	0.168	No	No Trend	31.1	col/100ml	25
Fecal Coliforms	7	0.023	Yes	Decreasing	180.8	col/100ml	25
Fecal Coliforms	8	0.225	No	No Trend	24.2	col/100ml	16
Fecal Coliforms	9	0.431	No	No Trend	158.4	col/100ml	10
Fecal Coliforms	10	0.281	No	No Trend	354.6	col/100ml	7
Lead	1	0.034	Yes	Decreasing	0.015	mg/L	19
Lead	2	0.093	No	No Trend	0.015	mg/L	19
Lead	3	0.105	No	No Trend	0.014	mg/L	
Lead	4	0.105	No	No Trend	0.014	mg/L	19
Lead	5	0.010	Yes	Decreasing	0.015	mg/L	
Lead	6	0.082	No	No Trend	0.015	mg/L	19
Lead	7	0.005	Yes	Decreasing	0.023	mg/L	
Lead	8	0.130	No	No Trend	0.004	mg/L	9
Lead	9	N/A	N/A	N/A	N/A	mg/L	
Nitrate	1	0.012	Yes	Decreasing	0.199	mg/L	

Nitroto	2	0.012	Voc	Dographing	0.232	ma/l	26
Nitrate	2		Yes	Decreasing No Trend	1	mg/L	26
Nitrate	3	0.131	No		0.167	mg/L	26
Nitrate	4	0.295	No	No Trend	0.217	mg/L	26
Nitrate	5	0.121	No	No Trend	0.231	mg/L	26
Nitrate	6	0.066	No	No Trend	0.199	mg/L	26
Nitrate	7	0.430	No	No Trend	0.636	mg/L	26
Nitrate	8	0.199	No	No Trend	0.201	mg/L	16
Nitrate	9	0.364	No	No Trend	0.508	mg/L	10
Nitrite	1	0.093	No	No Trend	0.037	mg/L	25
Nitrite	2	0.259	No	No Trend	0.035	mg/L	25
Nitrite	3	0.267	No	No Trend	0.035	mg/L	25
Nitrite	4	0.195	No	No Trend	0.047	mg/L	25
Nitrite	5	0.275	No	No Trend	0.035	mg/L	25
Nitrite	6	0.202	No	No Trend	0.036	mg/L	25
Nitrite	7	0.060	No	No Trend	0.039	mg/L	24
Nitrite	8	0.423	No	No Trend	0.047	mg/L	15
Nitrite	9	0.000	Yes	No Trend	0.096	mg/L	9
Orthophosphate	1	0.041	Yes	Decreasing	0.038	mg/L	26
Orthophosphate	2	0.033	Yes	Decreasing	0.036	mg/L	26
Orthophosphate	3	0.094	No	No Trend	0.033	mg/L	26
Orthophosphate	4	0.030	Yes	Decreasing	0.037	mg/L	26
Orthophosphate	5	0.029	Yes	No Trend	0.034	mg/L	25
Orthophosphate	6	0.049	Yes	Decreasing	0.033	mg/L	26
Orthophosphate	7	0.023	Yes	Decreasing	0.038	mg/L	25
Orthophosphate	8	0.345	No	No Trend	0.038	mg/L	16
Orthophosphate	9	0.242	No	No Trend	0.054	mg/L	10
рН	1	0.330	No	No Trend	6.90	su	26
pH	2	0.166	No	No Trend	6.77	su	26
pH	3	0.421	No	No Trend	7.20	su	26
pH	4	0.370		No Trend	7.08	su	26
pH	5	0.249	No	No Trend	7.23	su	25
pH	6	0.195	No	No Trend	7.26	su	26
pH	7	0.338	No	No Trend	8.02	su	26
pH	8	0.253	No	No Trend	7.76	su	16
pH	9	0.190	No	No Trend	8.04	su	10
Phosphorus	1	0.086		No Trend	0.065	mg/L	25
Phosphorus	2	0.234	No	No Trend	0.104	mg/L	25
Phosphorus	3	0.234	Yes	Decreasing	0.104	mg/L	25
Phosphorus	4	0.044	Yes	Decreasing	0.070	mg/L	25
Phosphorus	5	0.033		No Trend	0.061	mg/L	24
	6	0.178	No	No Trend	0.062		25
Phosphorus Phosphorus	7	0.241	Yes		0.060	mg/L	25
Phosphorus				Decreasing	1	mg/L	
Phosphorus	8	0.349	No	No Trend	0.091	mg/L	15
Phosphorus	9	0.460		No Trend	0.134	mg/L	9
Specific conductivity	1	0.261	No	No Trend	118.2	umohs/cm	26
Specific conductivity	2	0.183		No Trend	130.5	umohs/cm	26
Specific conductivity	3	0.330		No Trend	134.0	umohs/cm	26
Specific conductivity	4	0.076		No Trend	131.7	umohs/cm	26
Specific conductivity	5	0.000	Yes	Increasing	201.1	umohs/cm	25
Specific conductivity	6	0.003	Yes	Increasing	167.8	umohs/cm	26
Specific conductivity	7	0.100	No	No Trend	455.6	umohs/cm	26
Specific conductivity	8	0.199	No	No Trend	245.3	umohs/cm	16
Specific conductivity	9	0.078	No	No Trend	553.5	umohs/cm	10
TKN	1	0.395	No	No Trend	0.856	mg/L	23

TKN	2	0.110	No	No Trend	2.832	mg/L	24
TKN	3	0.263	No	No Trend	0.803	mg/L	24
TKN	4	0.231	No	No Trend	0.520	mg/L	24
TKN	5	0.162	No	No Trend	0.782	mg/L	24
TKN	6	0.264	No	No Trend	0.723	mg/L	24
TKN	7	0.226	No	No Trend	0.853	mg/L	24
TKN	8	0.225	No	No Trend	1.033	mg/L	14
TKN	9	0.238	No	No Trend	0.565	mg/L	9
TSS	1	0.424	No	No Trend	6.58	mg/L	25
TSS	2	0.032	Yes	Decreasing	4.01	mg/L	25
TSS	3	0.015	Yes	Decreasing	2.82	mg/L	25
TSS	4	0.005	Yes	Decreasing	4.26	mg/L	25
TSS	5	0.109	No	Decreasing	9.44	mg/L	25
TSS	6	0.006	Yes	Decreasing	3.36	mg/L	25
TSS	7	0.453	No	No Trend	6.61	mg/L	25
TSS	8	0.218	No	No Trend	16.87	mg/L	15
TSS	9	0.540	No	No Trend	15.92	mg/L	9

Notes:

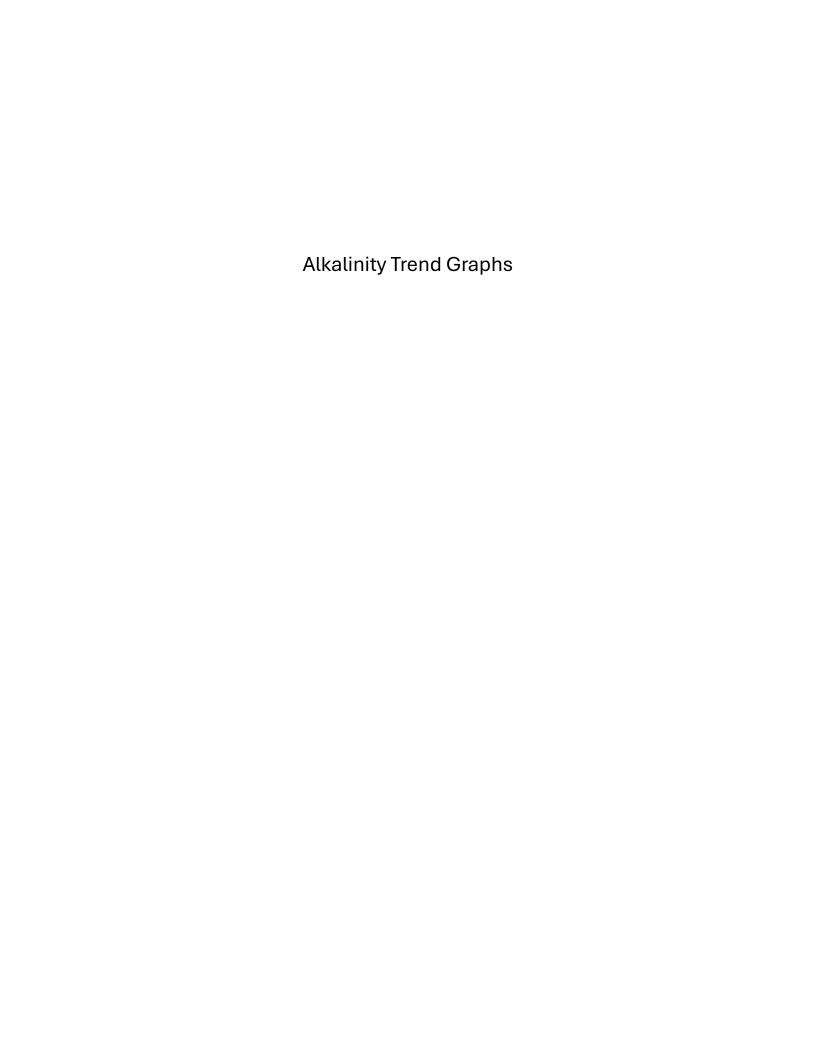
N/A = not applicable; 95UCL not calculated due to small sample size (n).

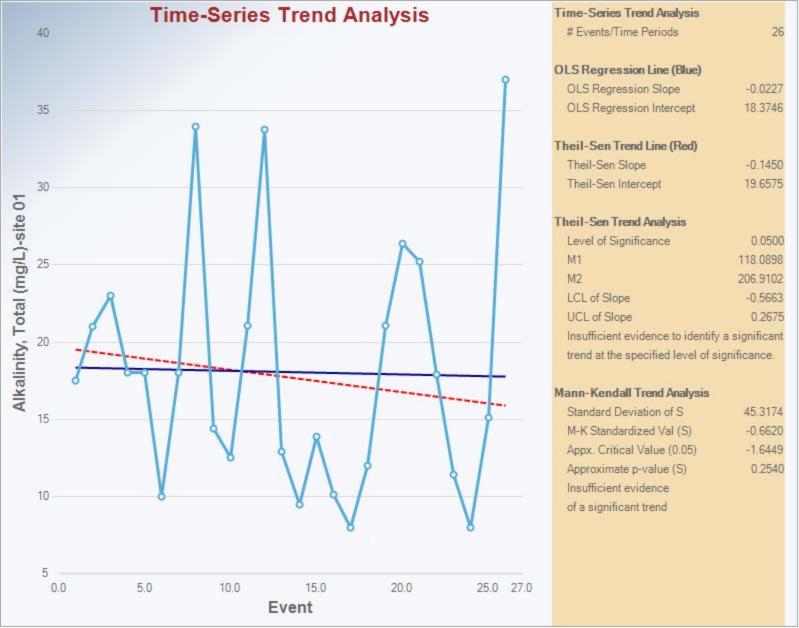
ND = Not Detected or not detected above SWCS; 95UCL not calculated due to low detection frequency.

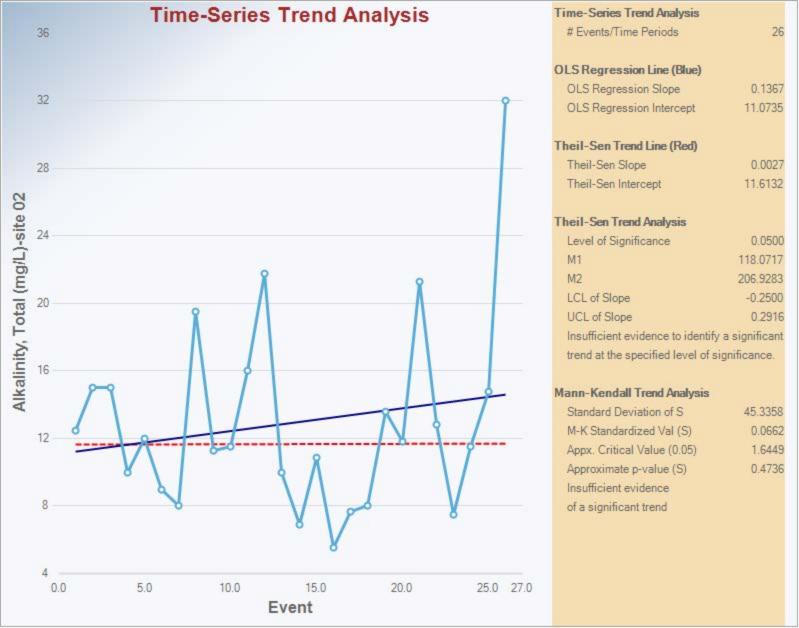
Significant trend identified if P>0.05.

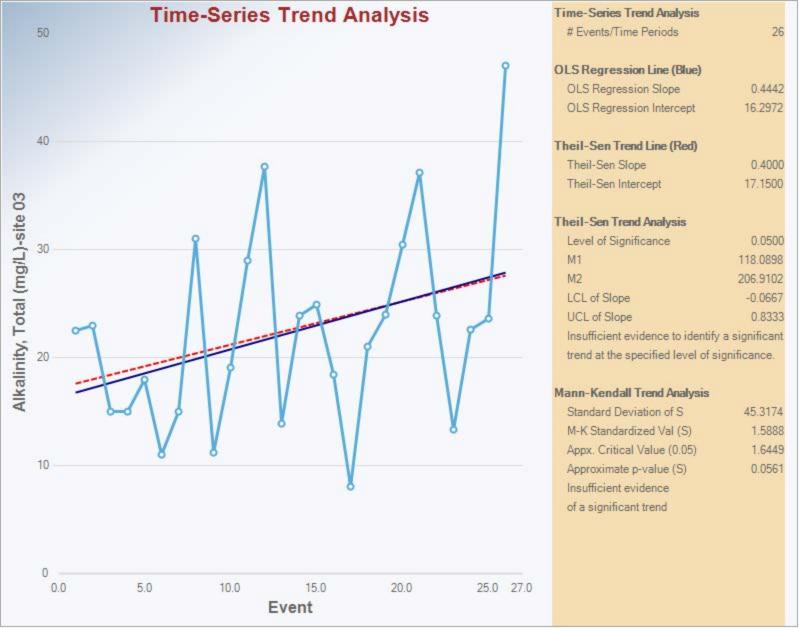
95UCL based on 95% Student's-t UCL.

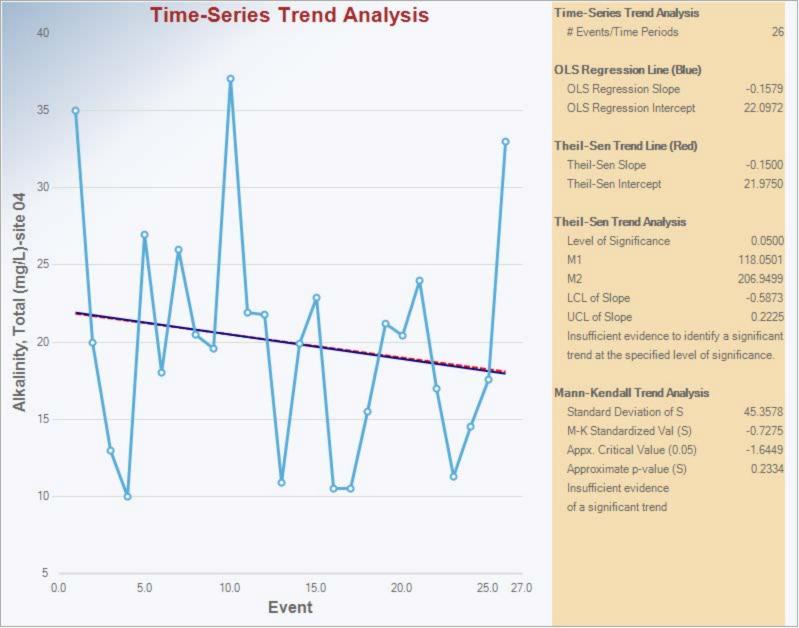
 $^{^\}star$ 95UCL qulaified due to low sample size (n<10)

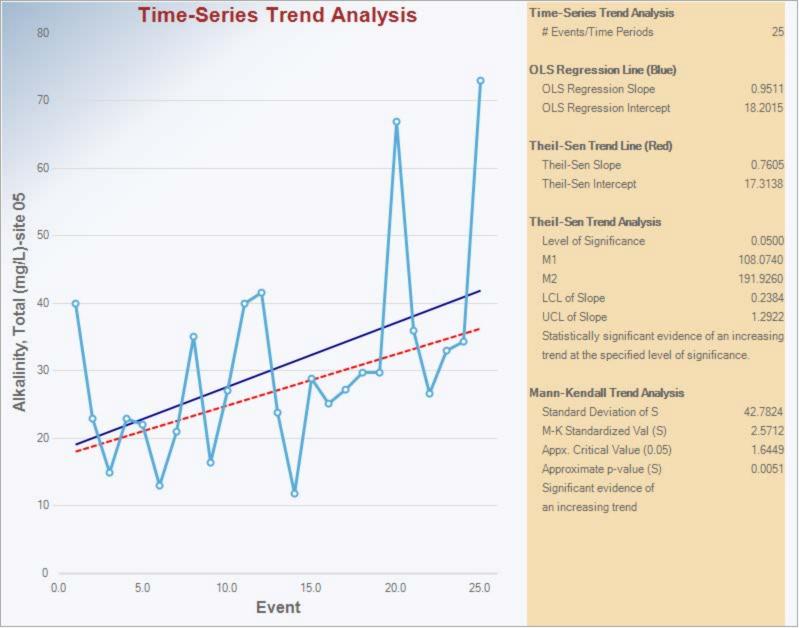


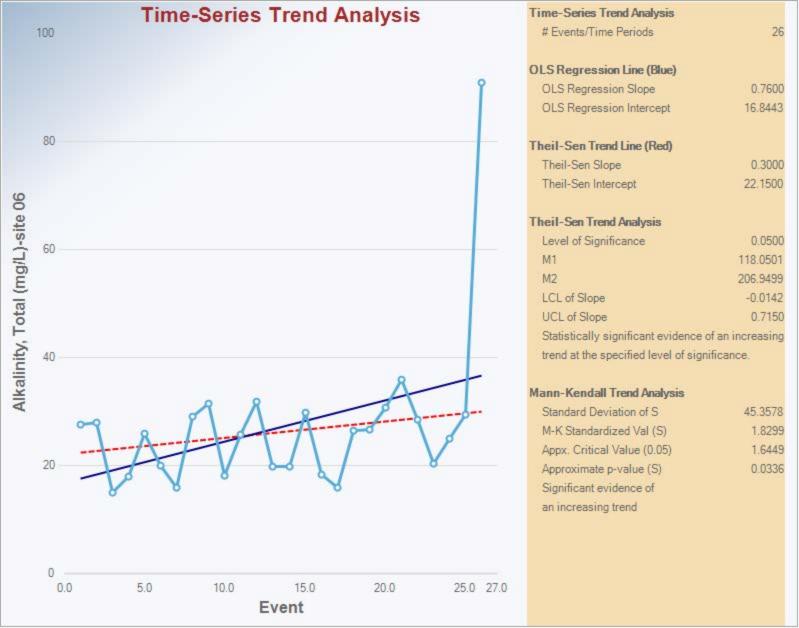


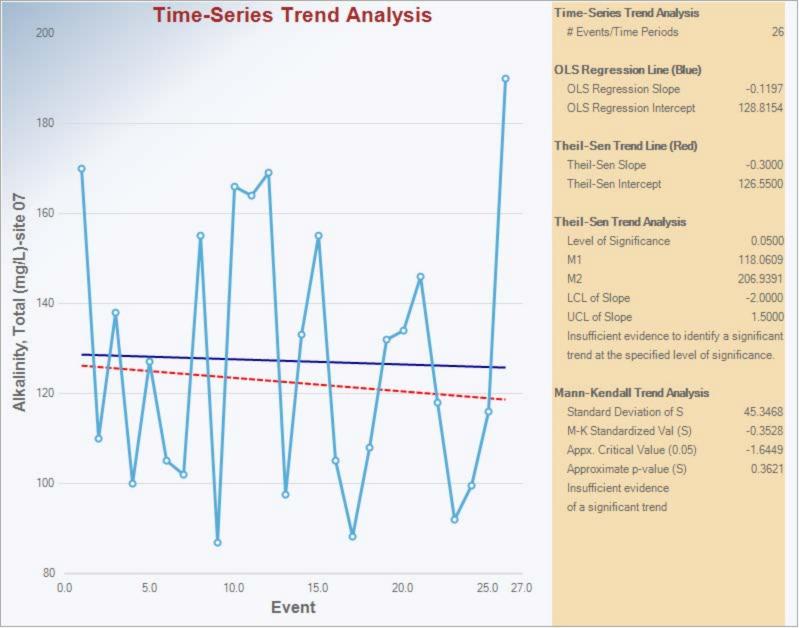


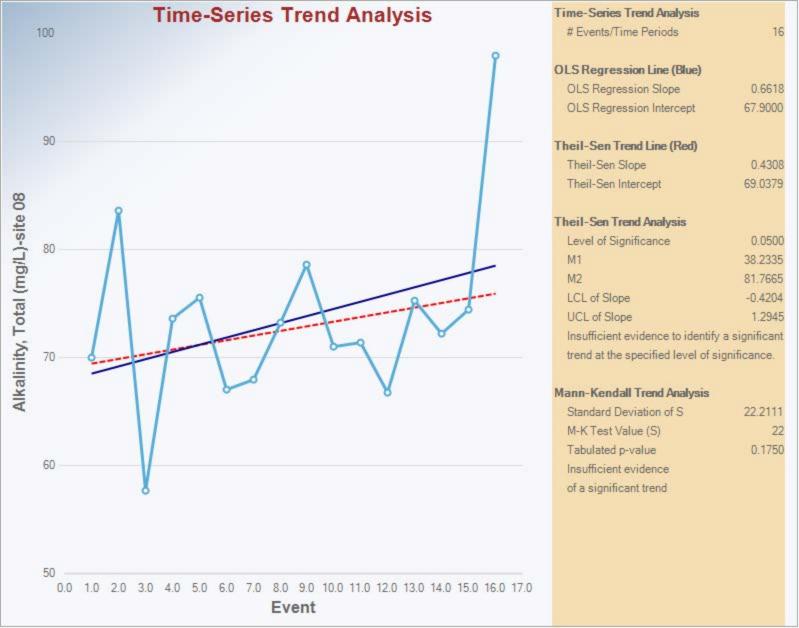


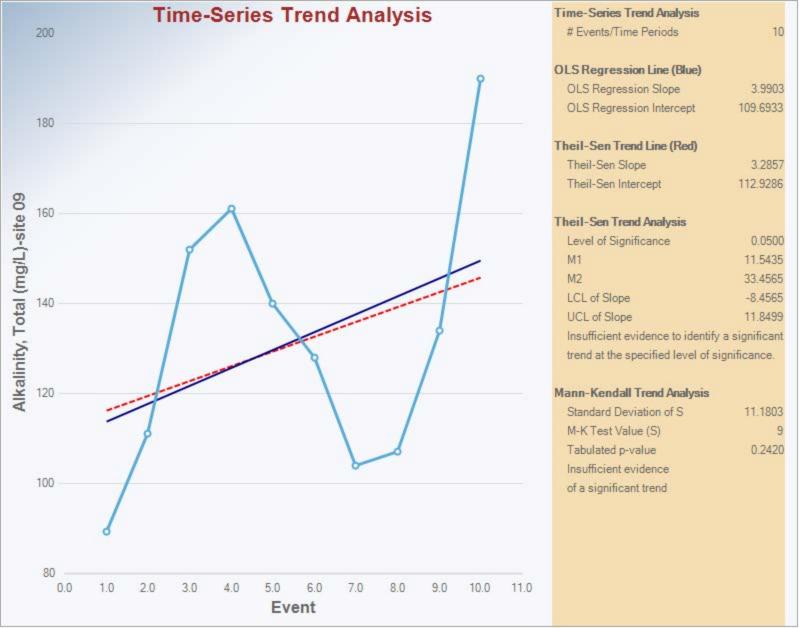










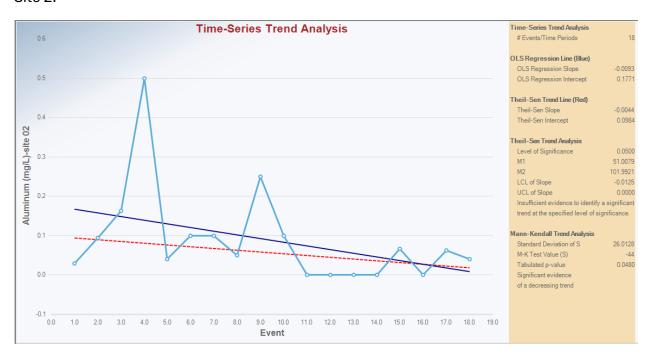


Aluminum graphs

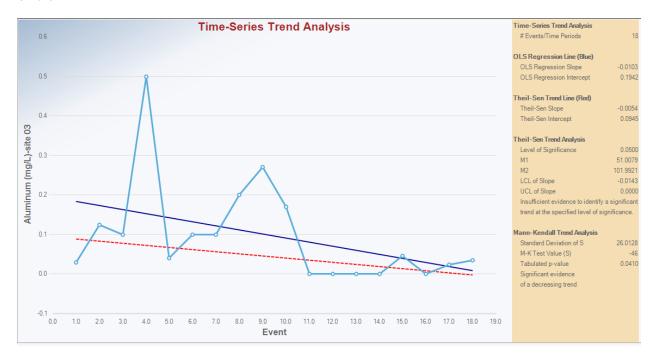
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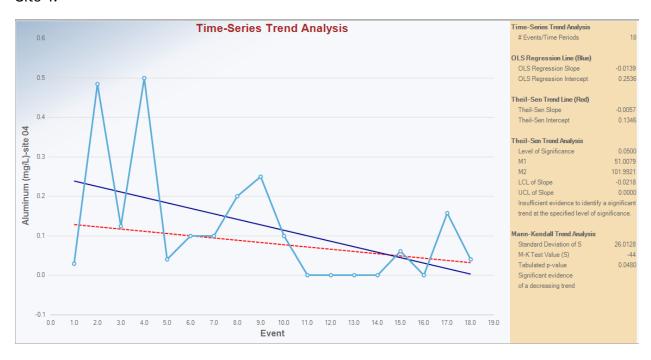
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Site 3:



Site 4:



Site 5:



Site 6:



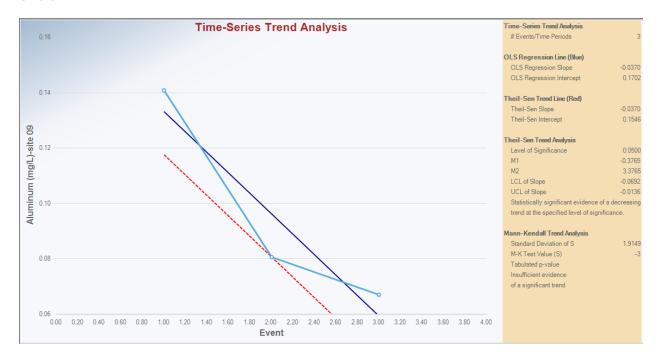
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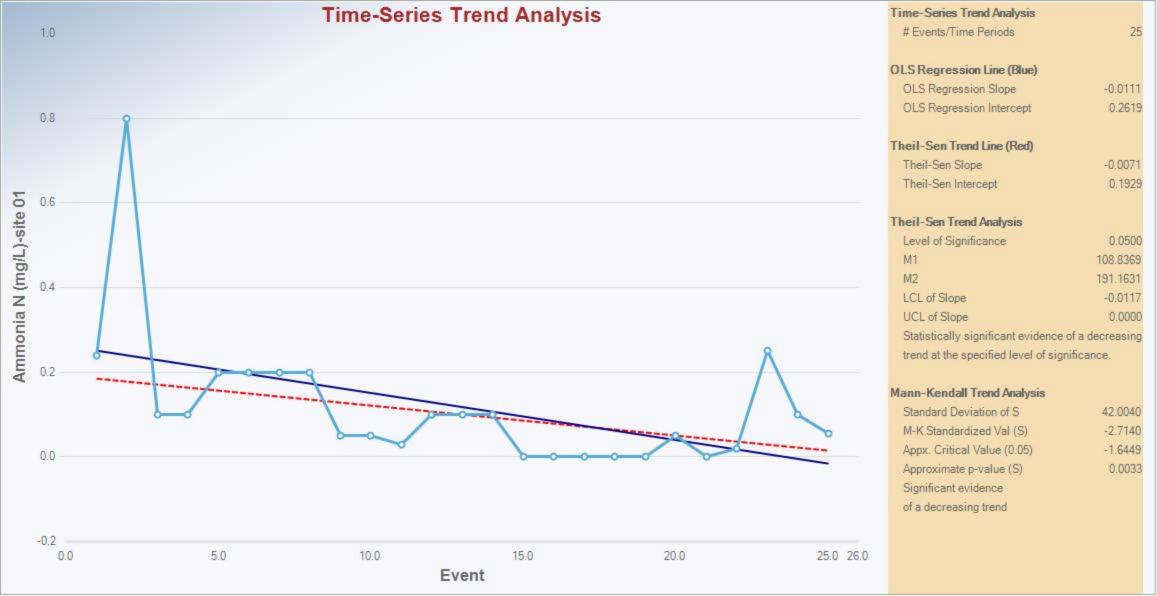
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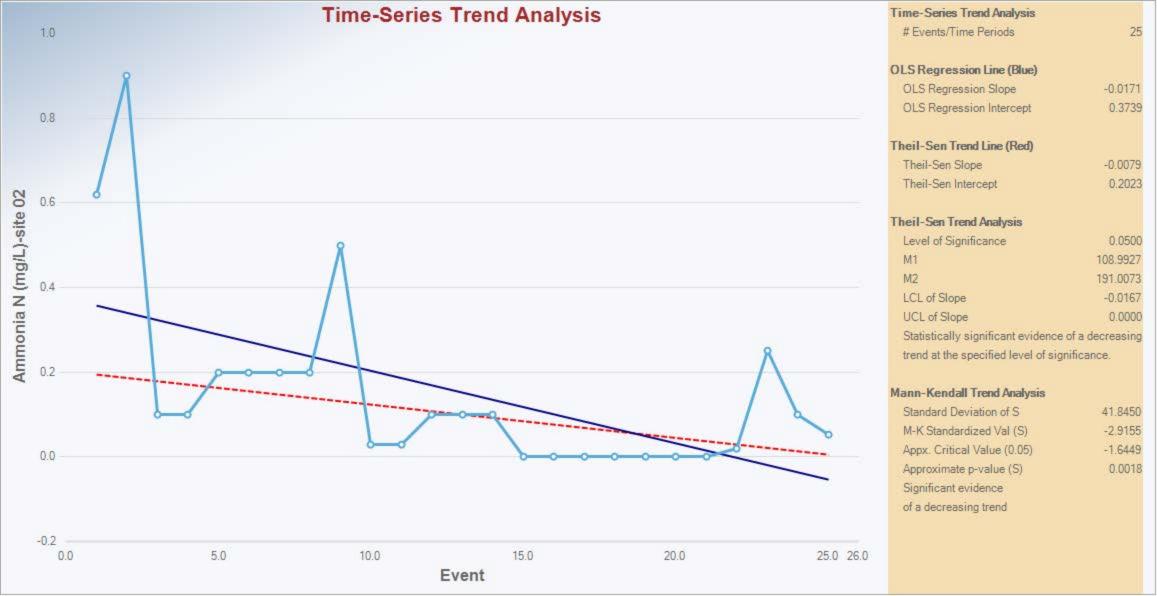


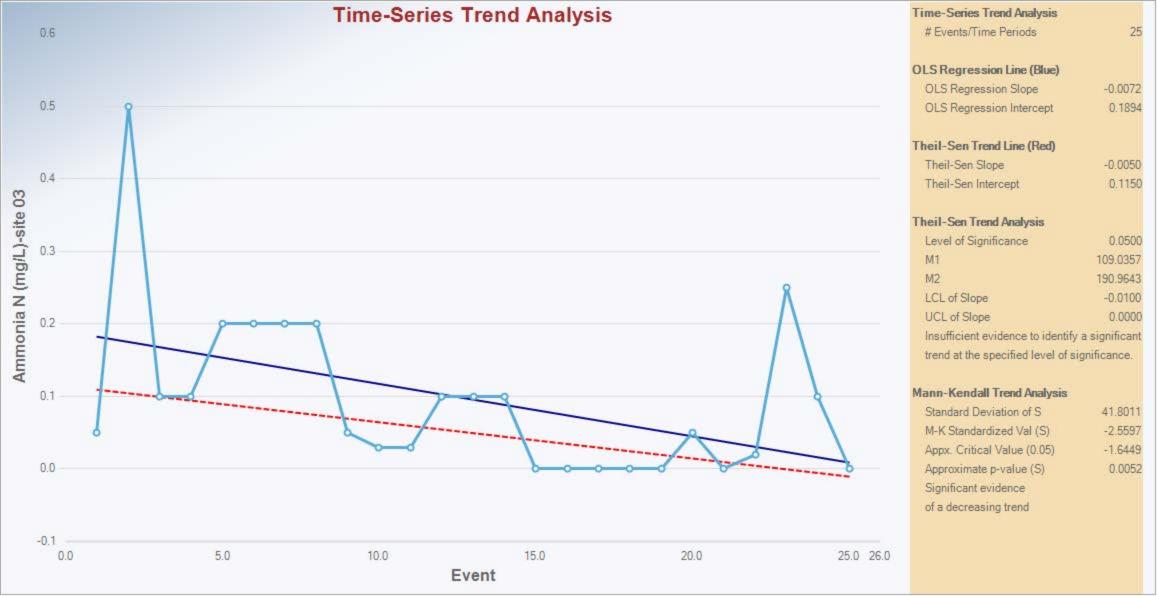
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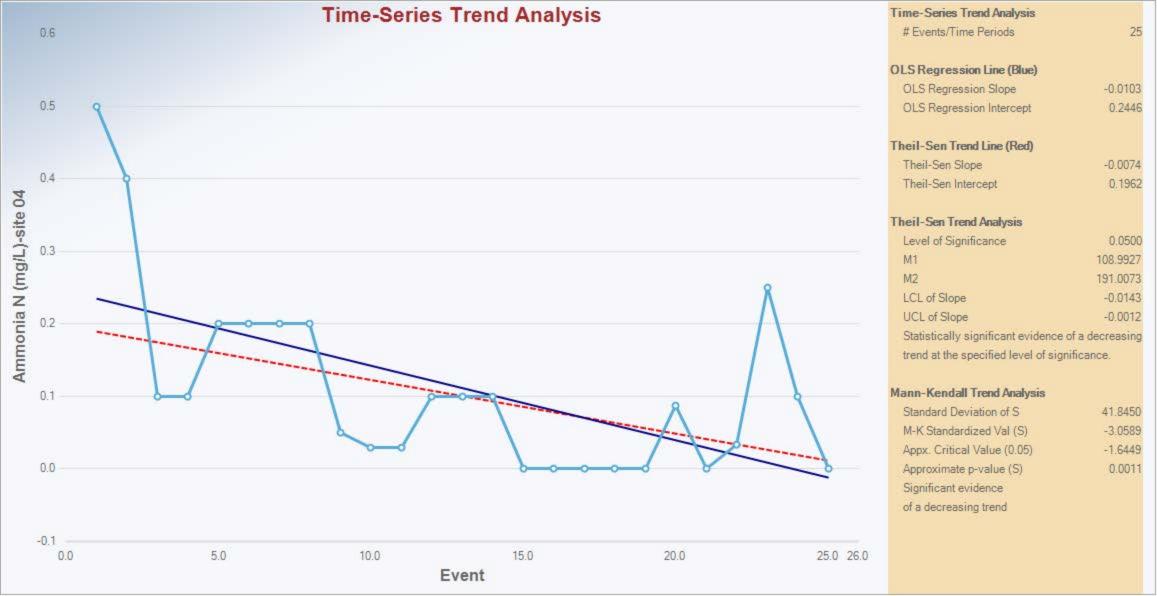


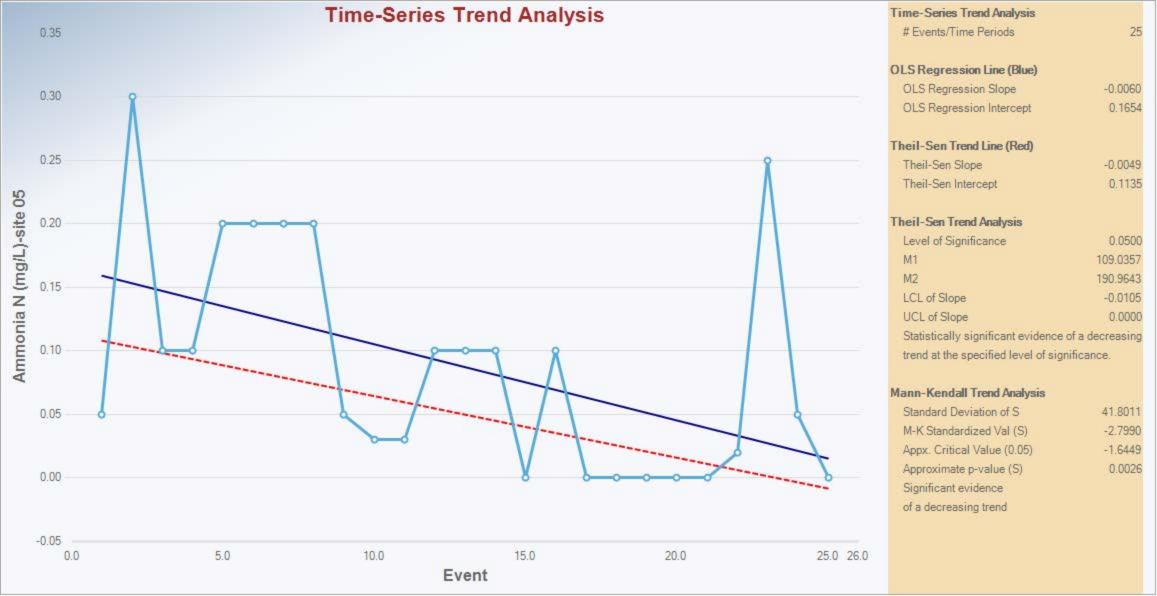


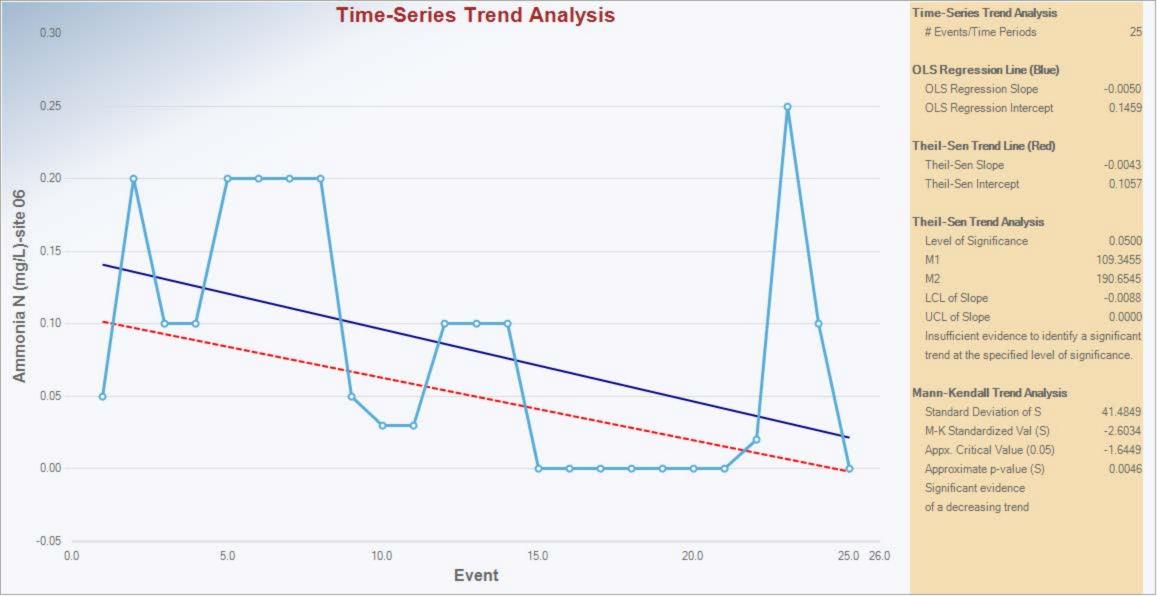


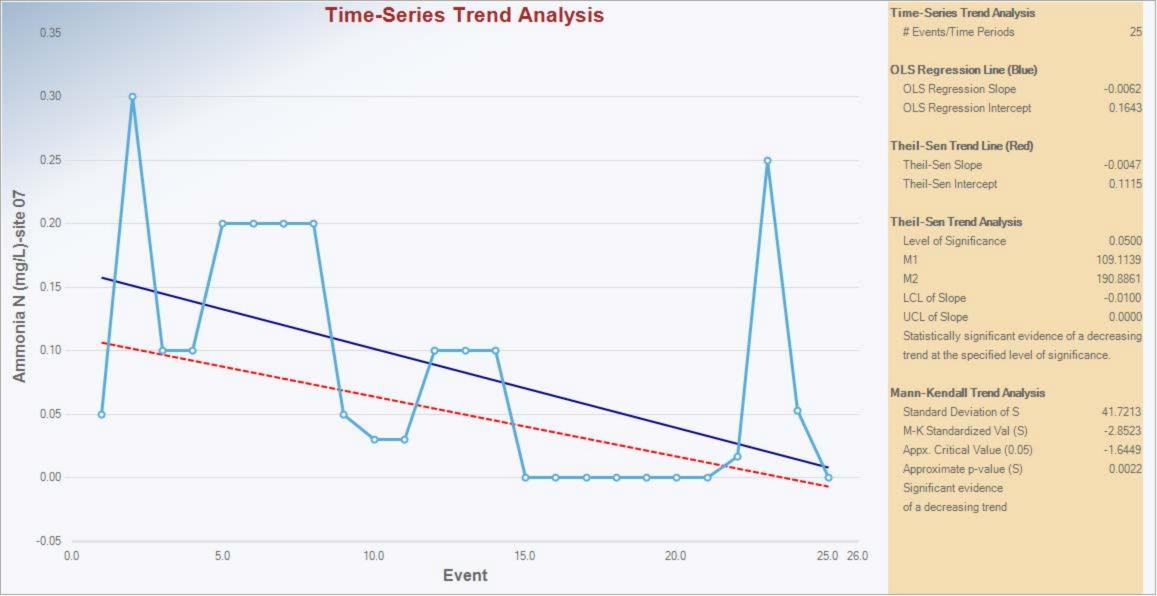


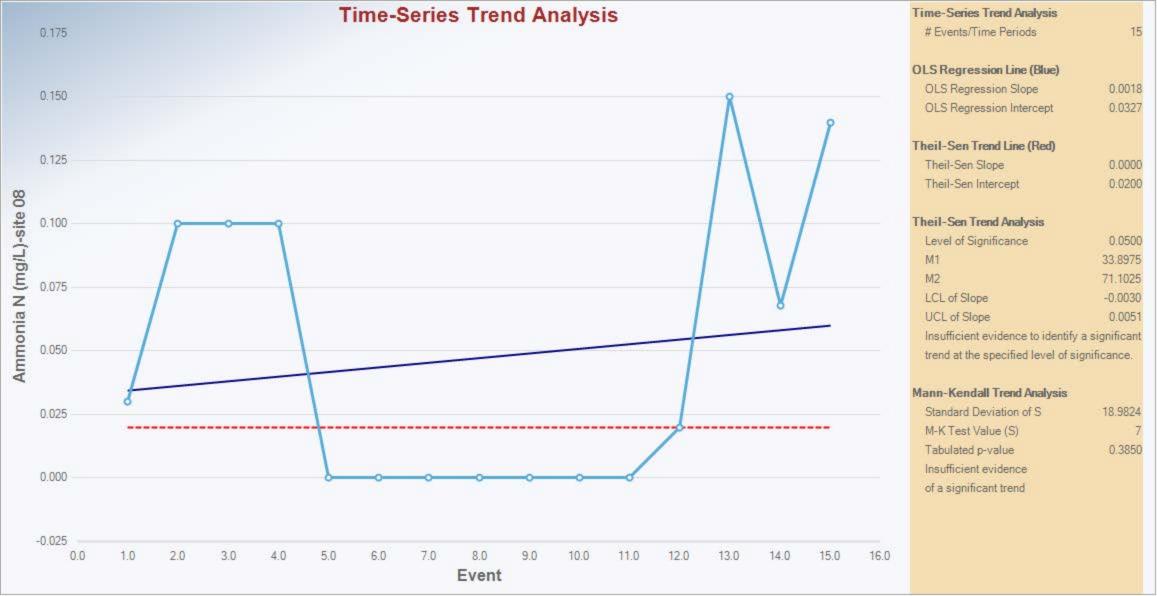


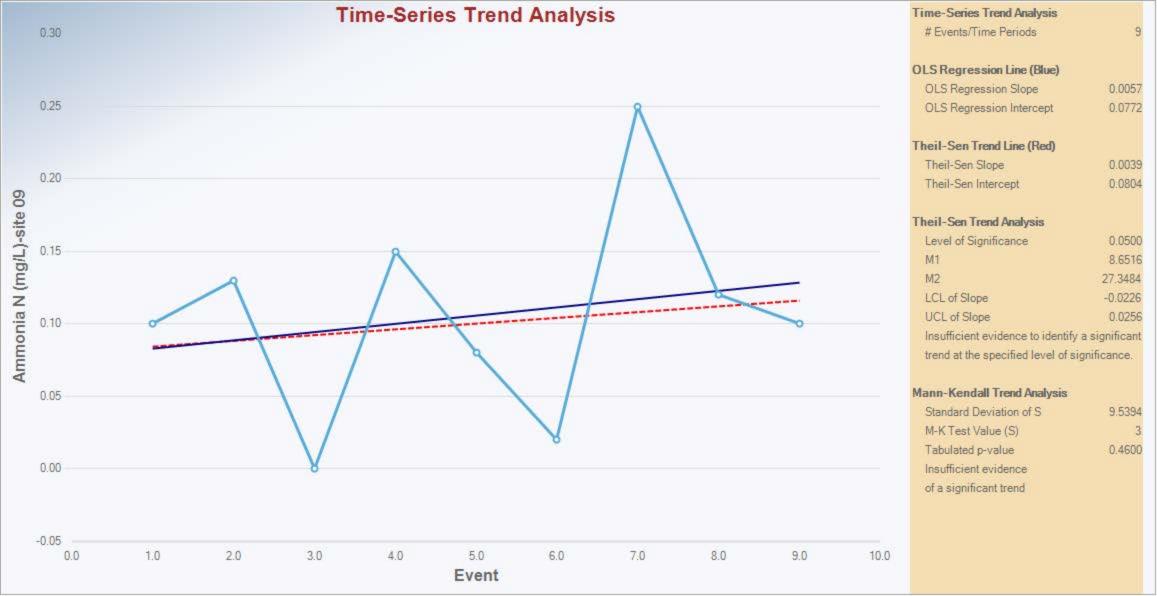








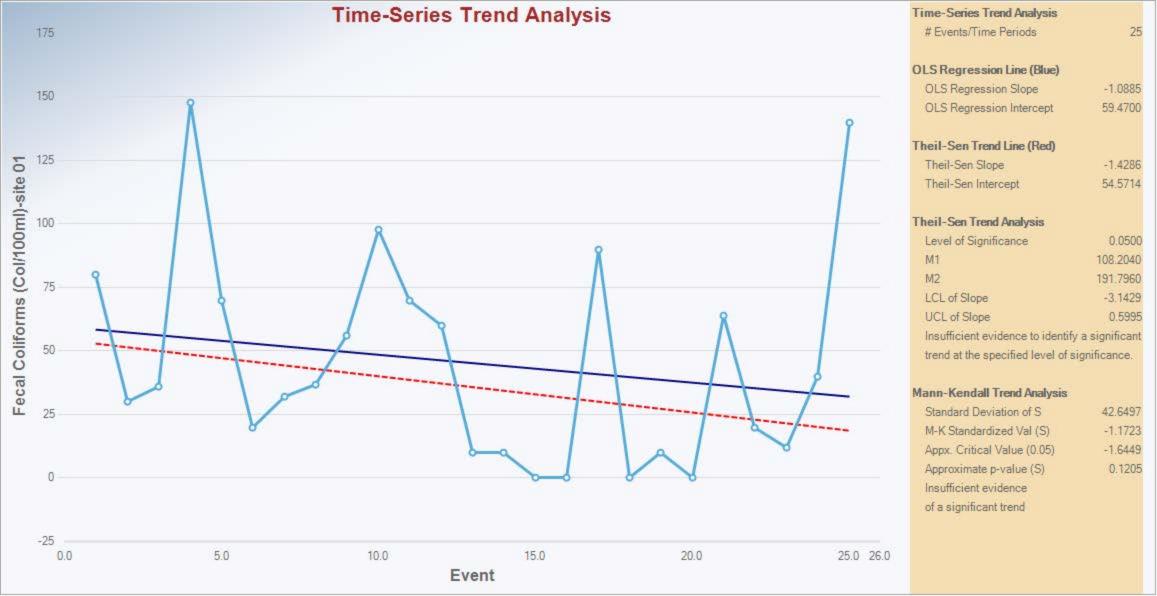


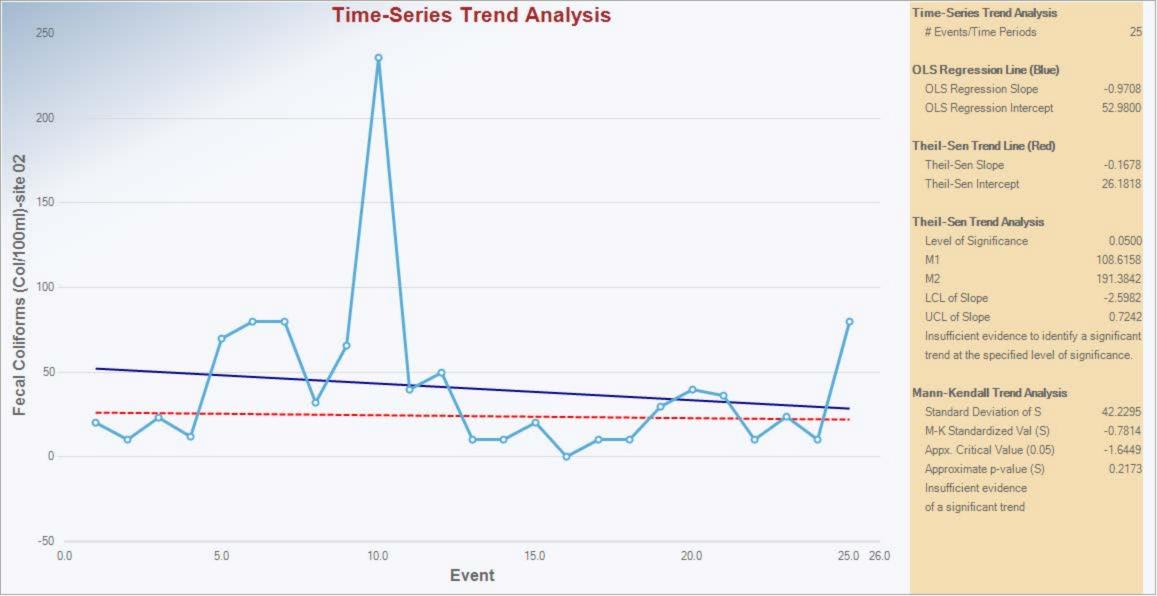


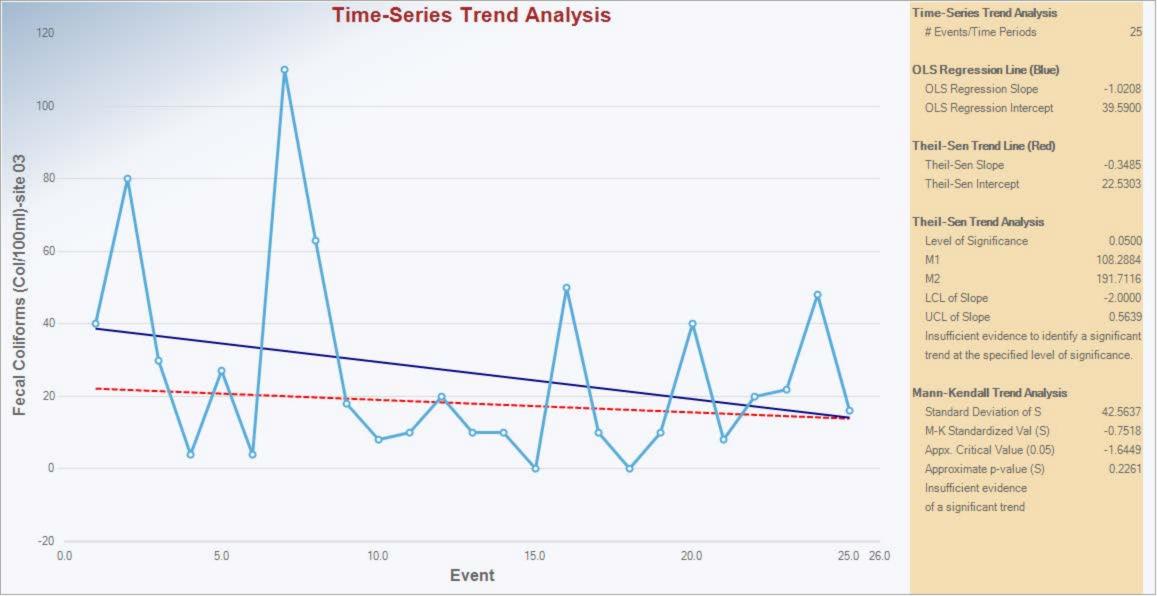
Trend Graph – E. coli in Surface Water (2015-2024) Station 10

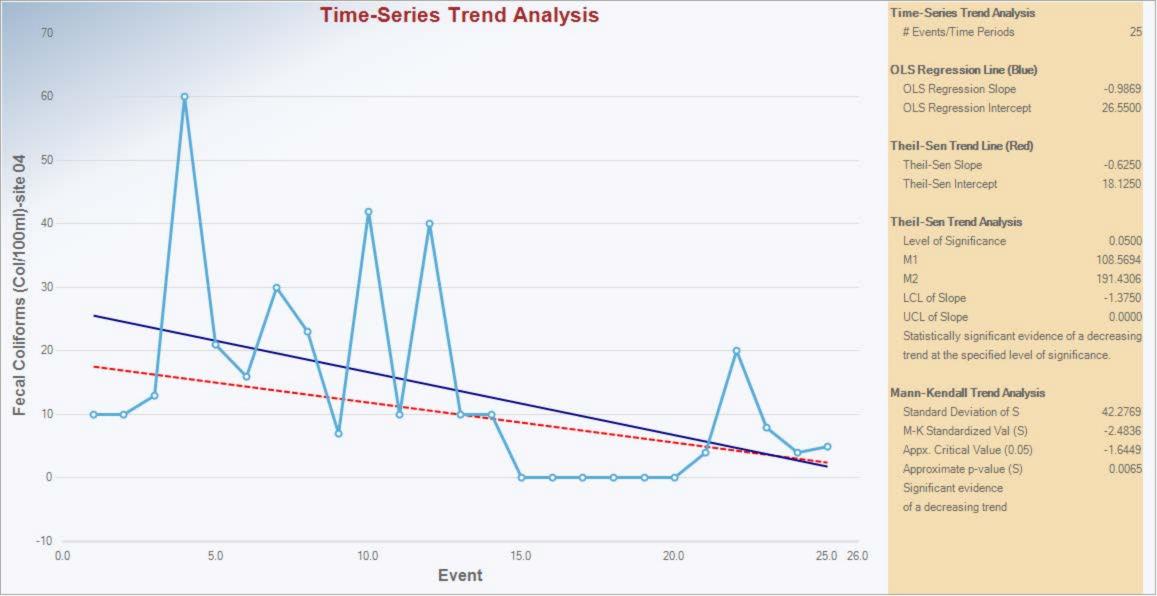


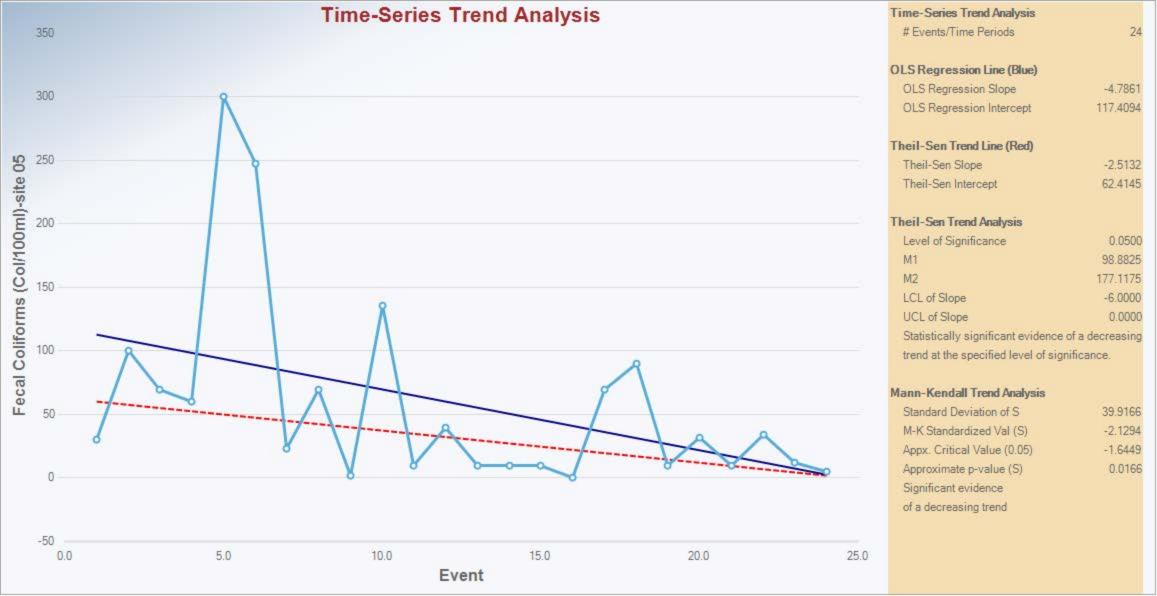


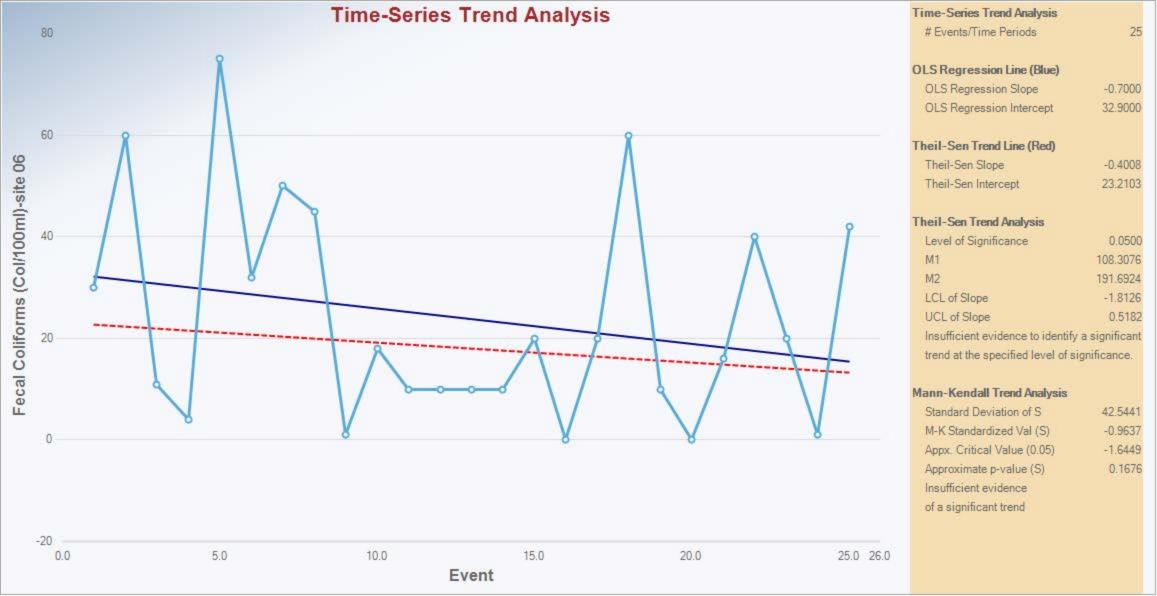


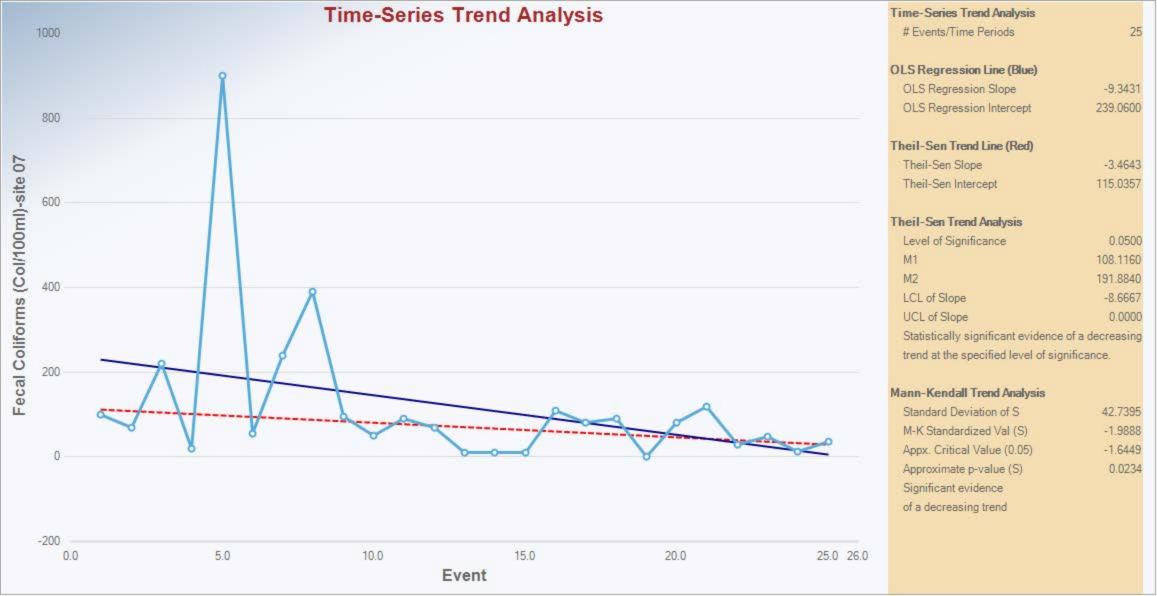


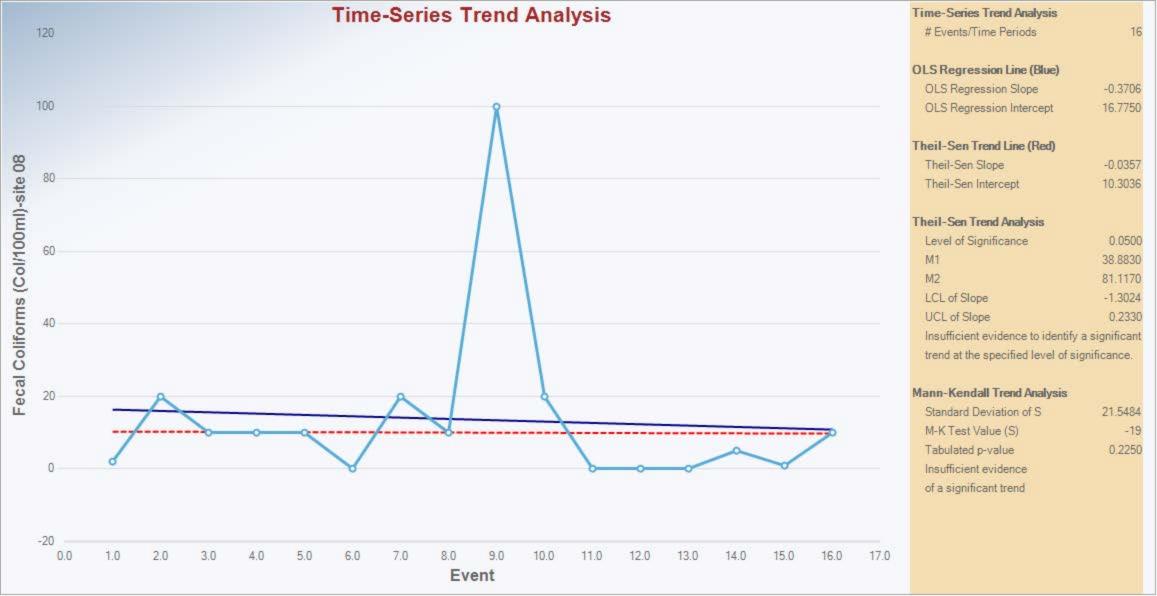


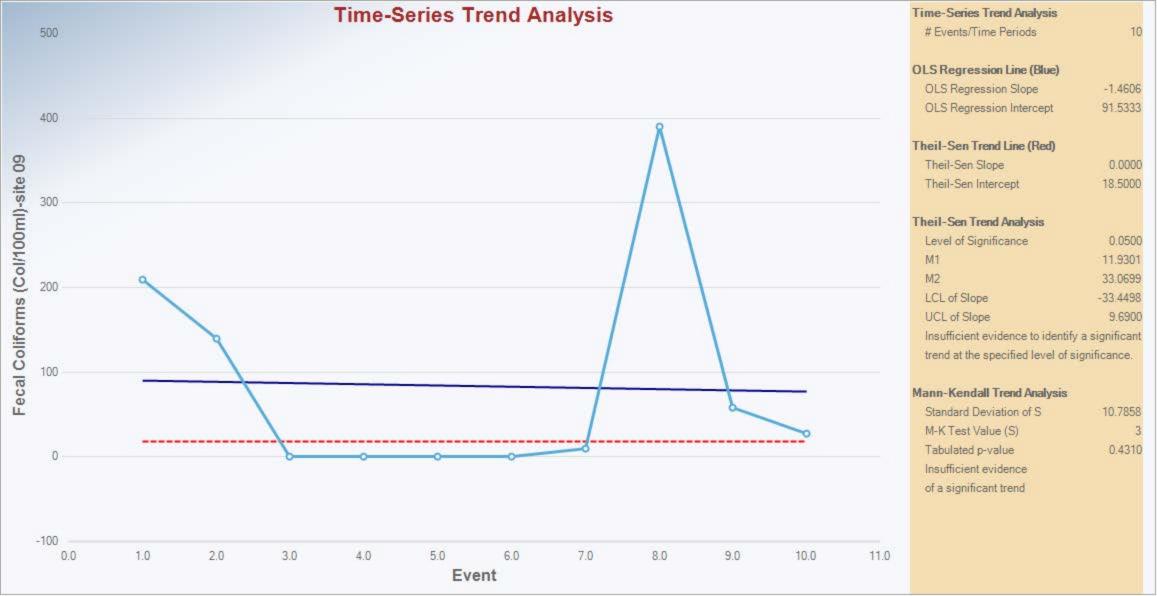


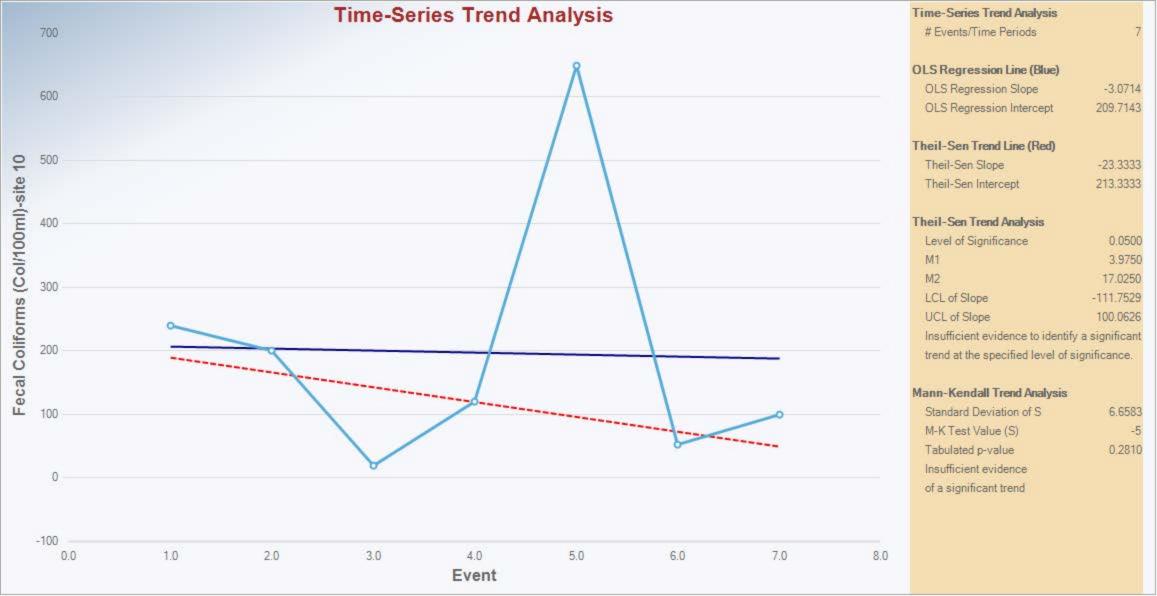






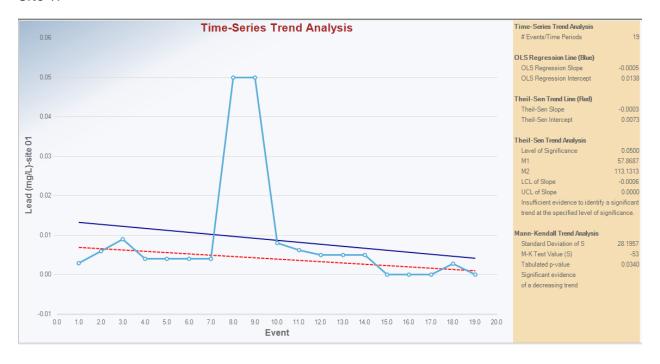






Lead graphs

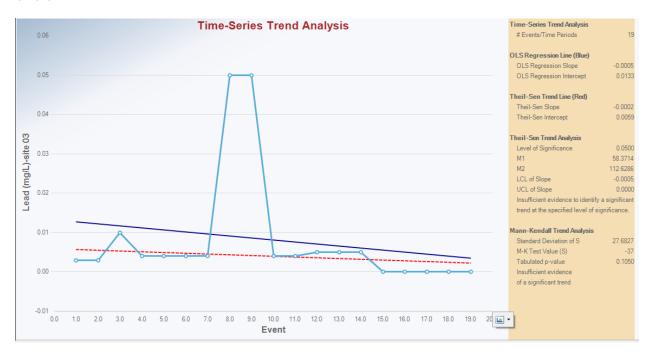
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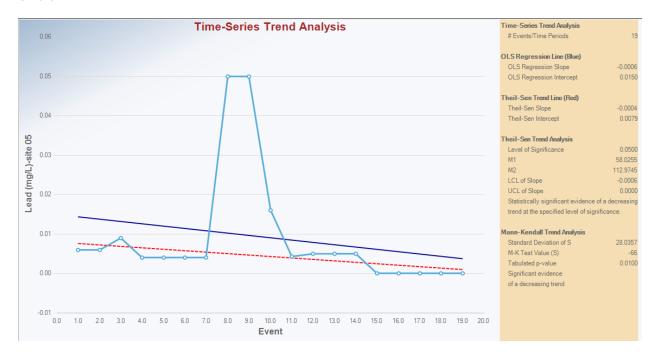
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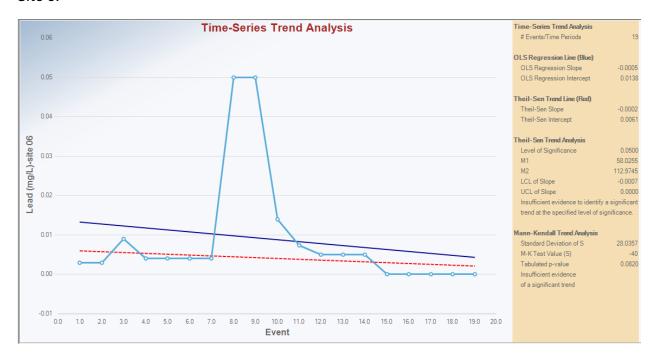
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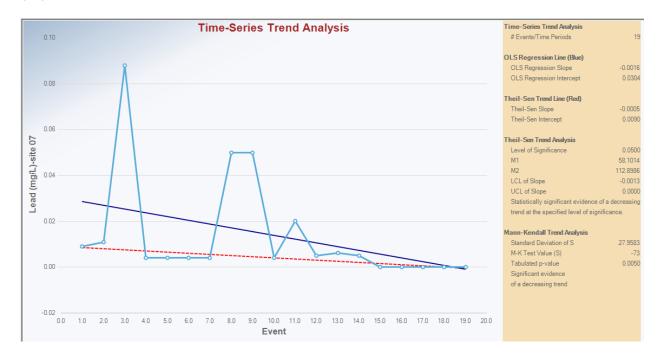
Site 5:



Site 6:



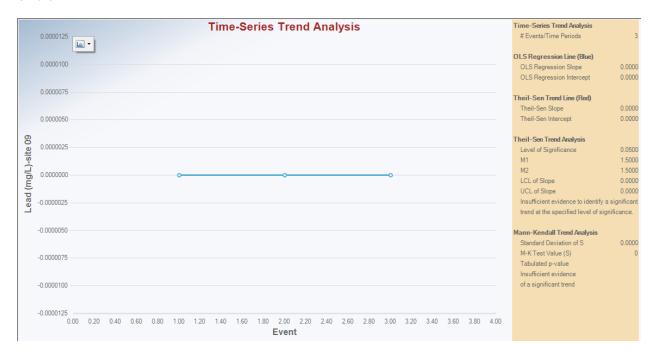
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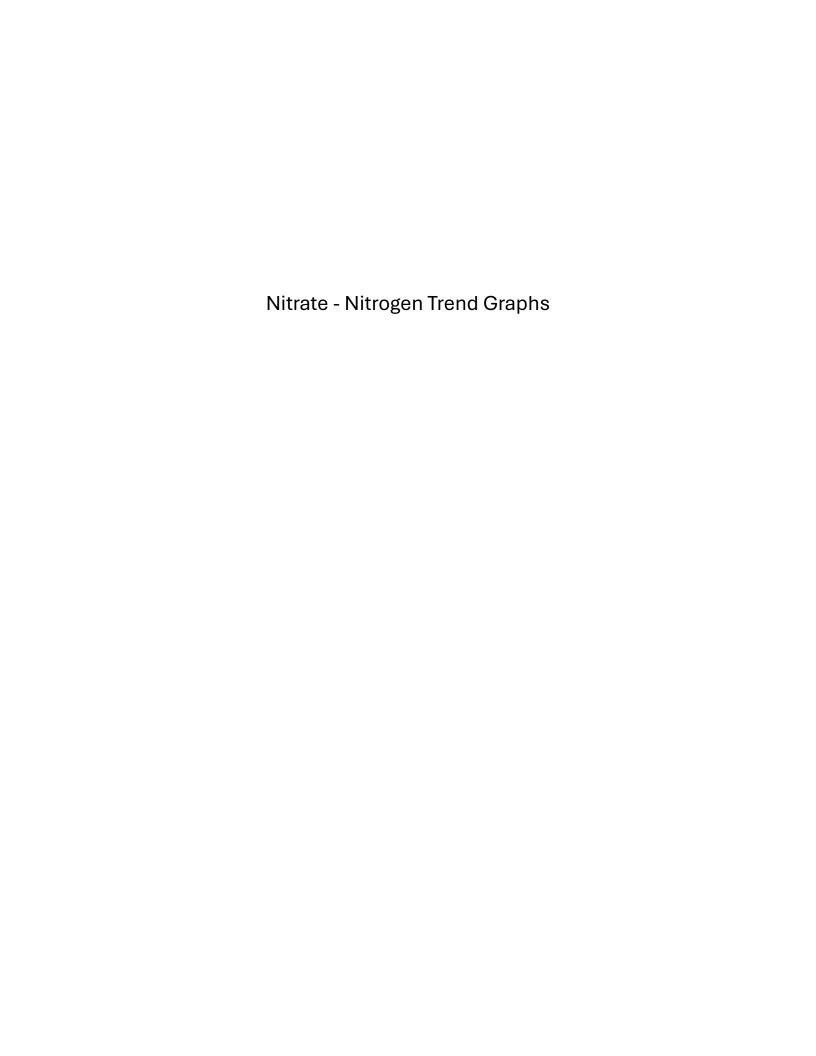


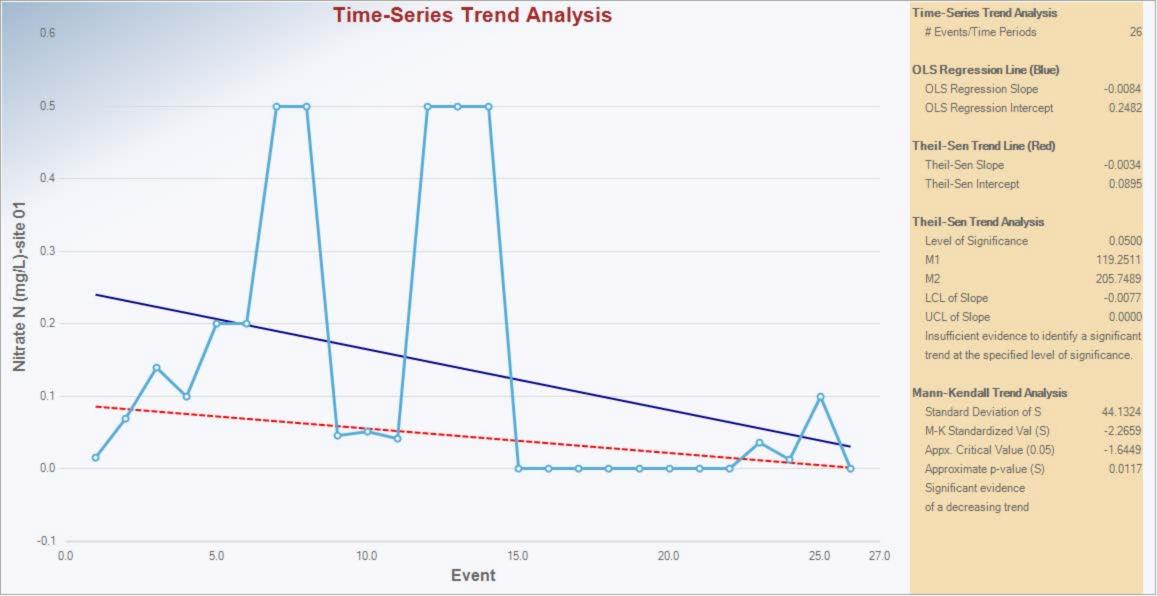
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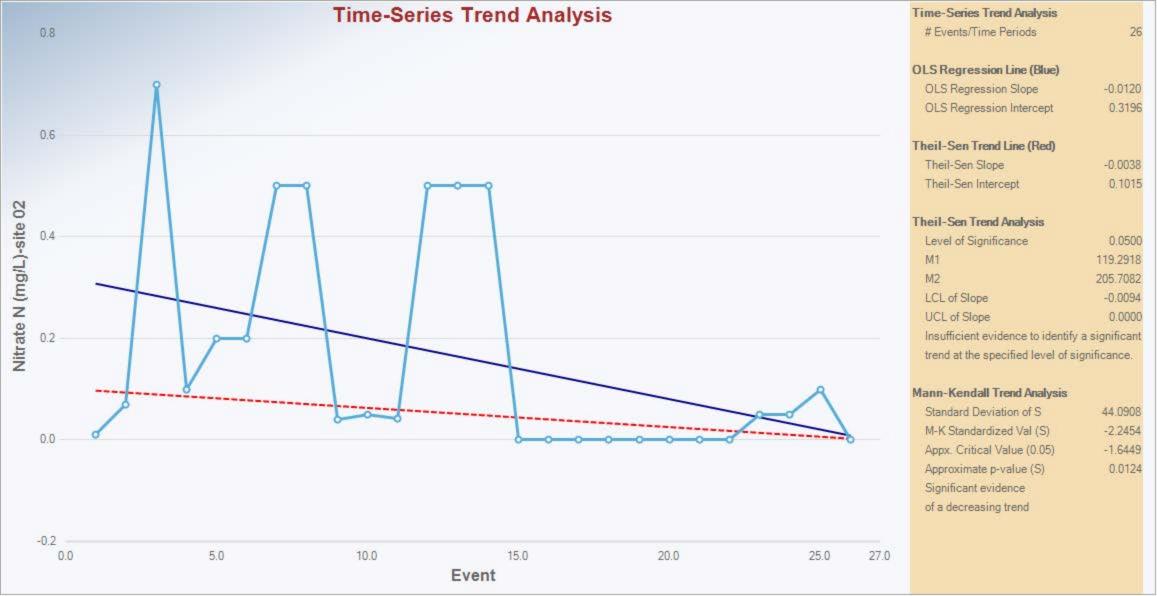


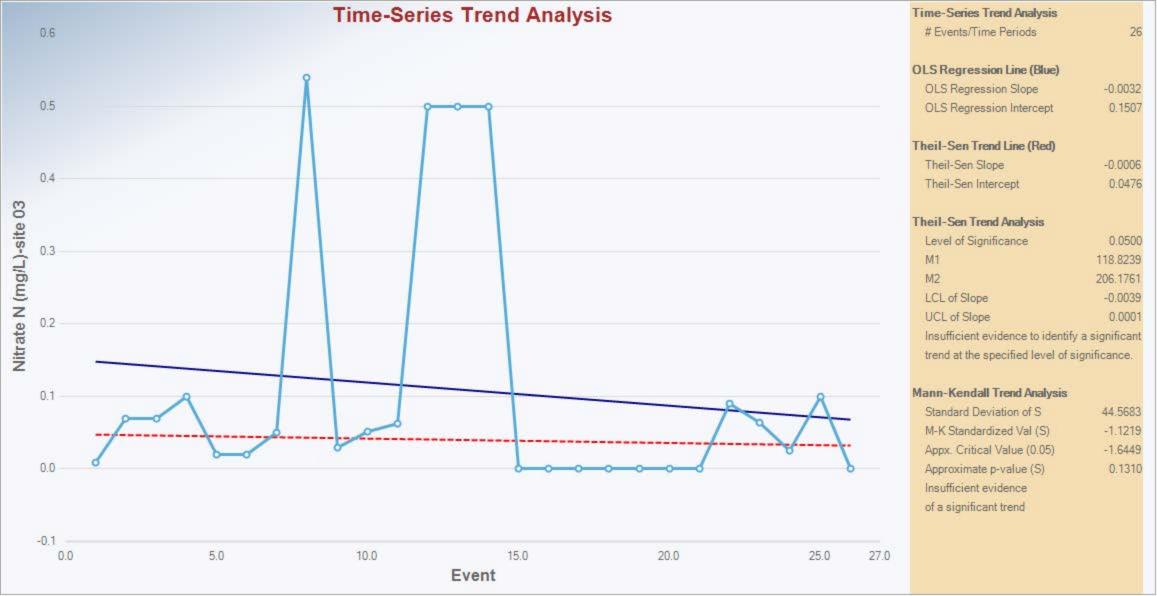
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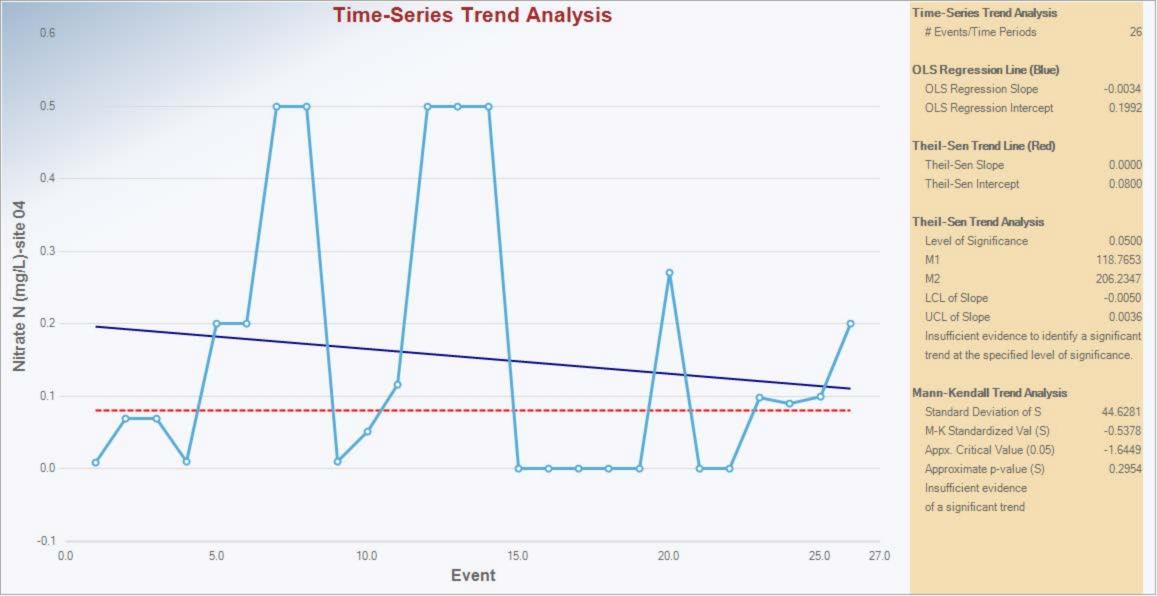


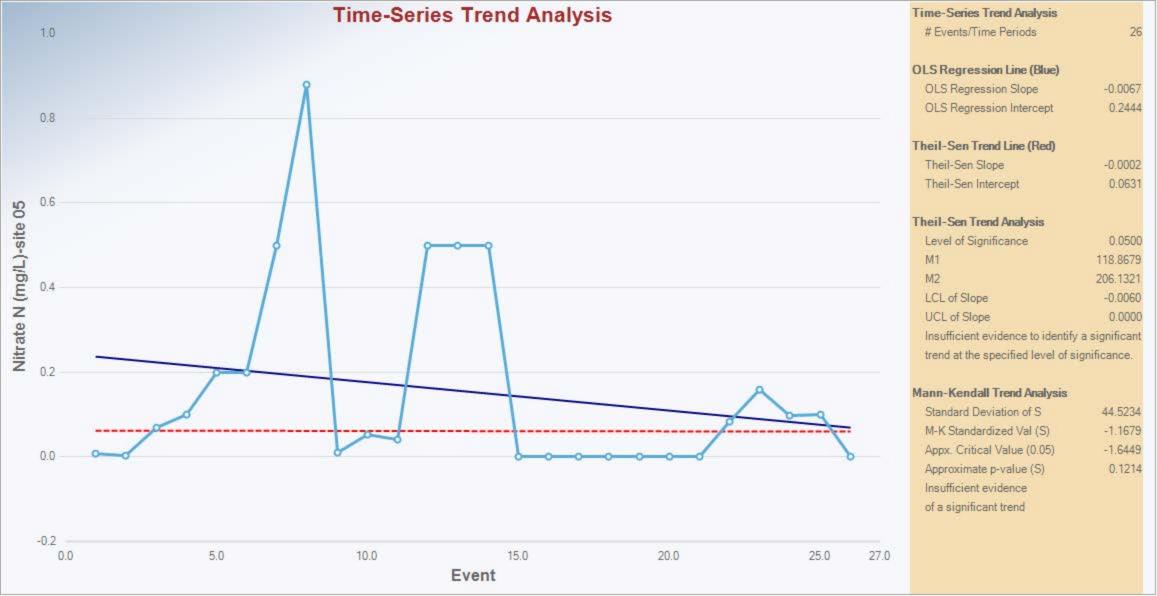


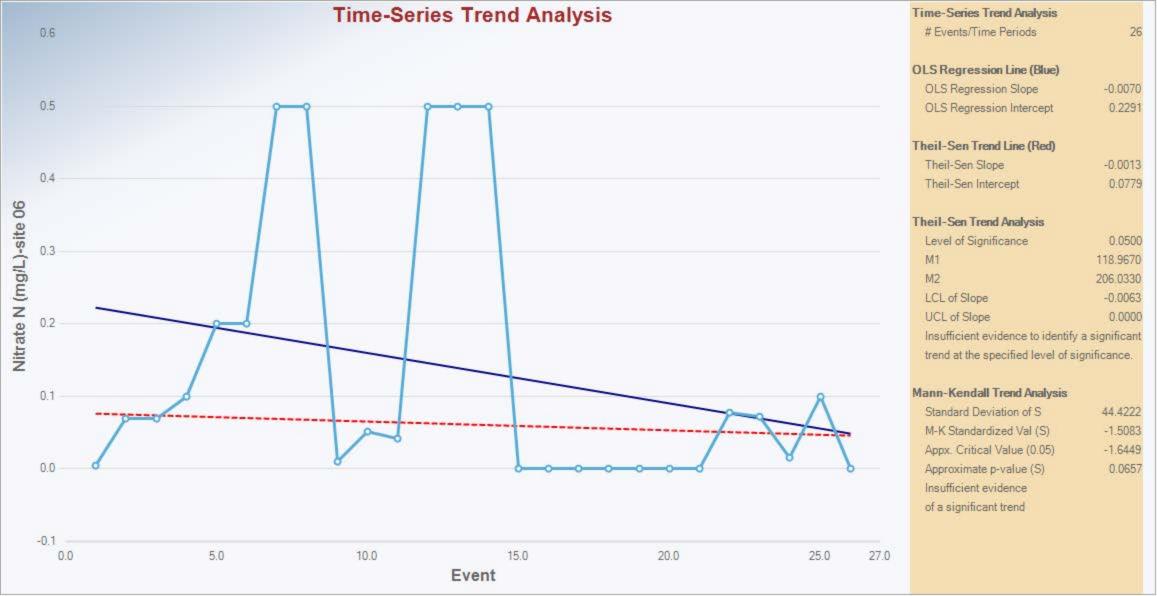


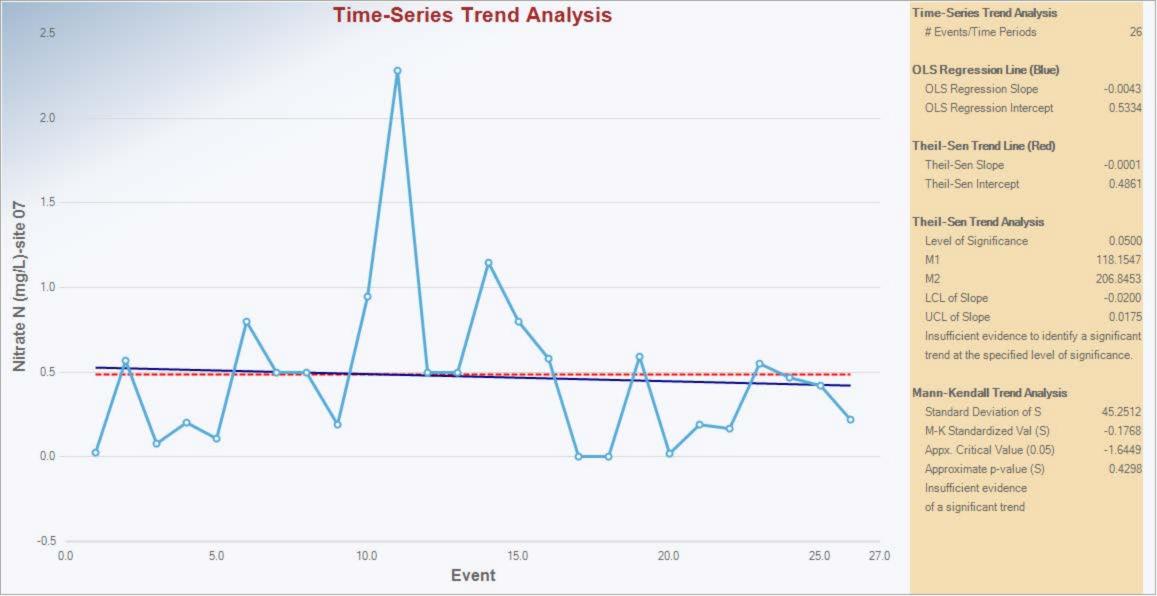


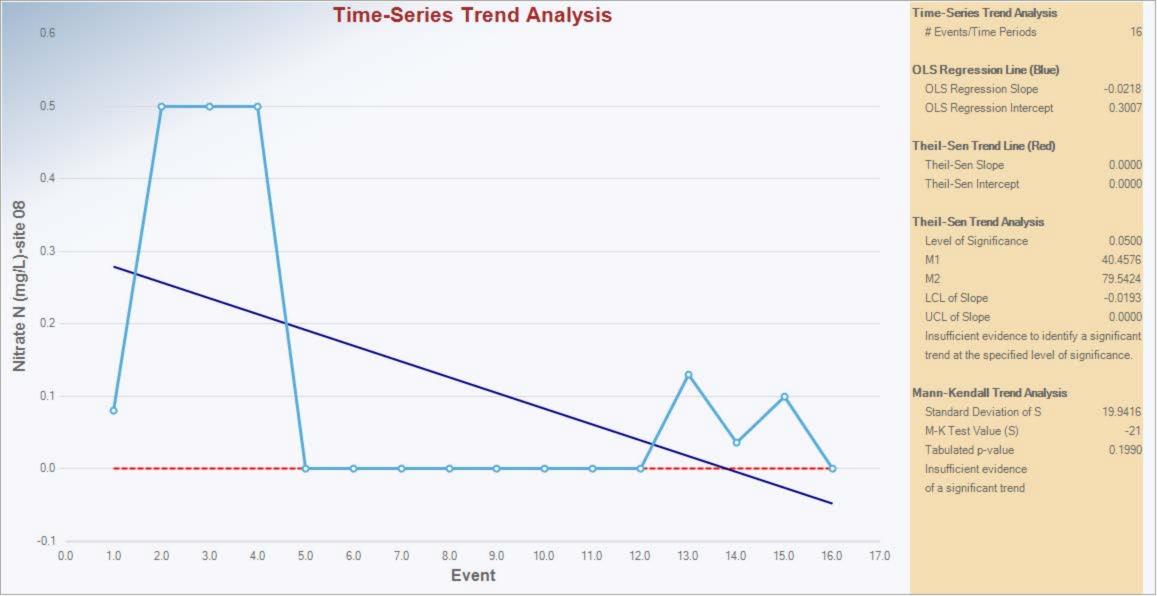


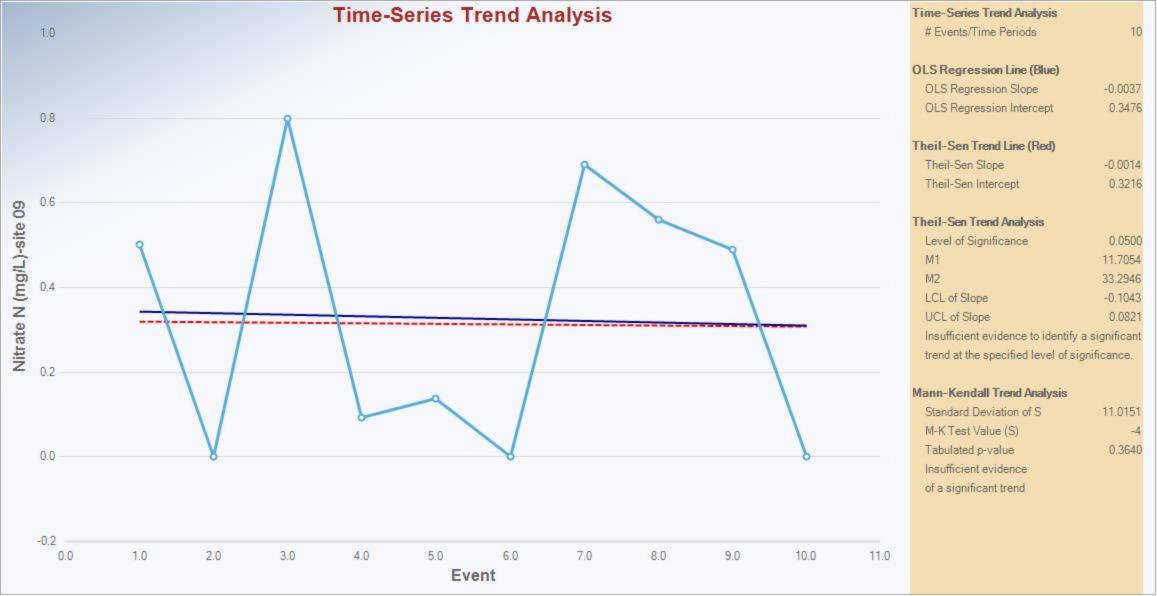


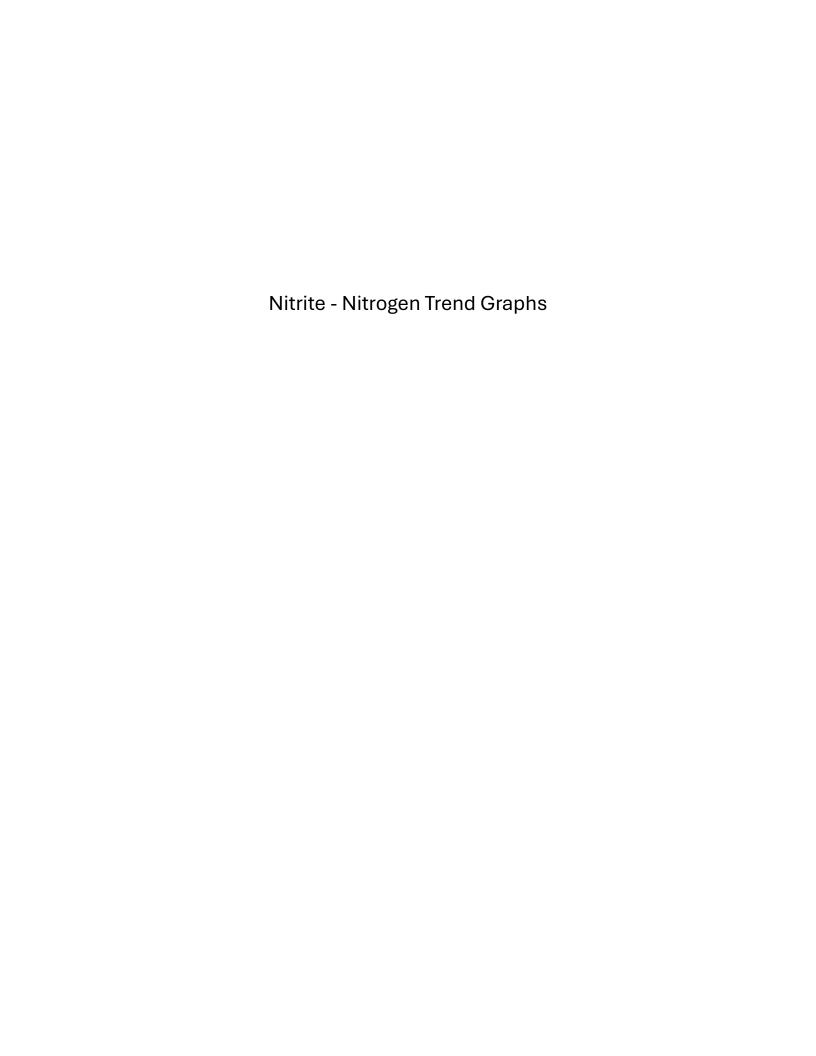


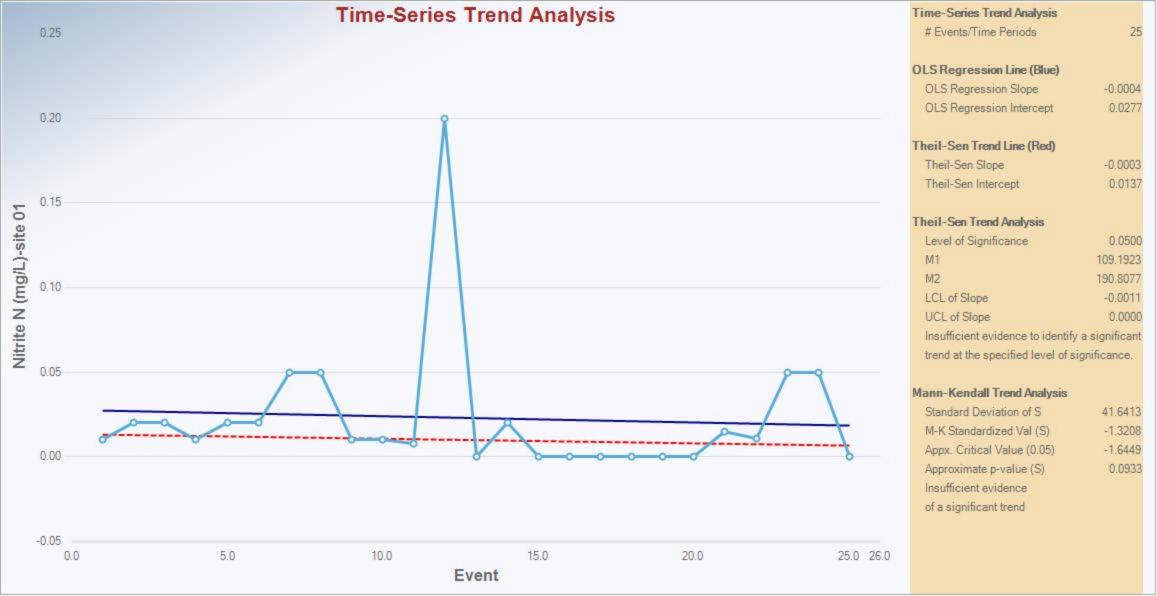


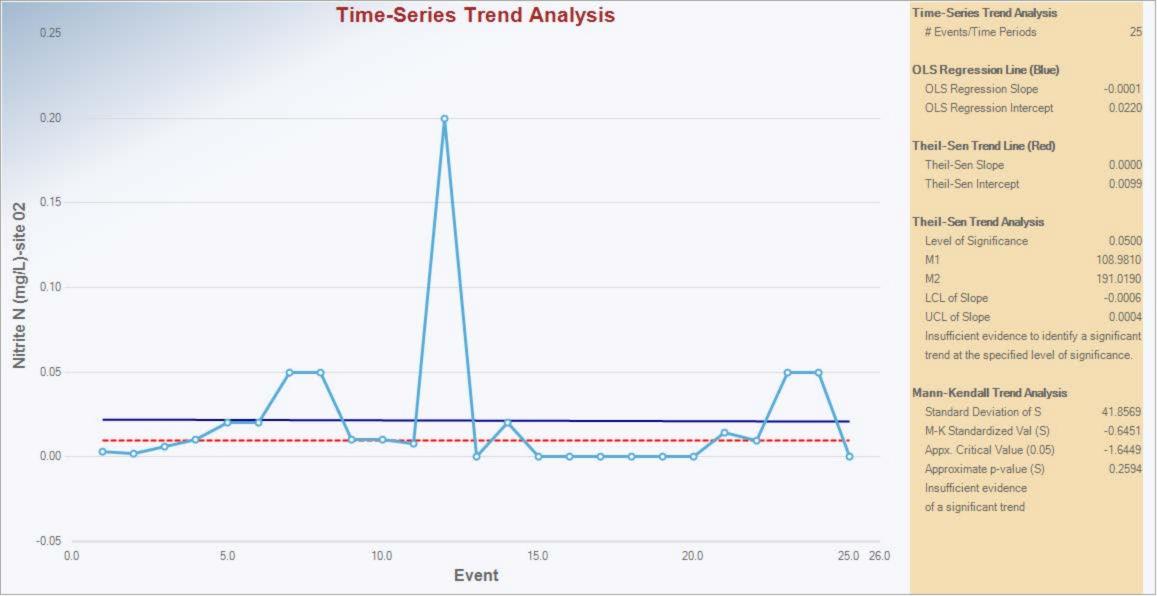


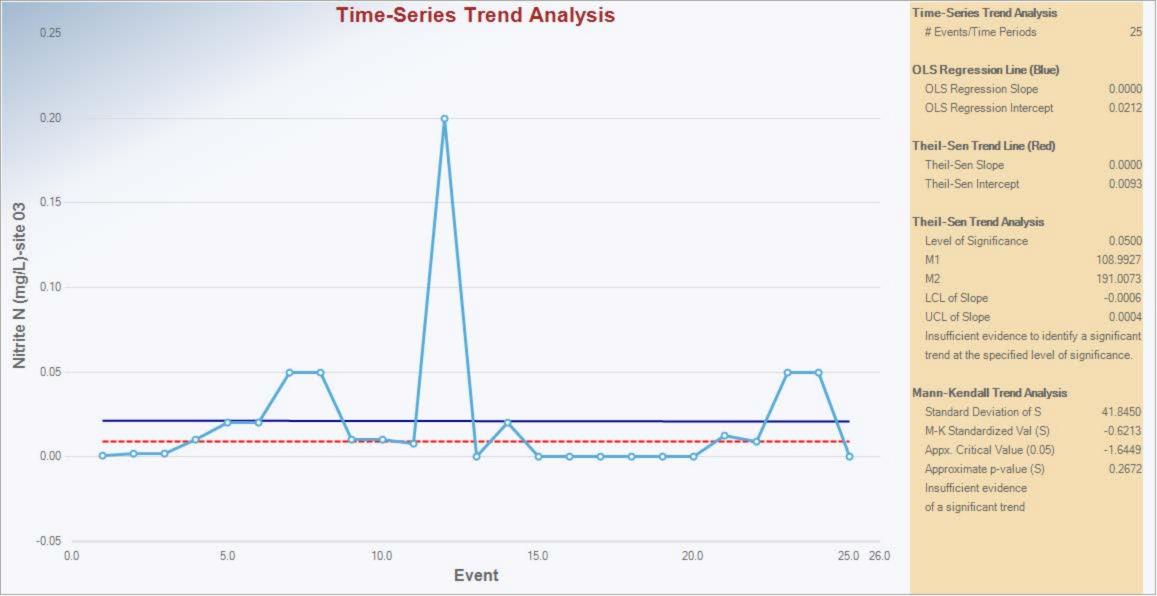


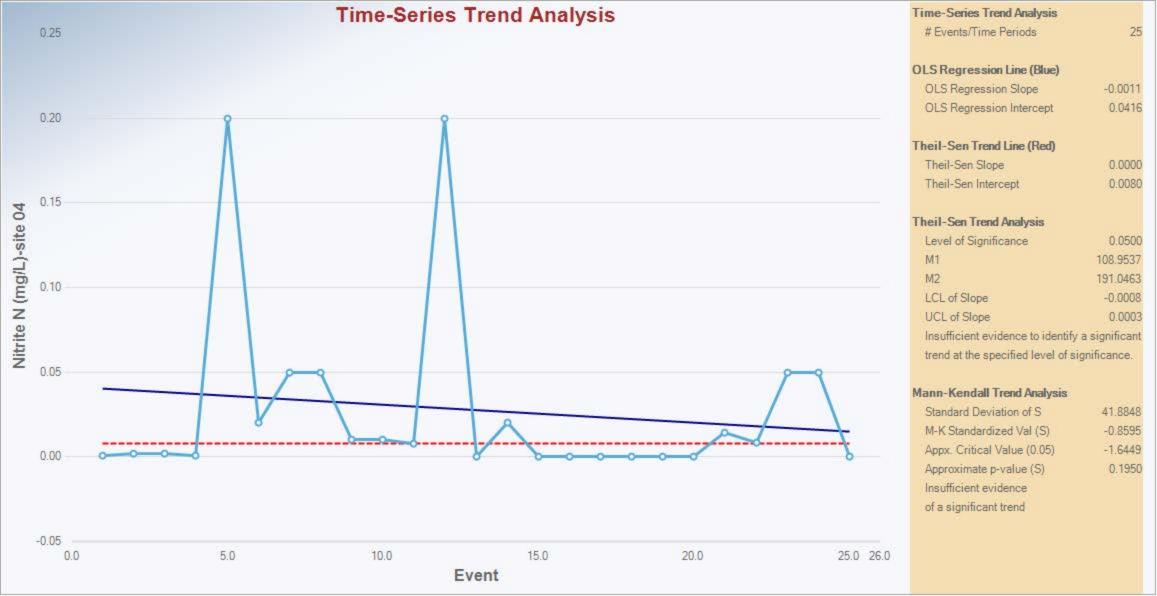


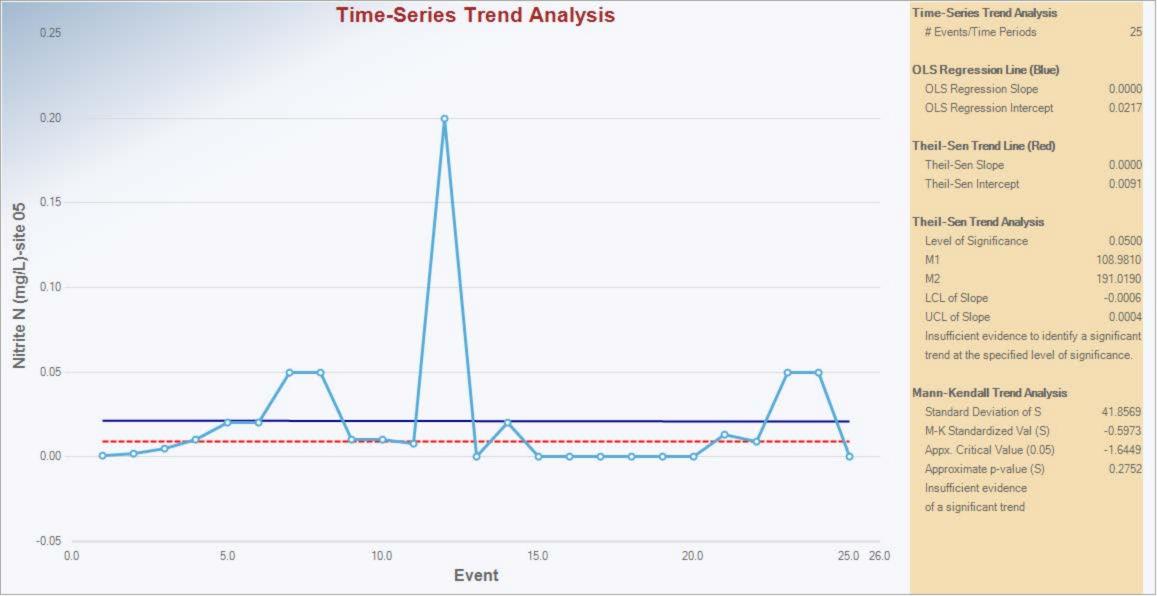


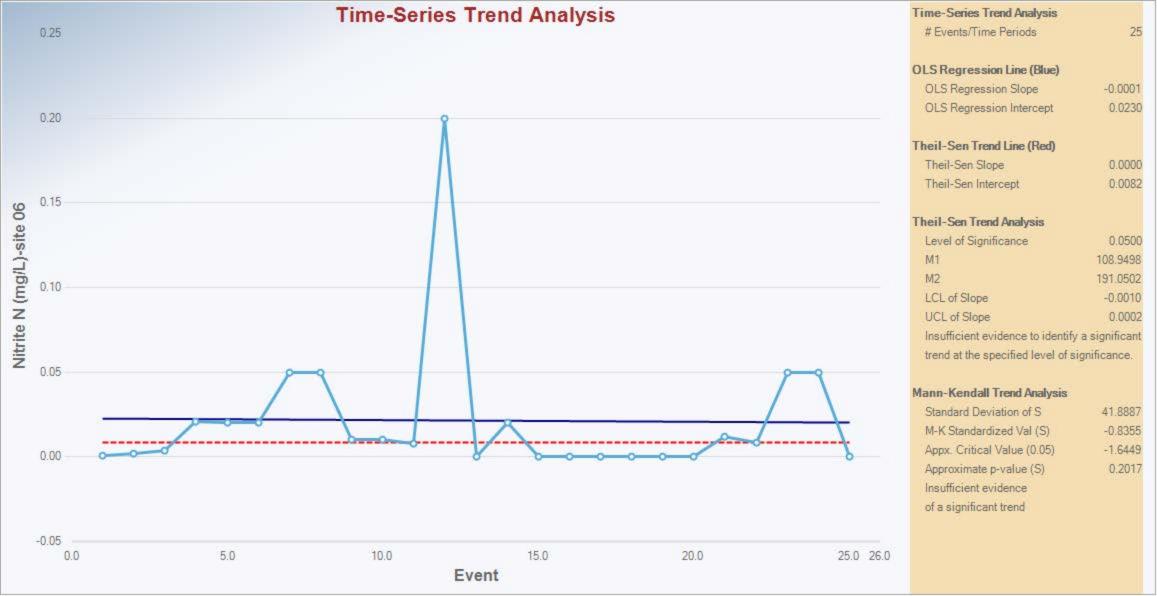


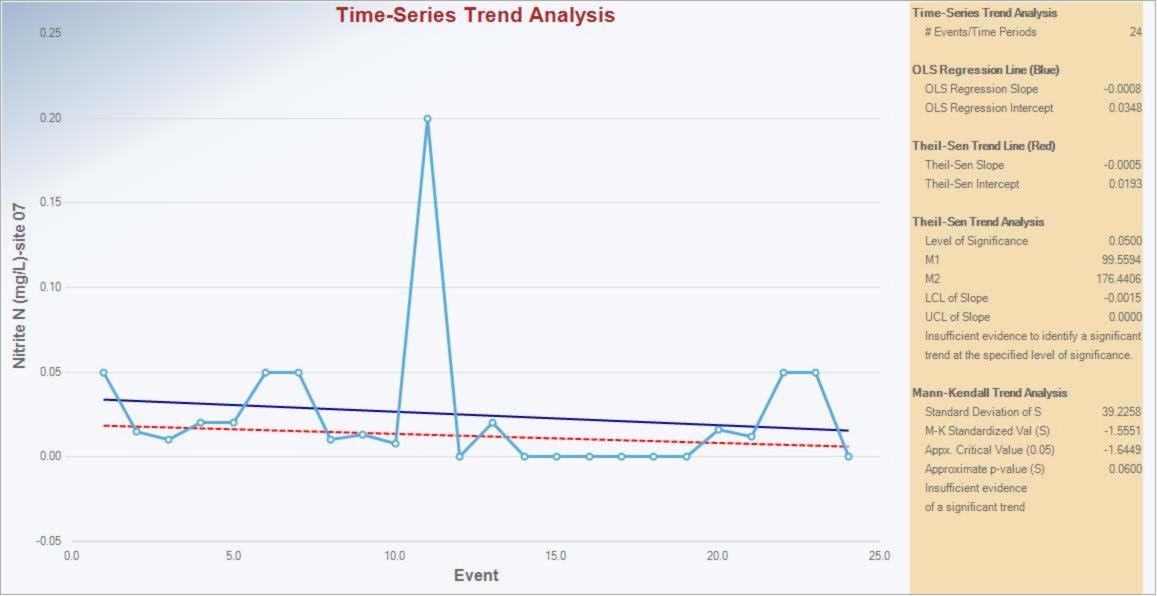


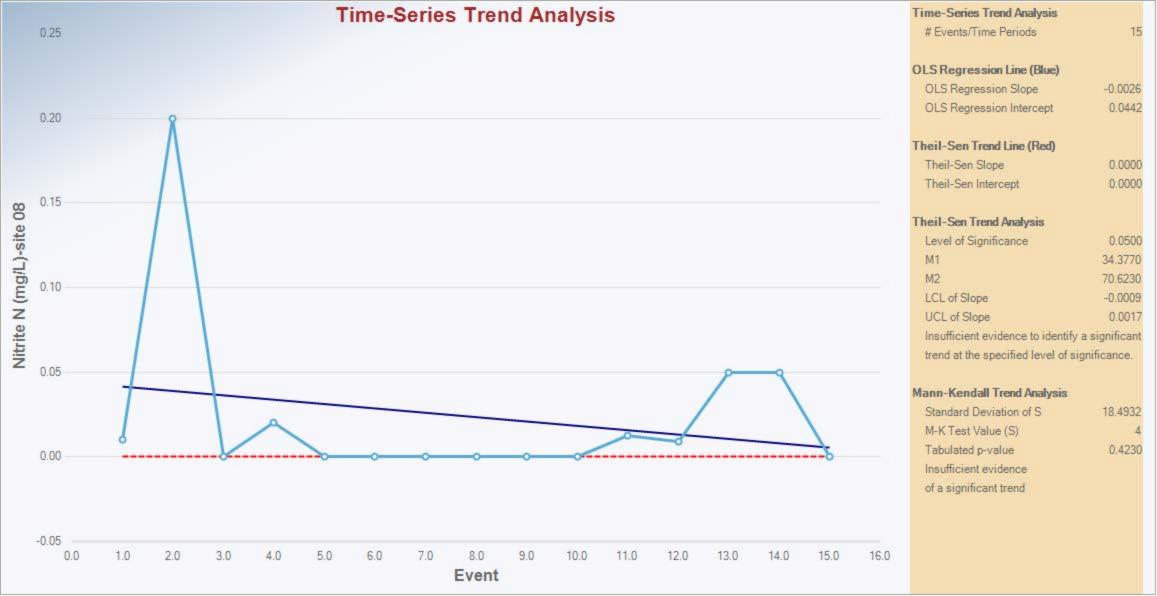


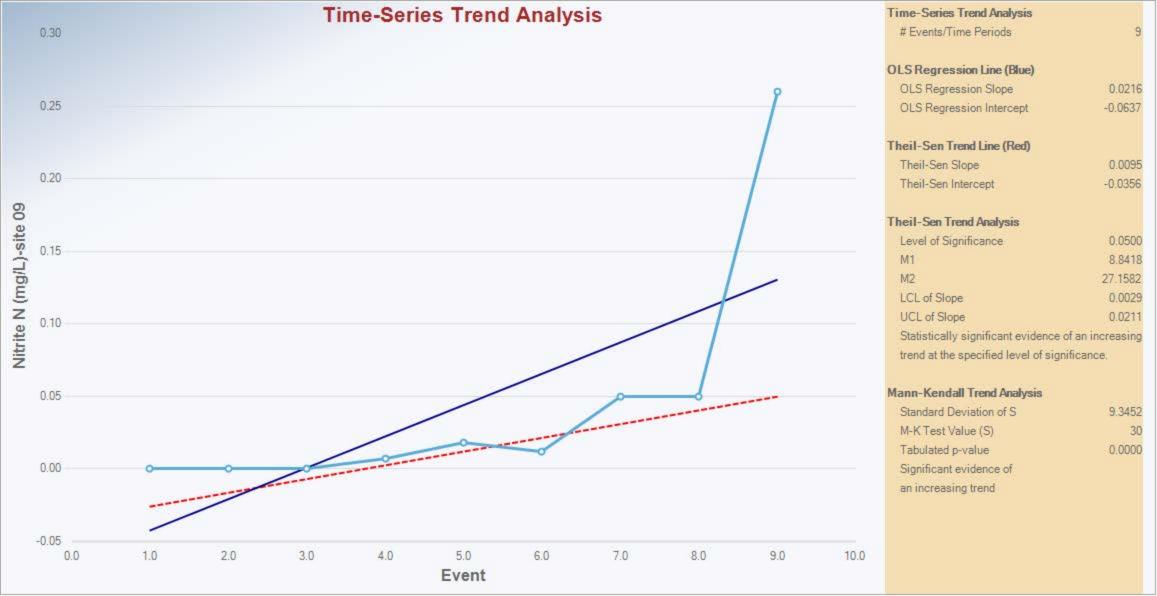




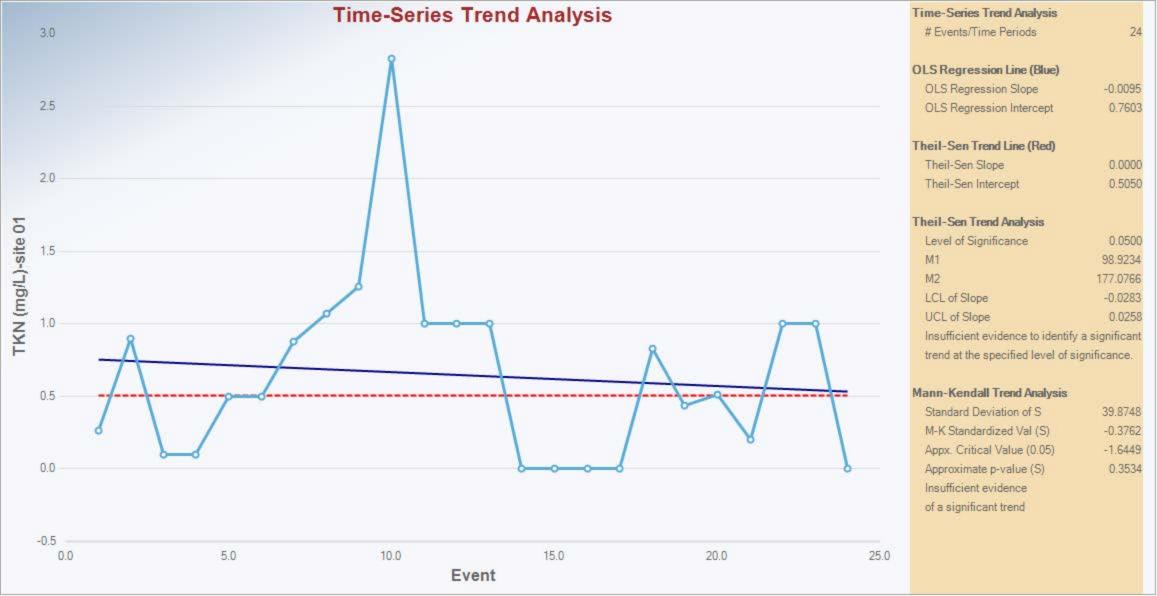


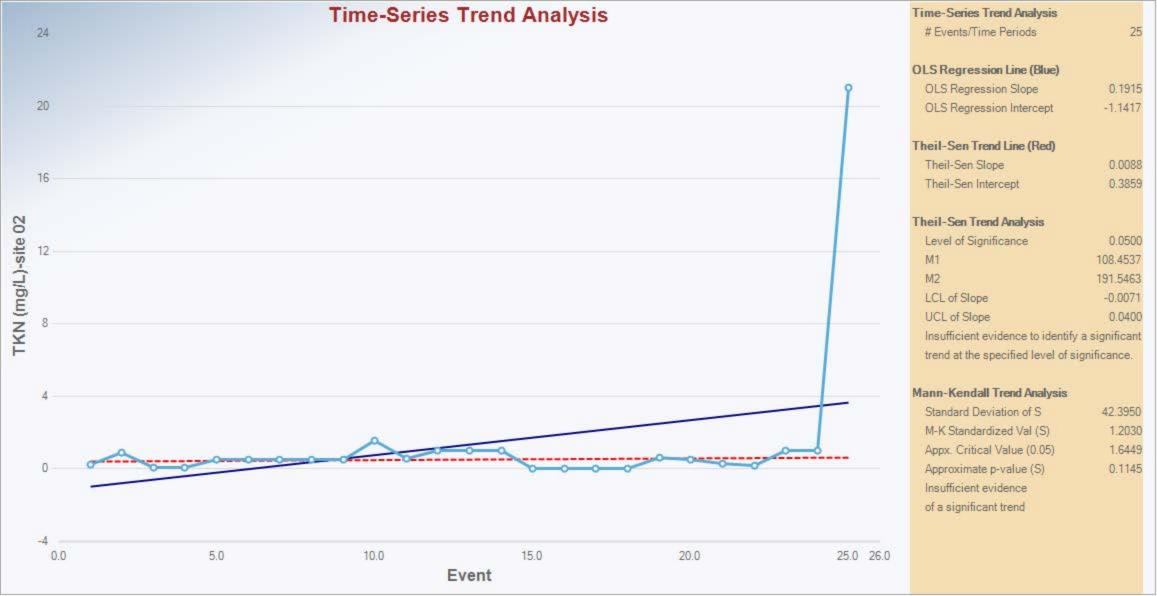


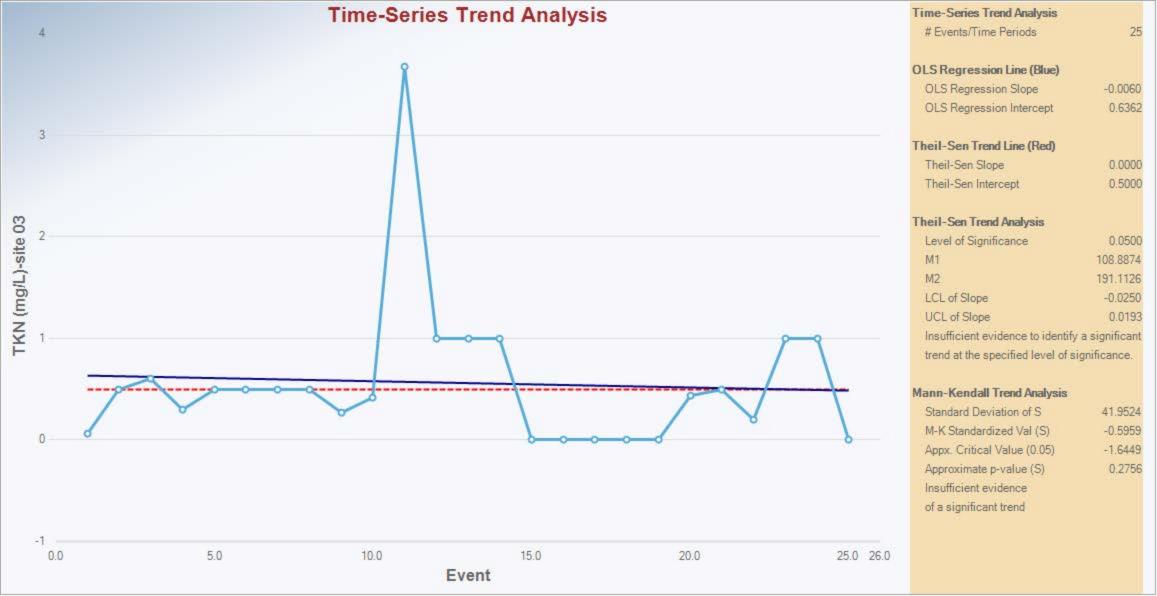


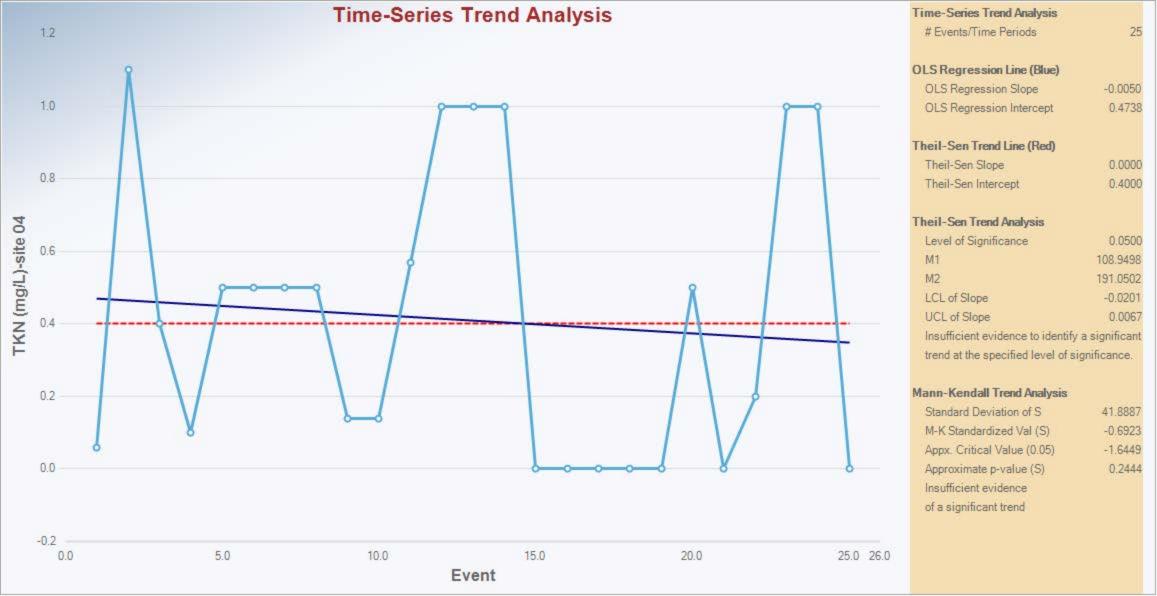


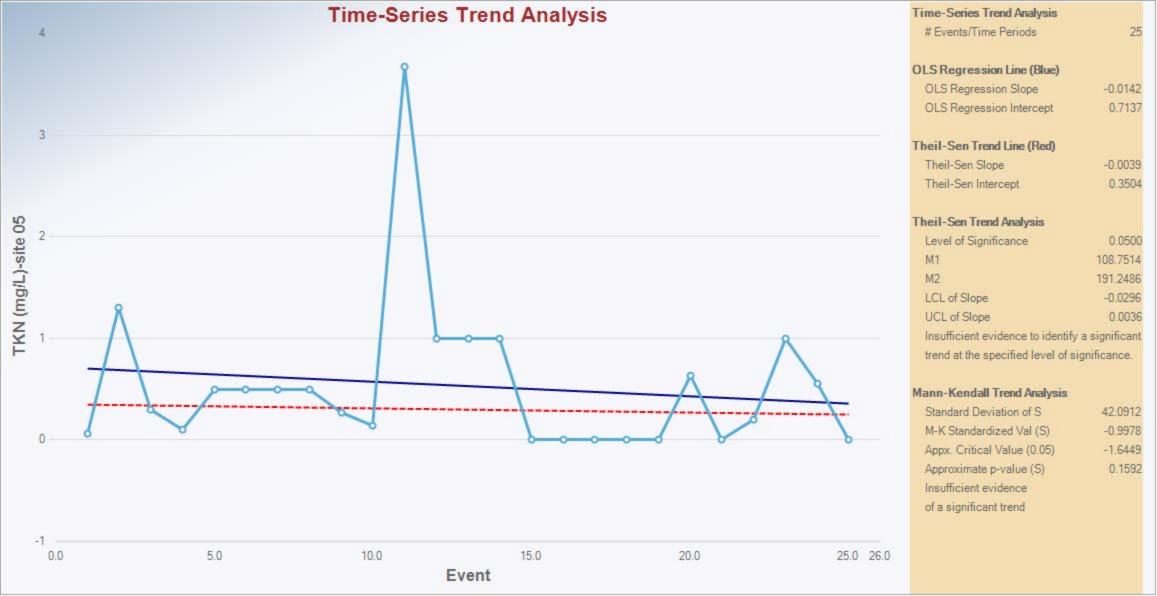


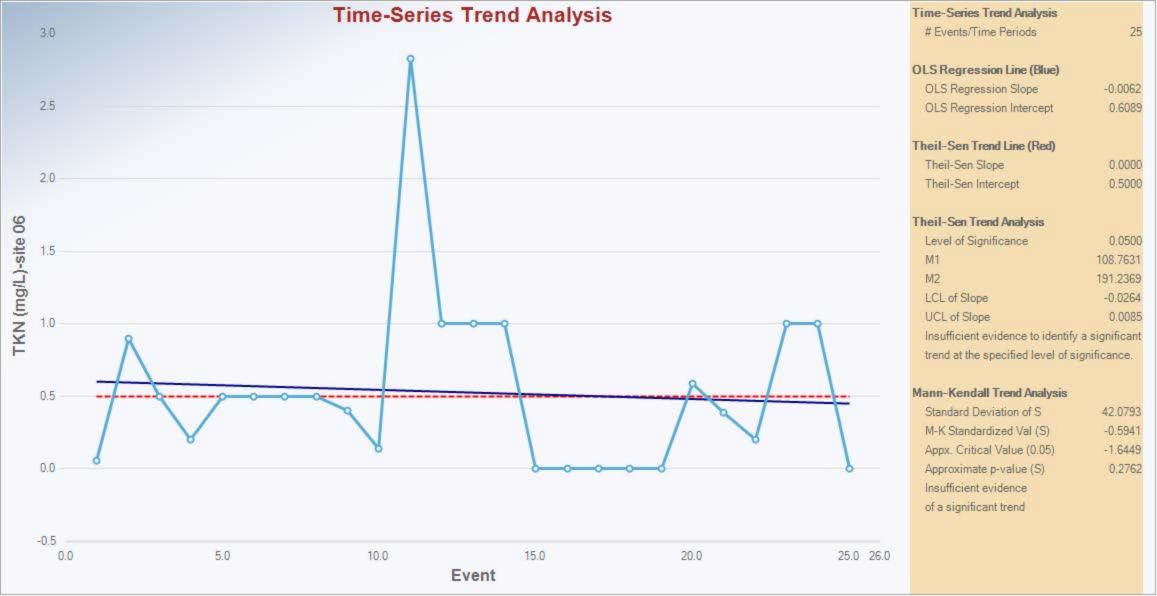


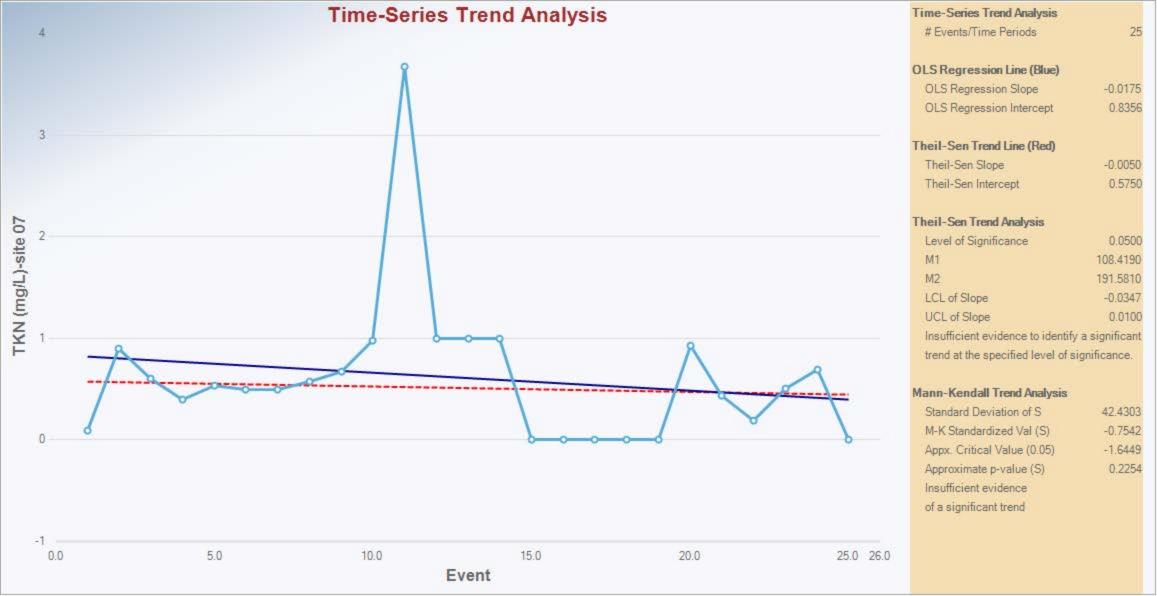


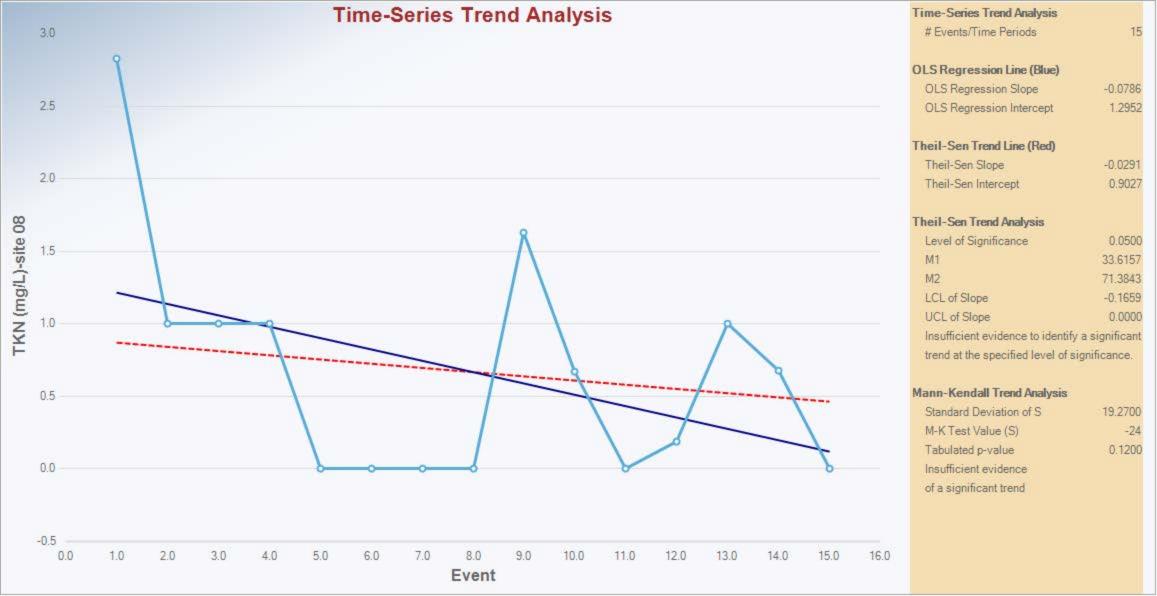


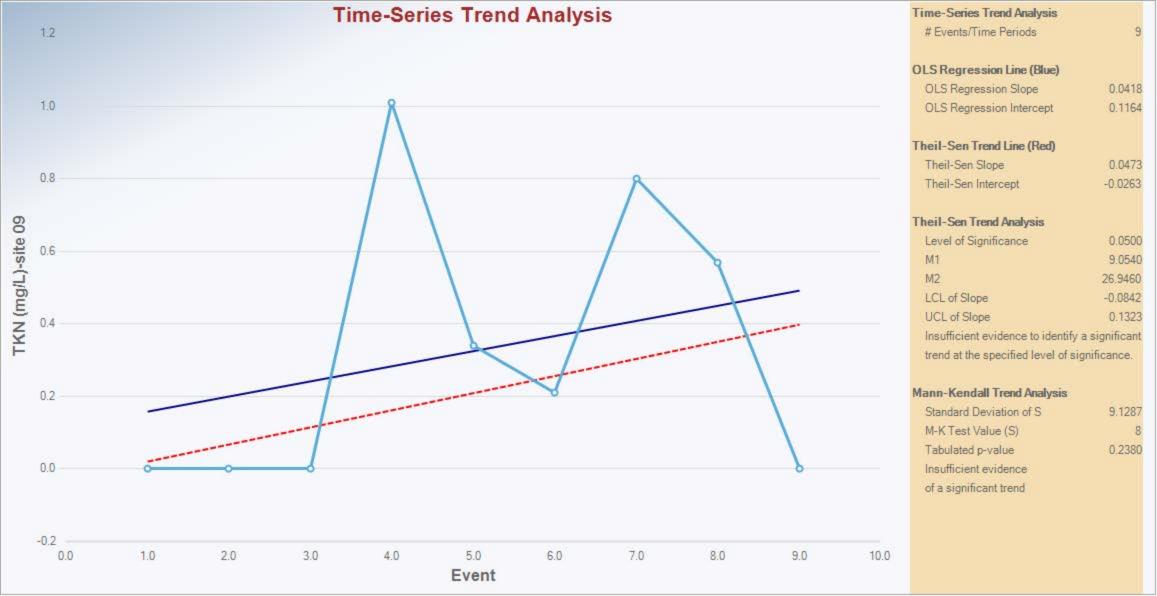




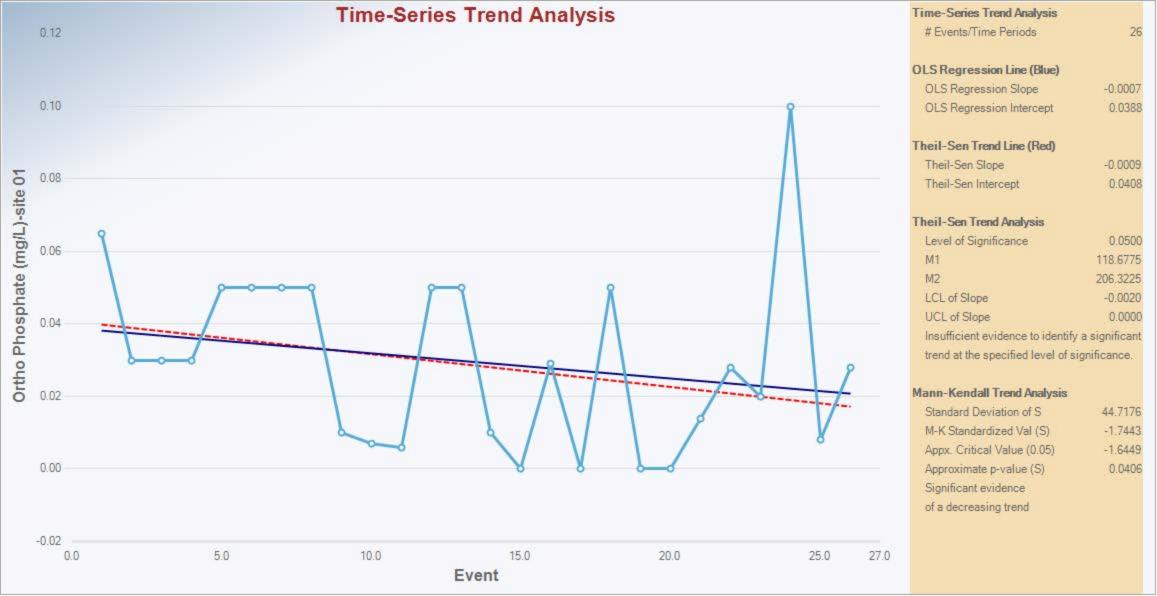


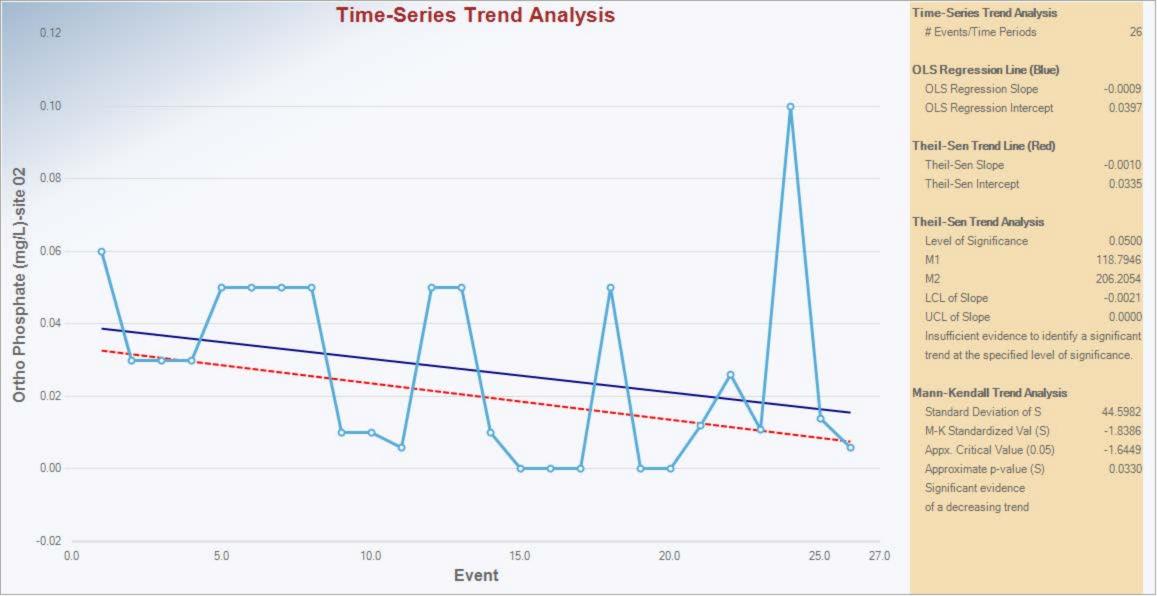


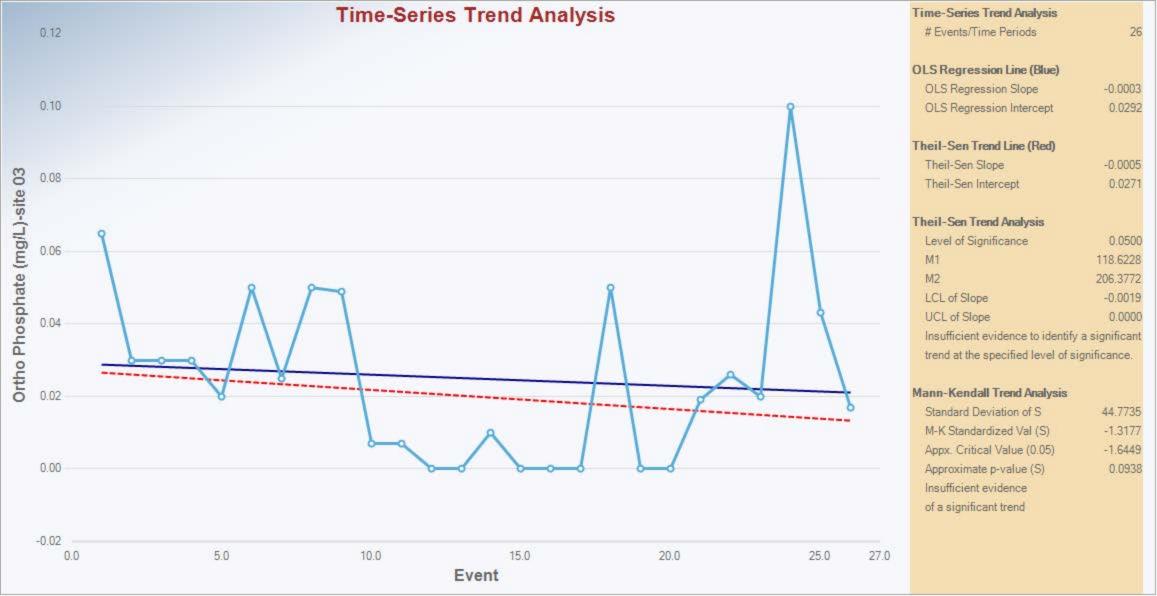


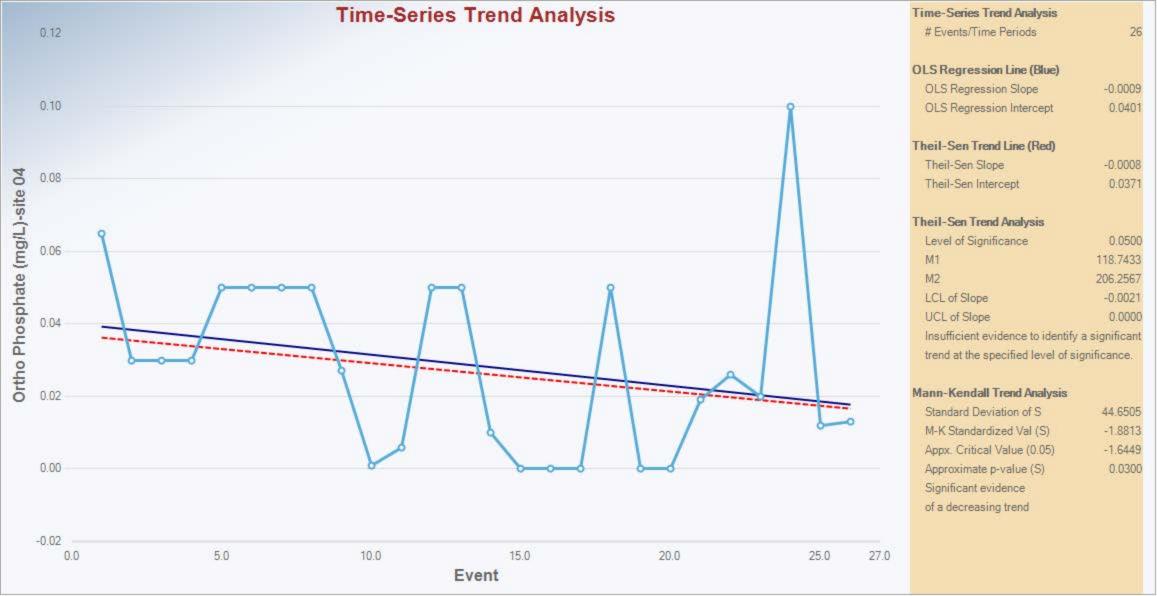


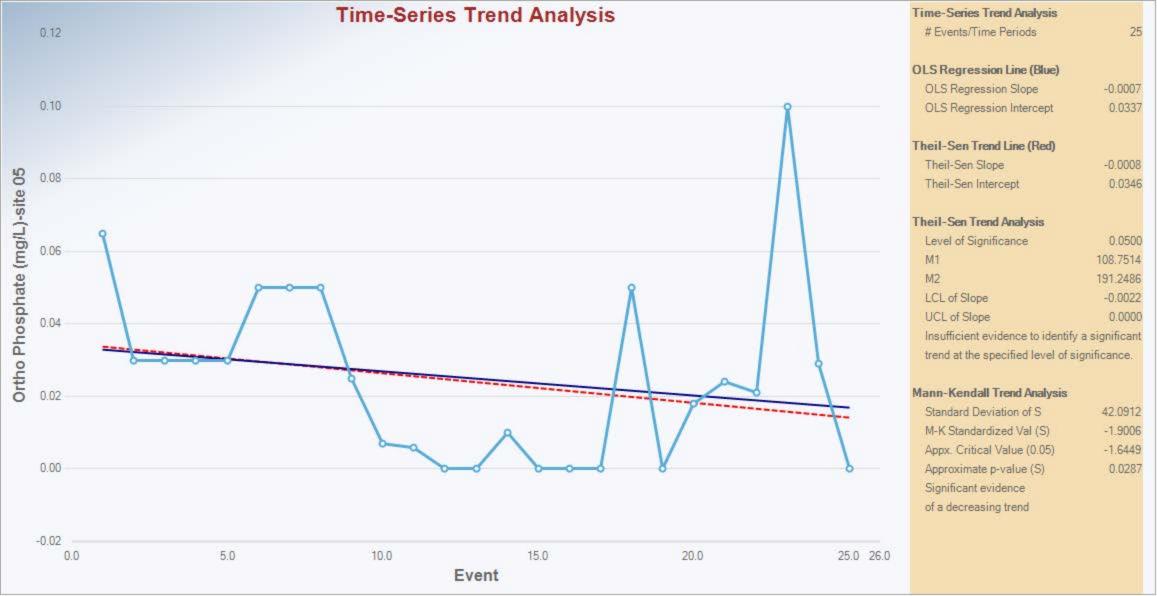


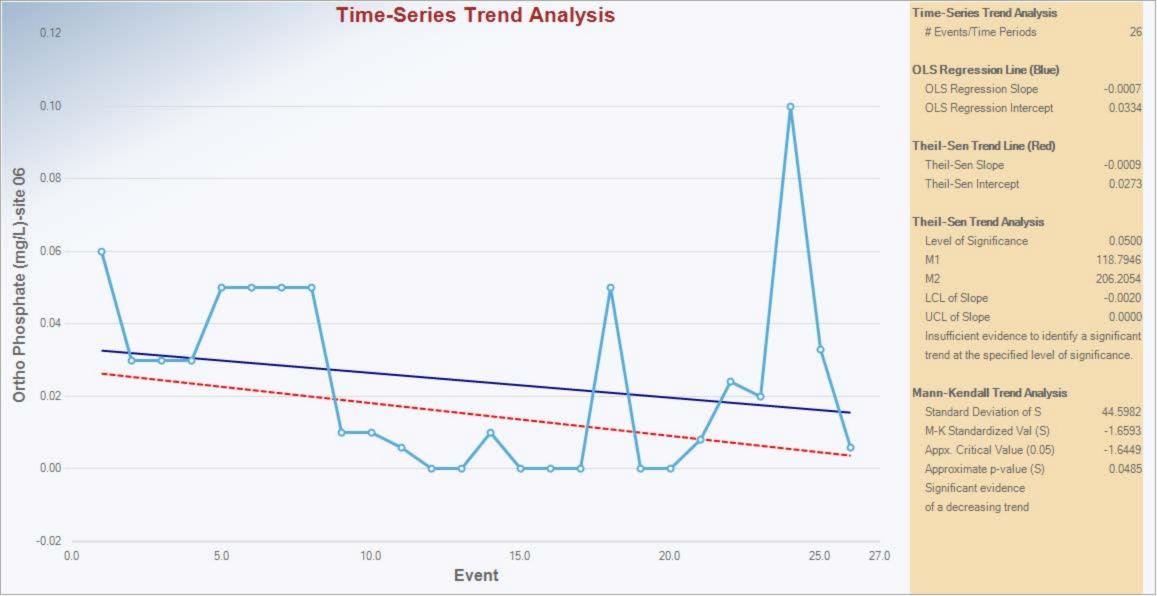


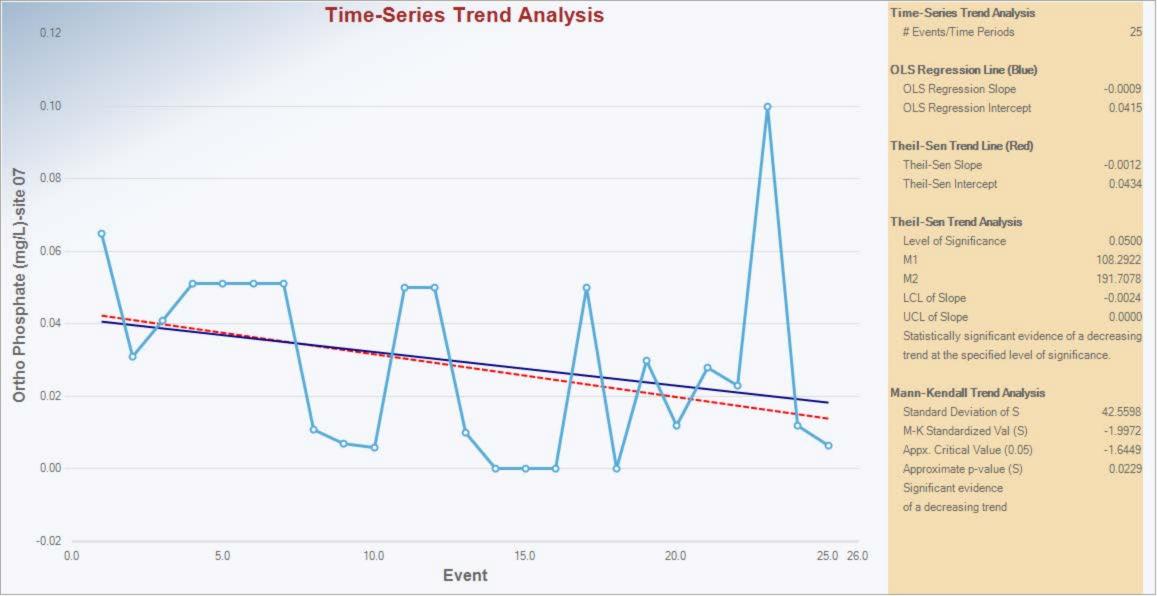


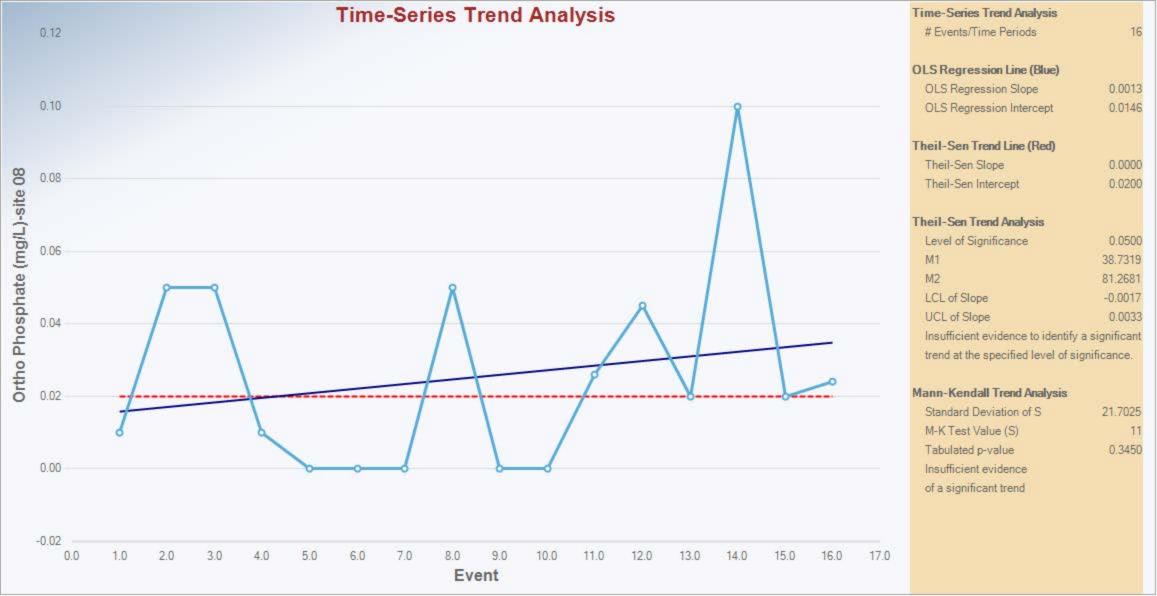


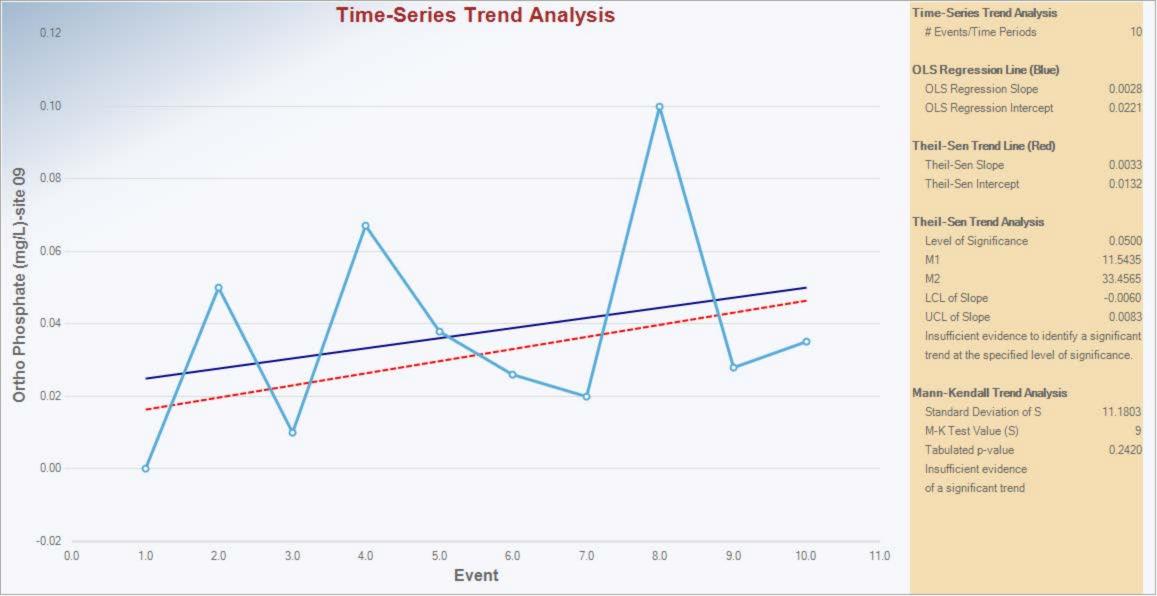




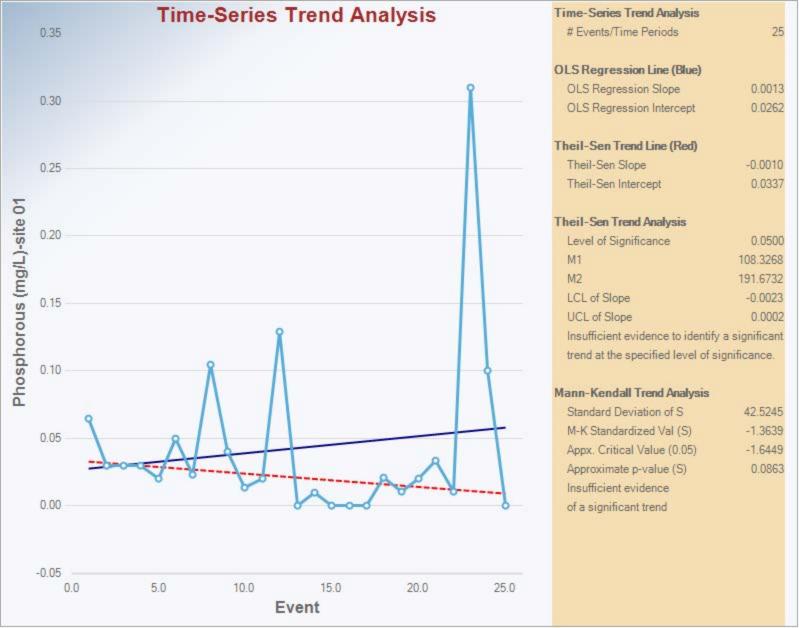


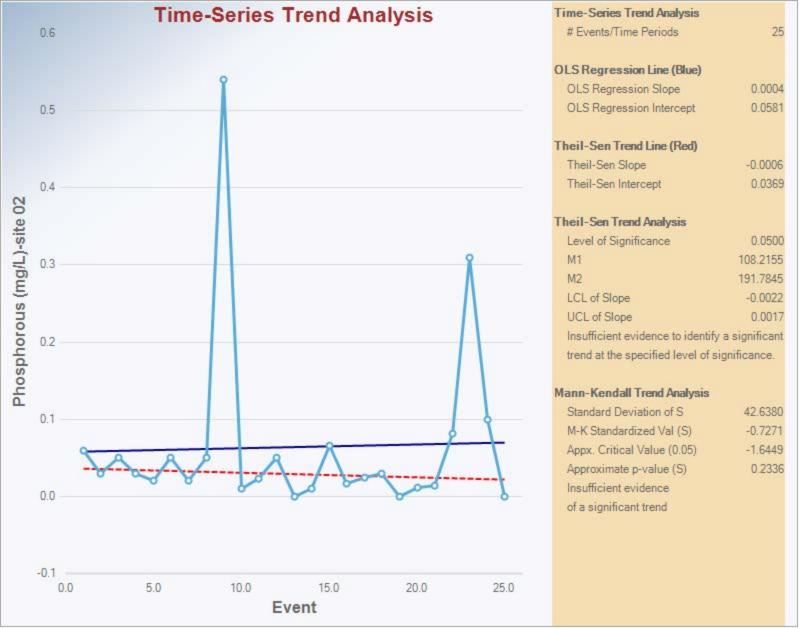


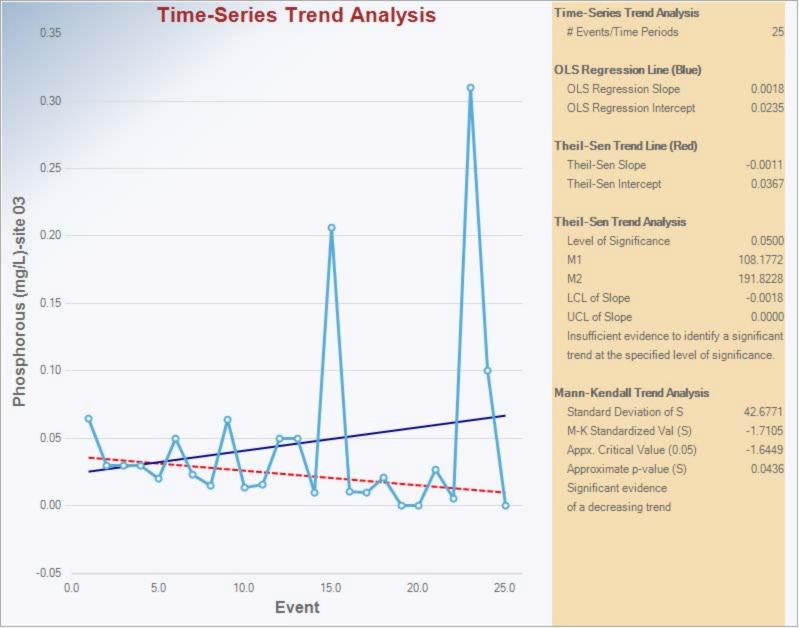


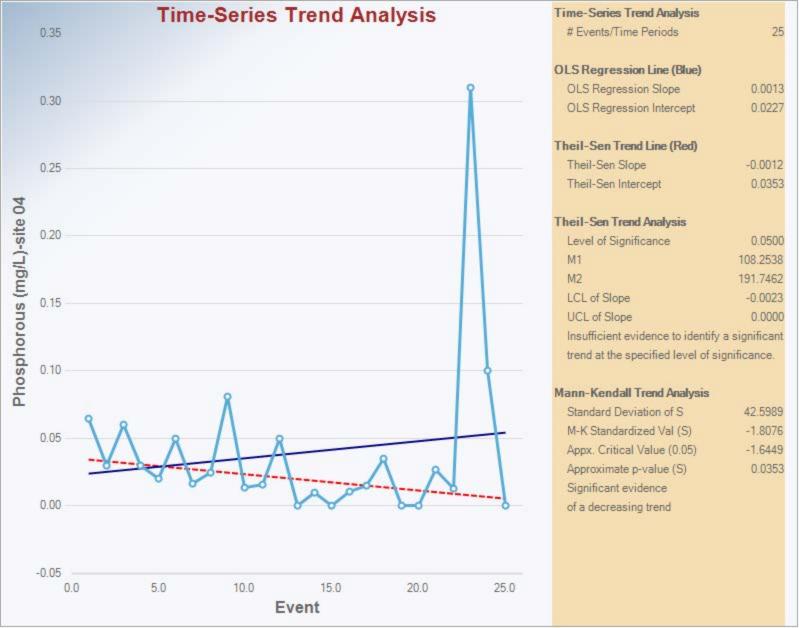


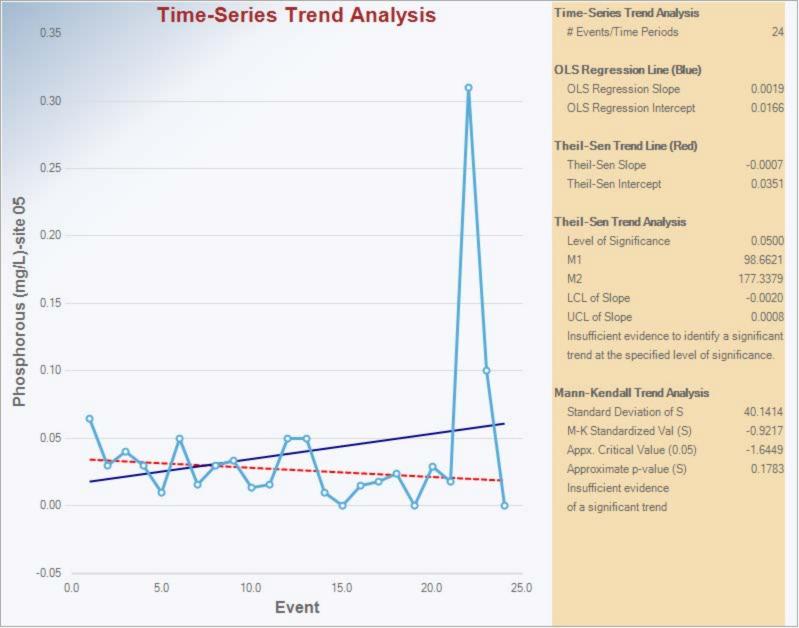


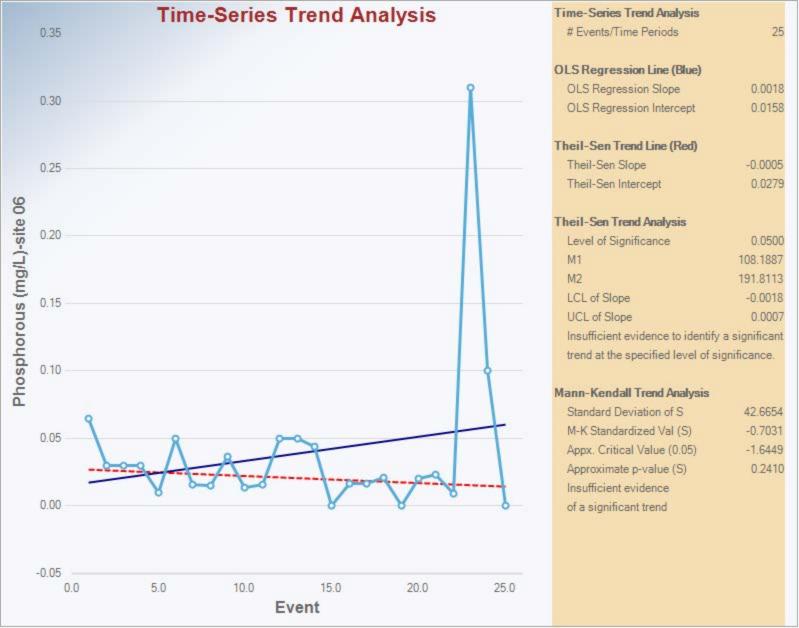


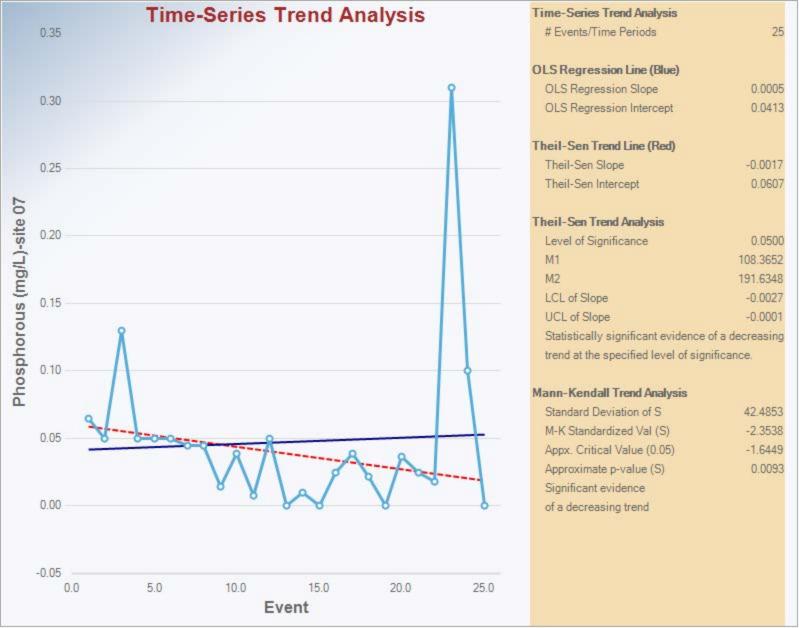


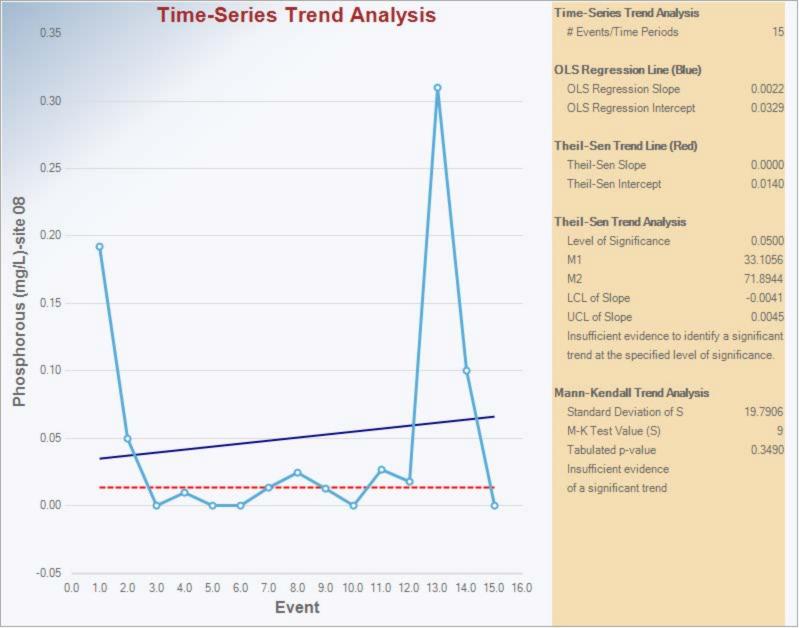


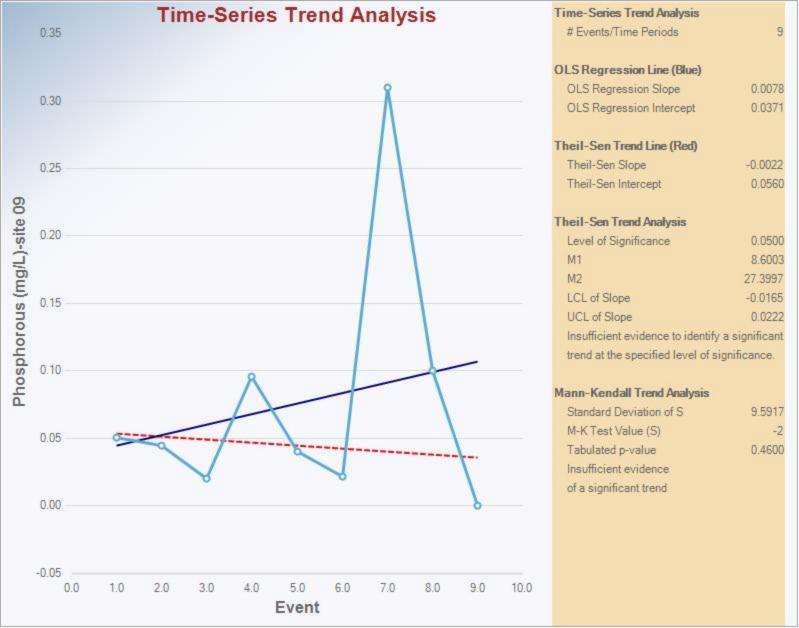




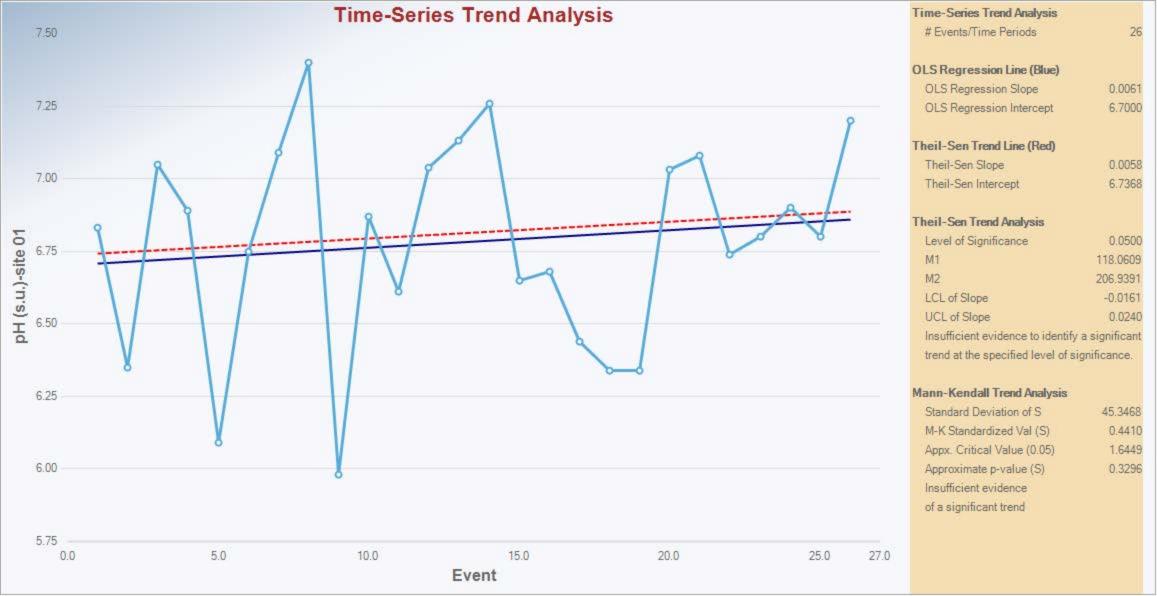


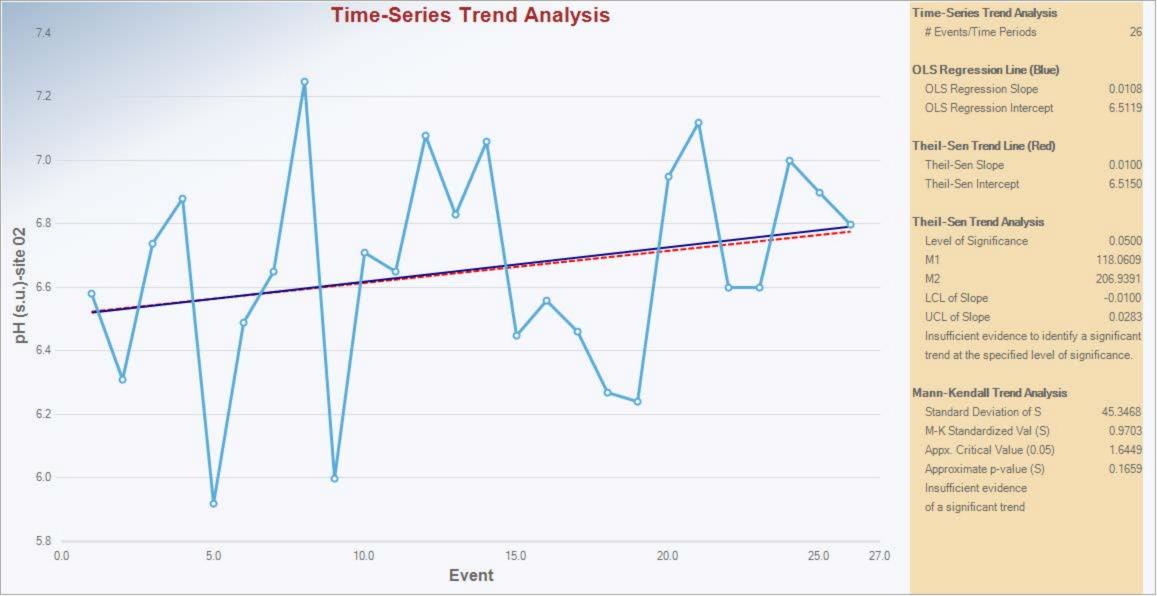


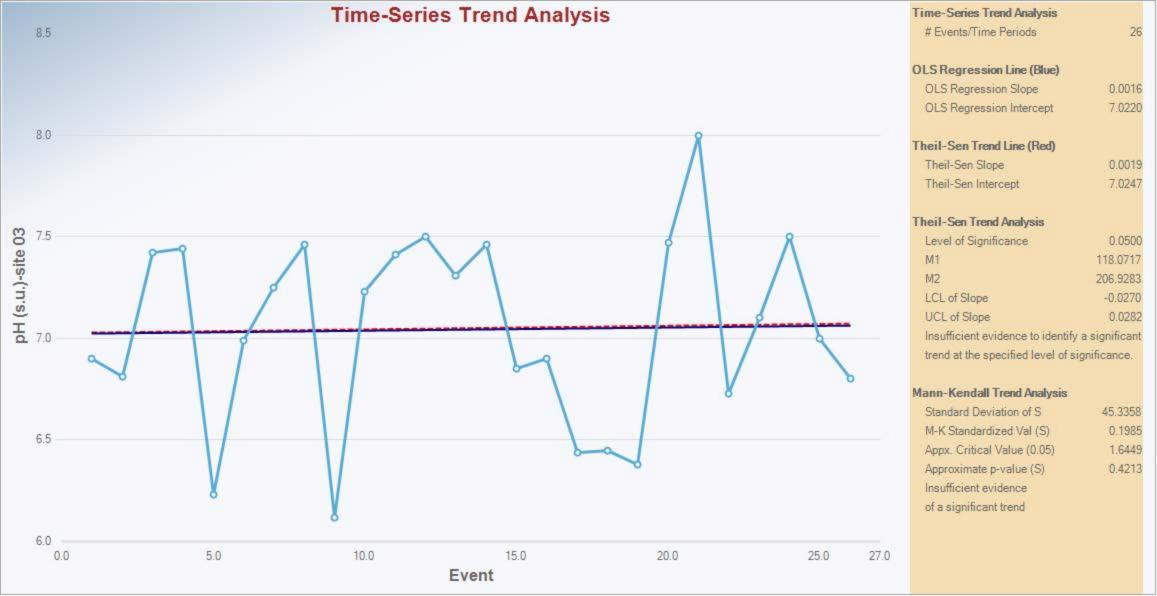


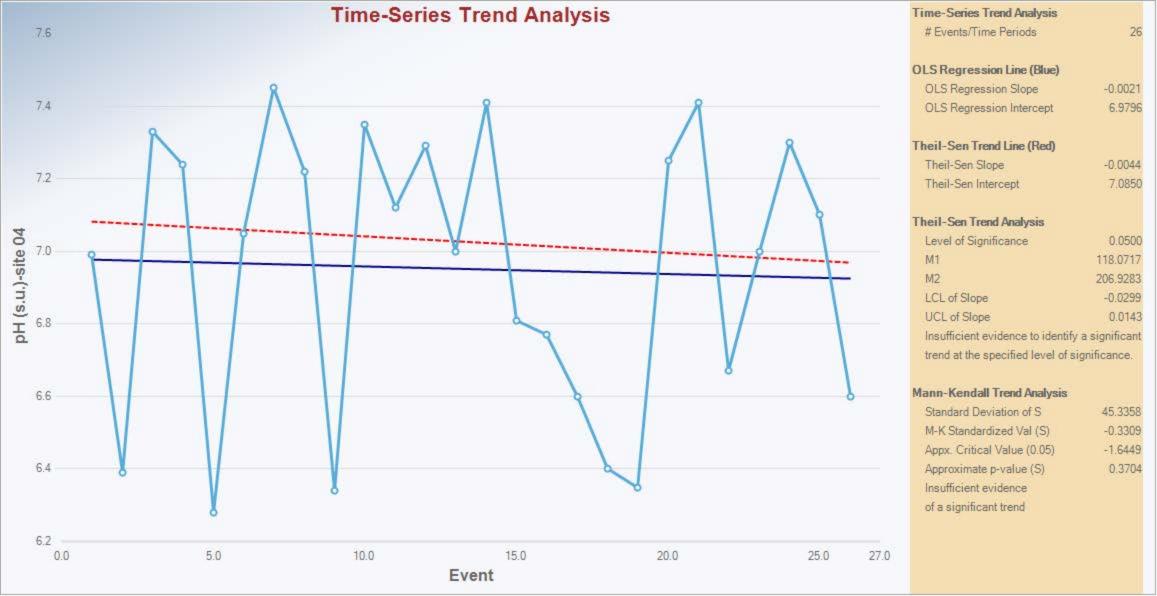


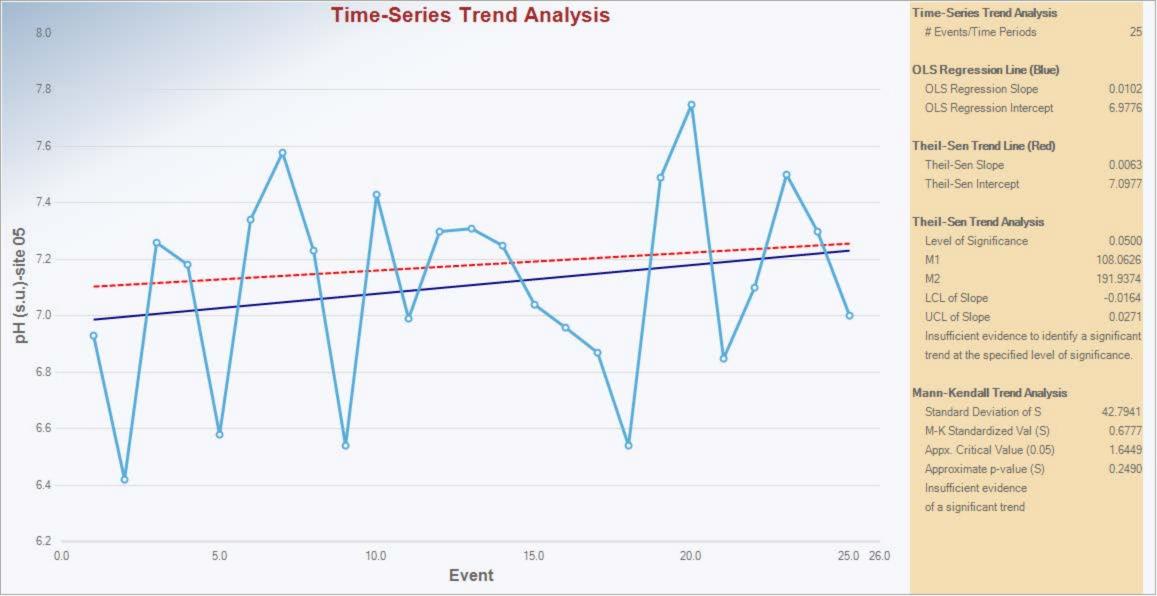


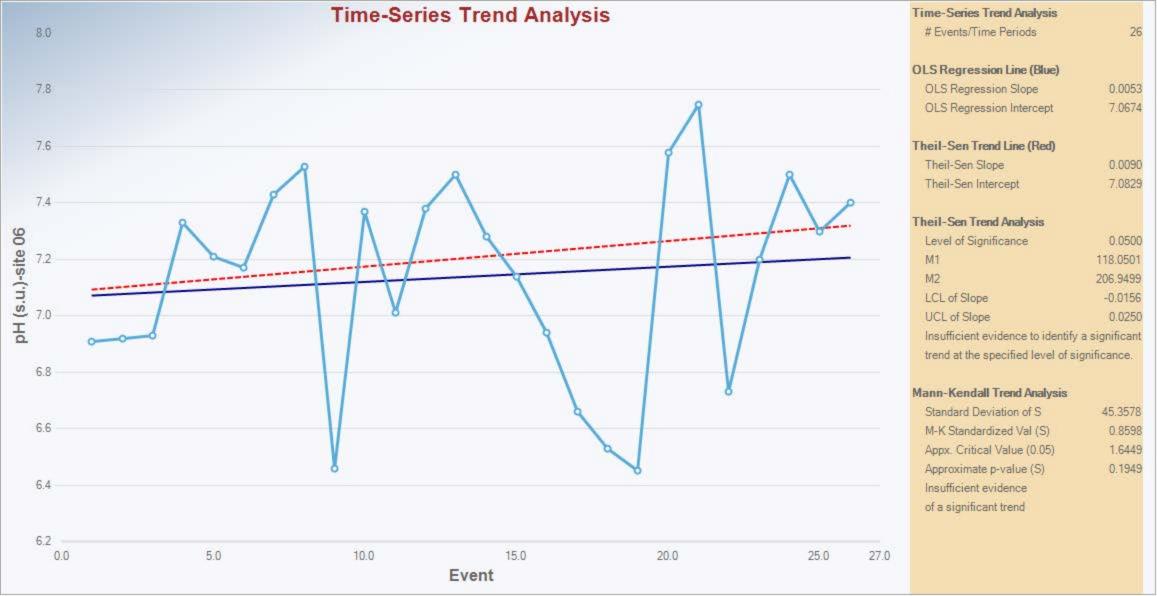


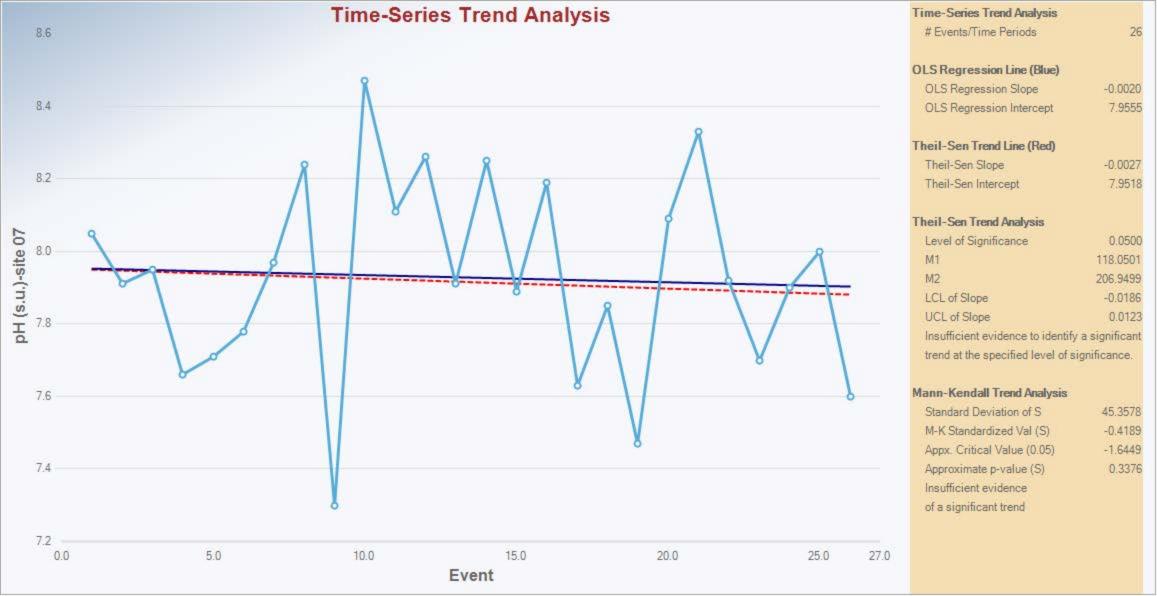


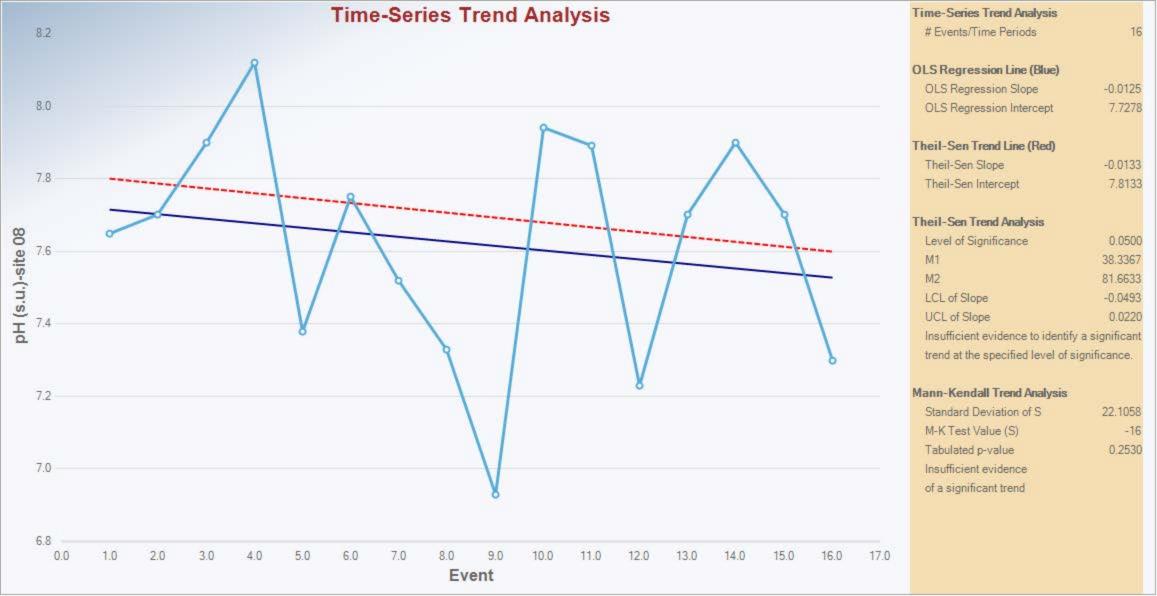


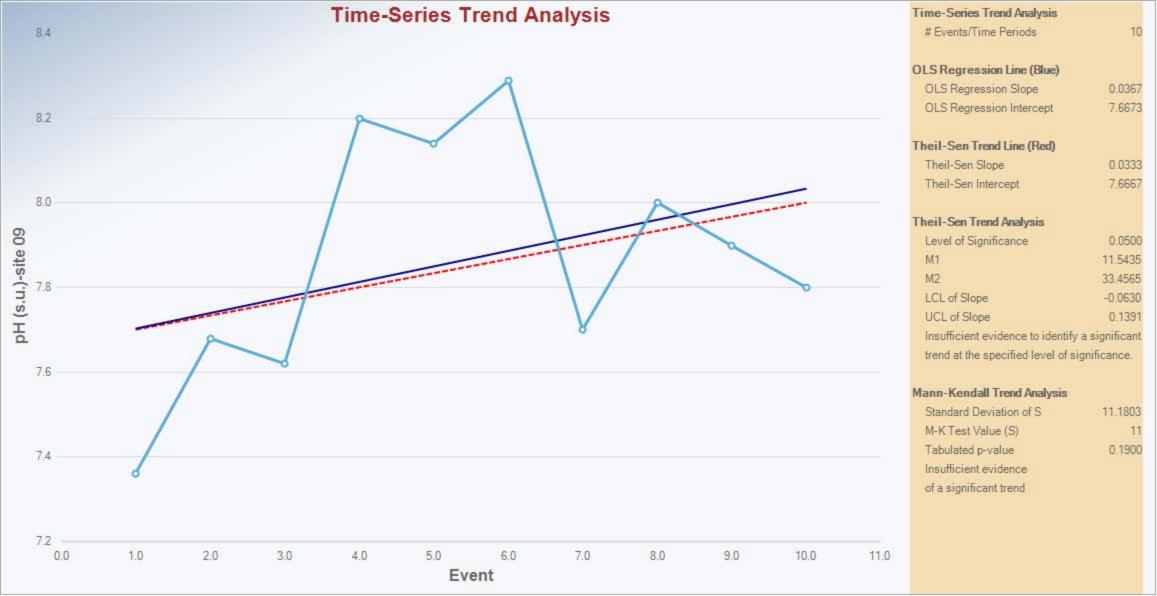




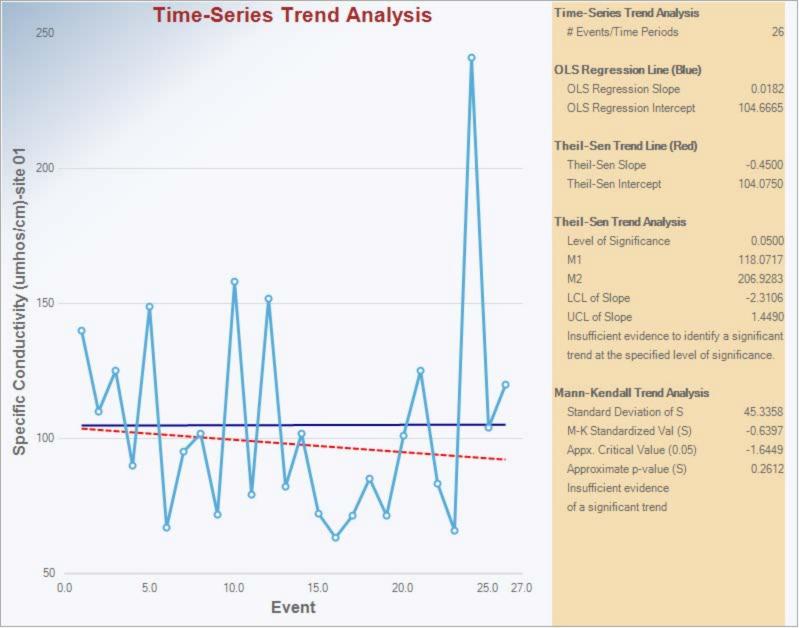


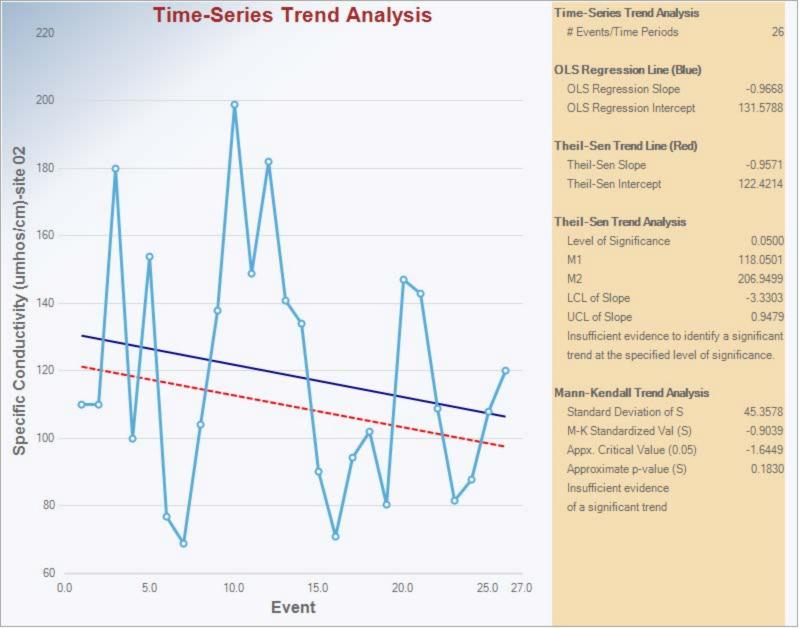


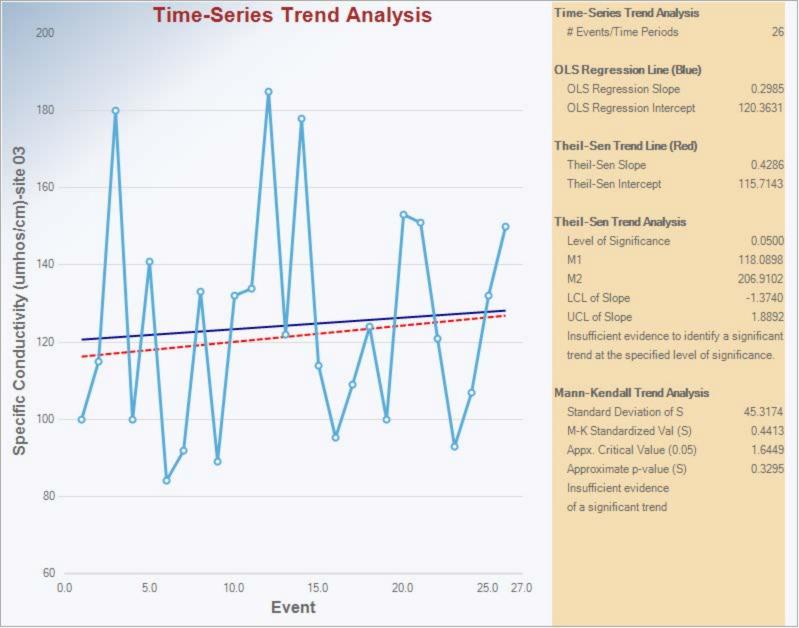


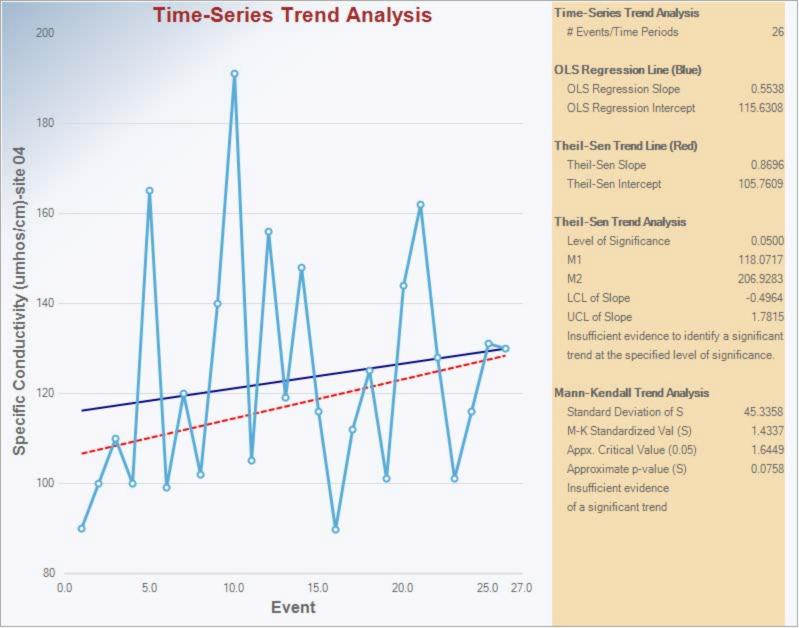


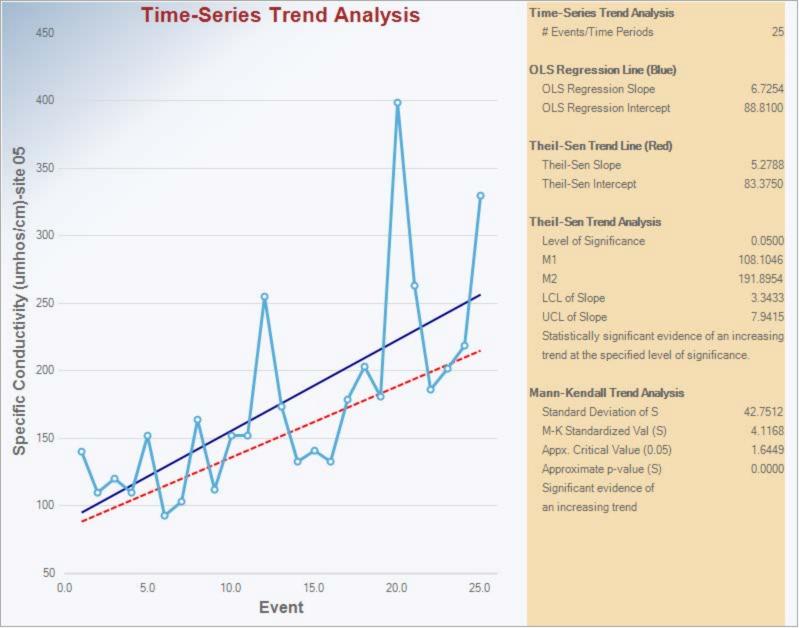


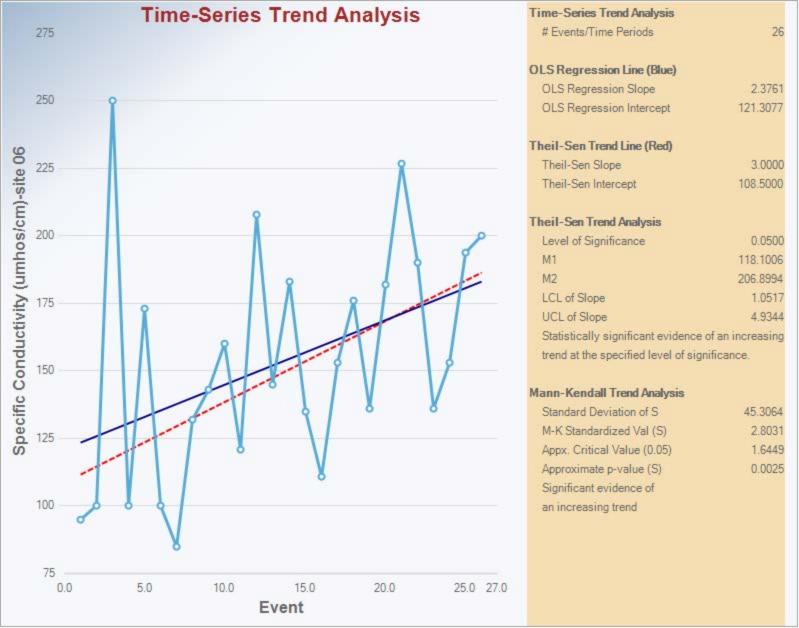


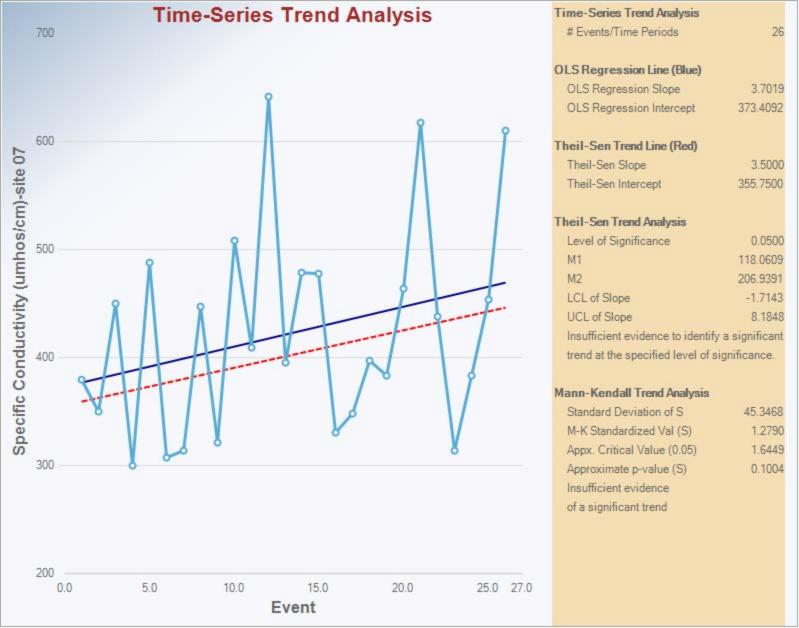






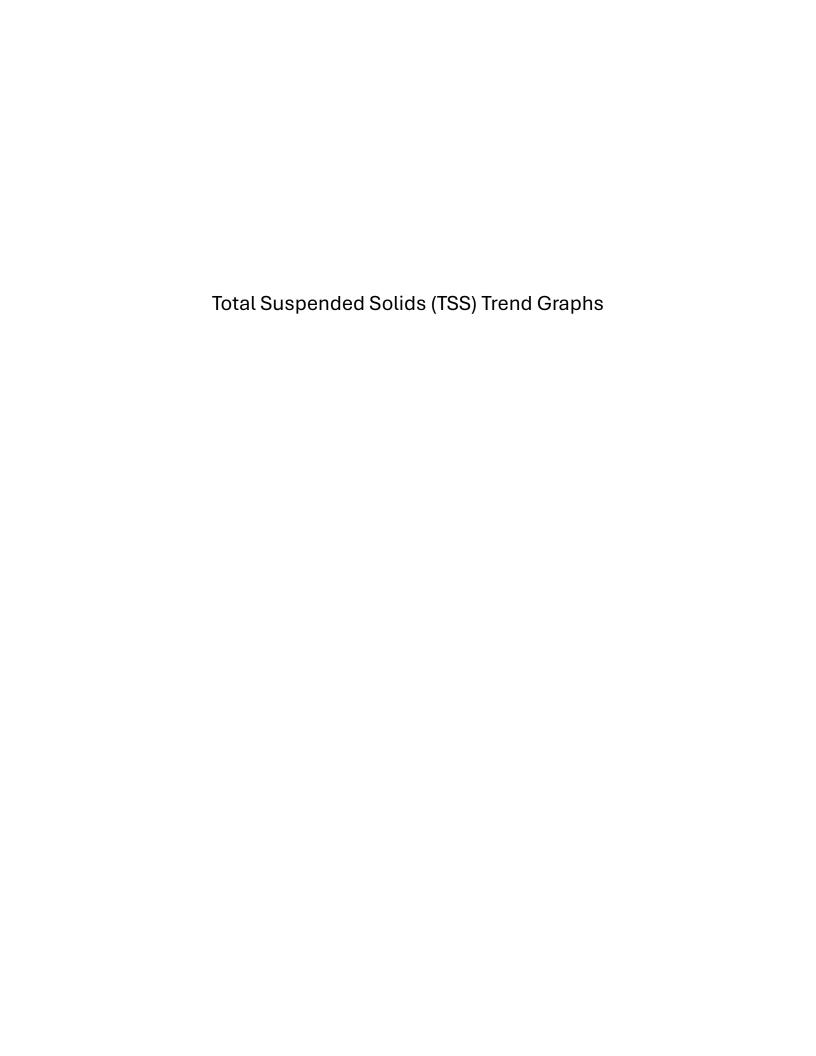


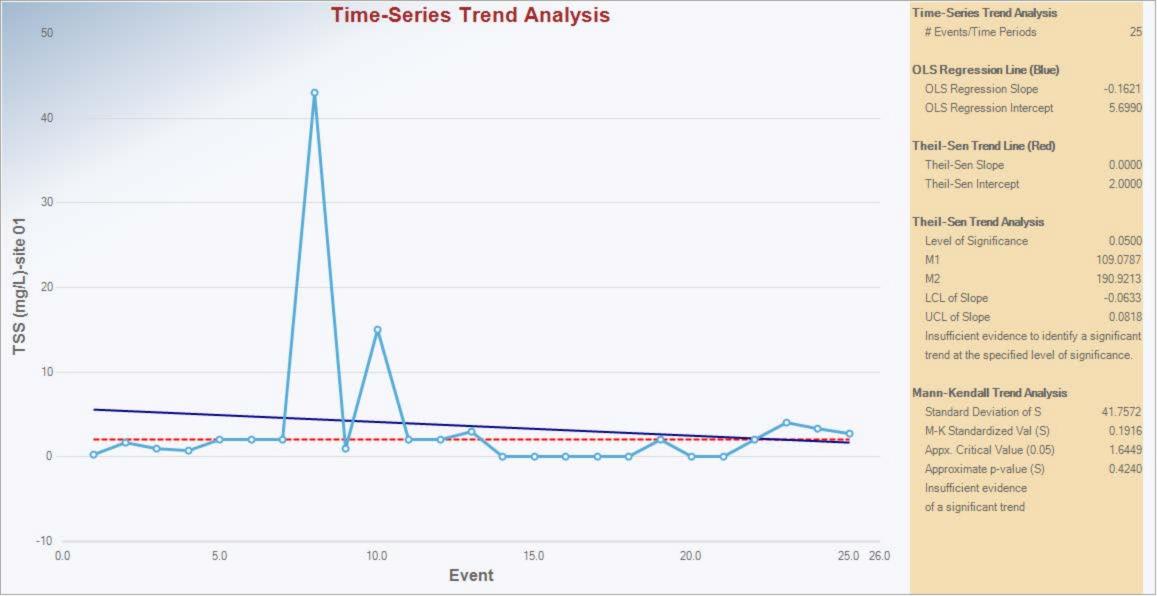


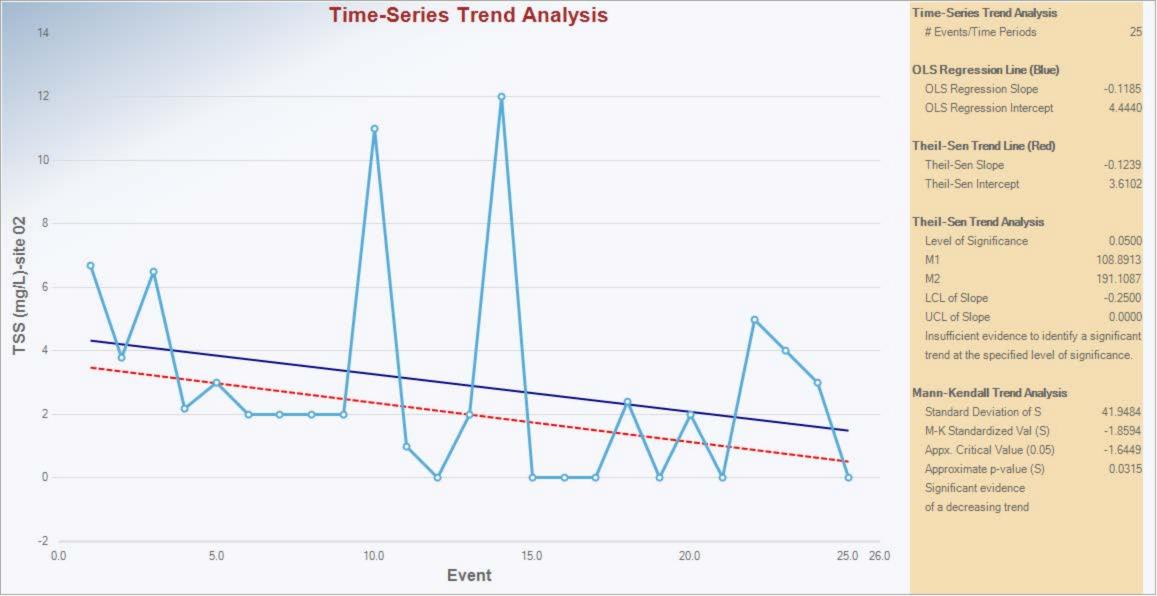


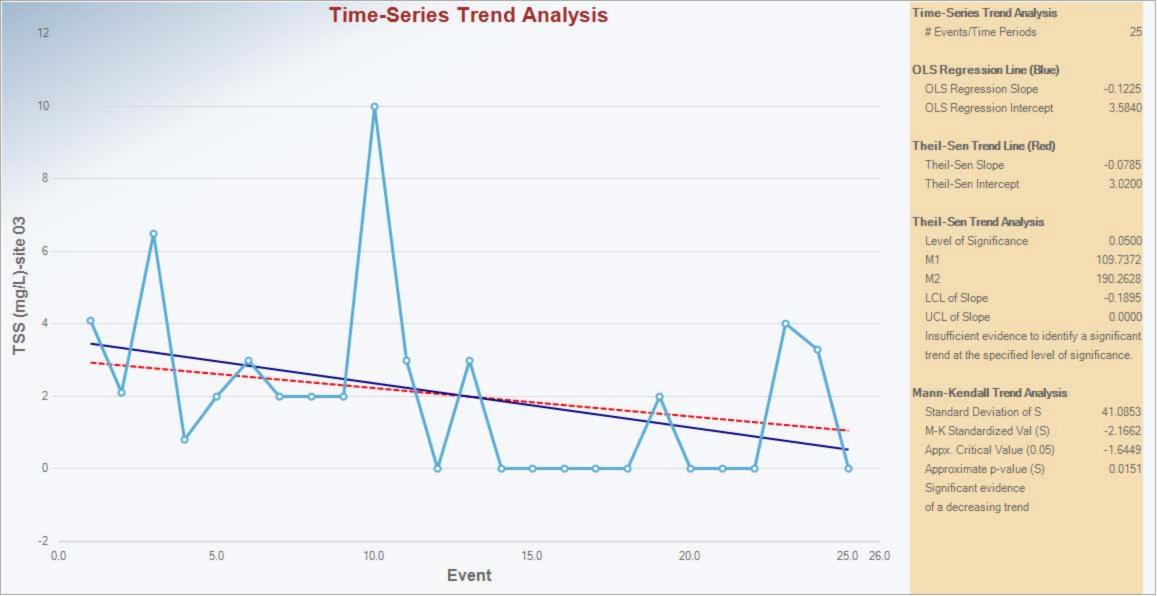


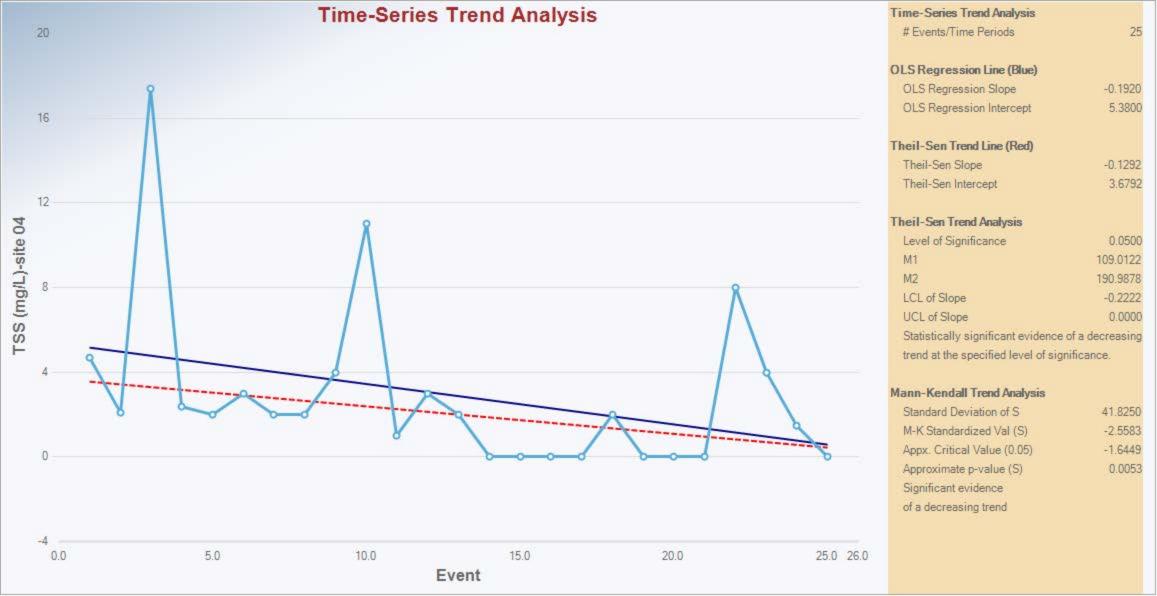


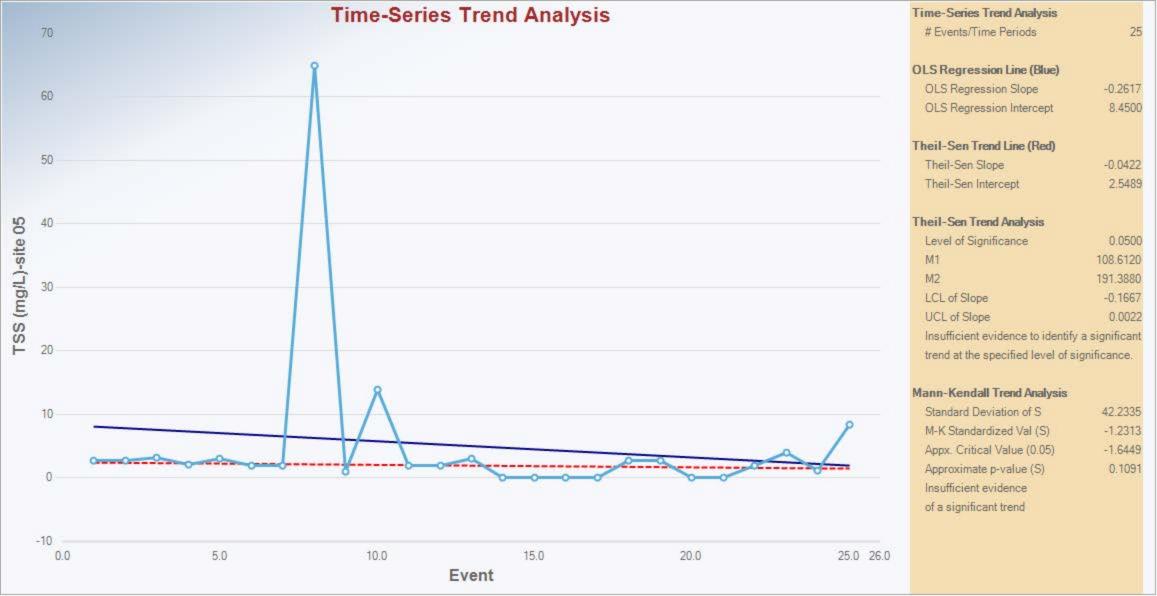


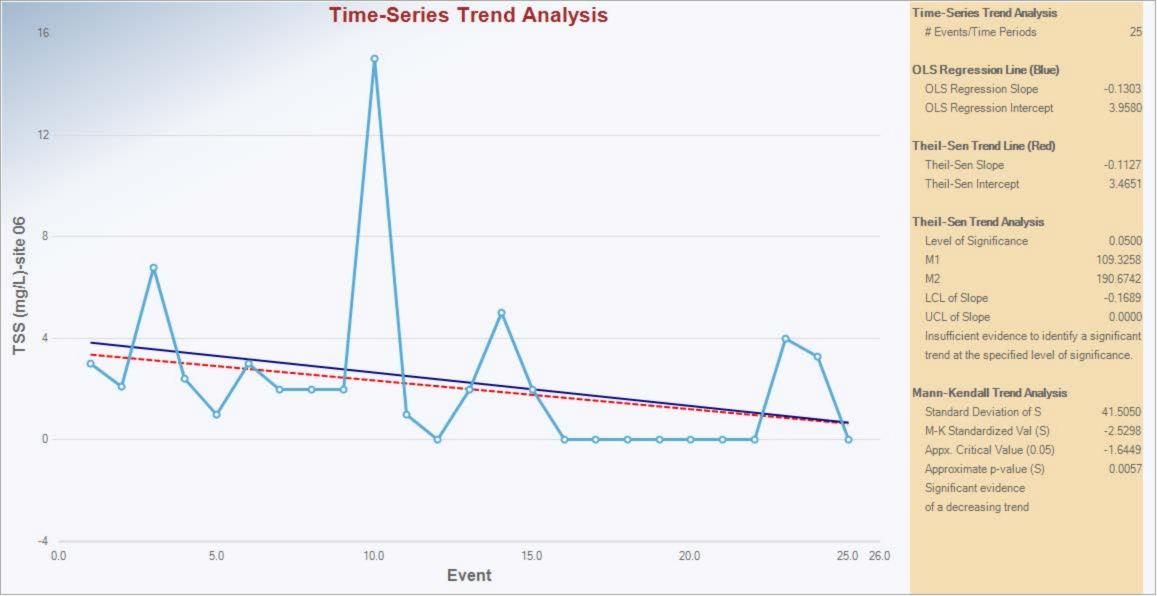


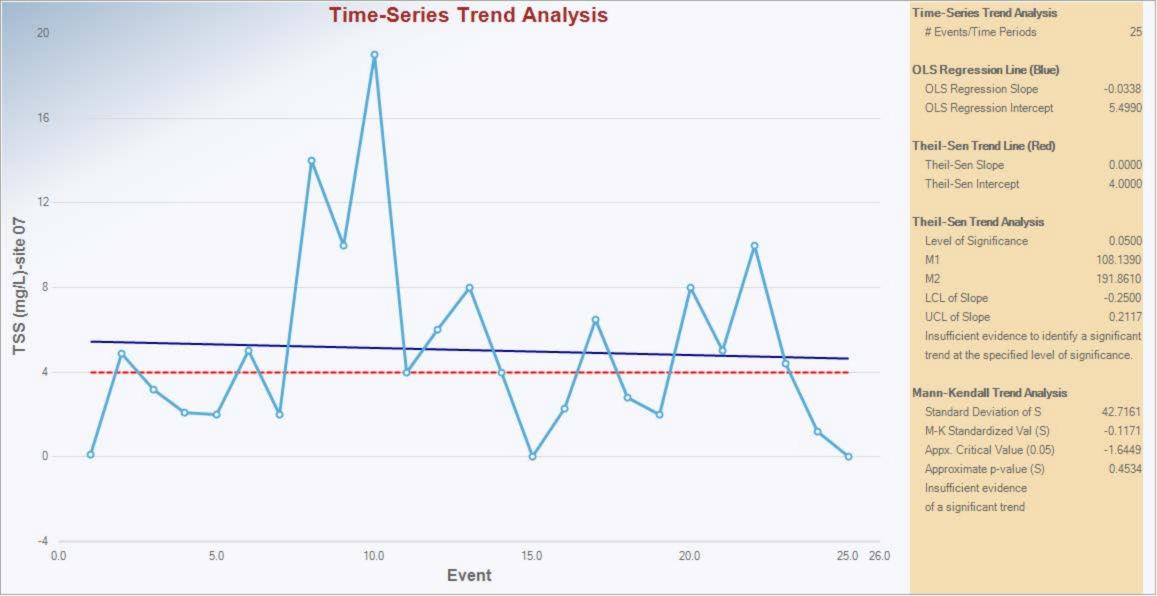


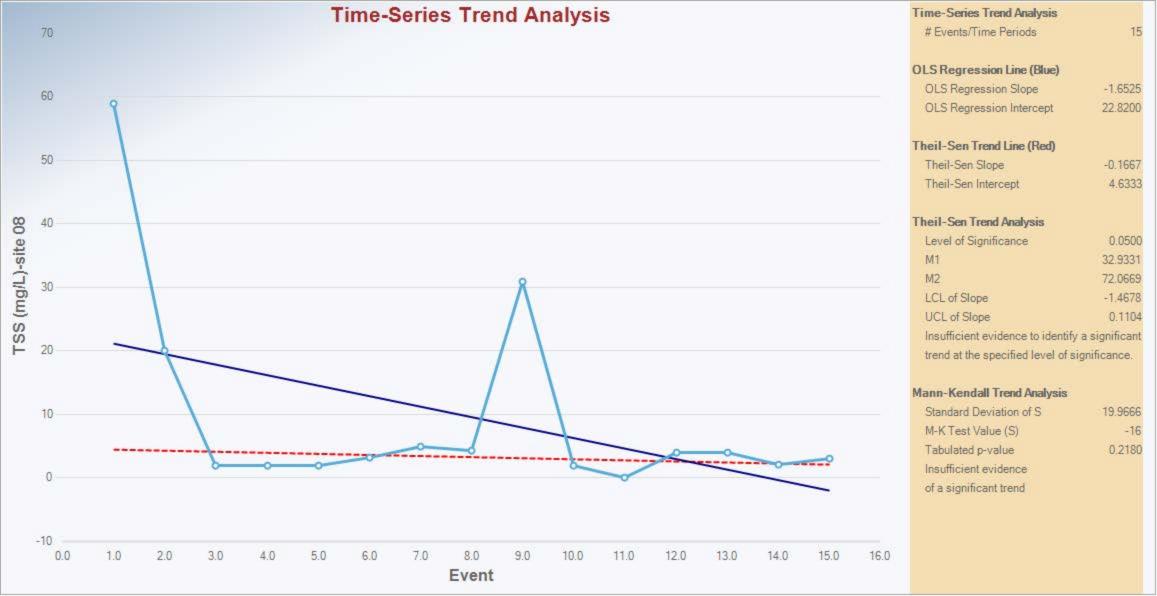


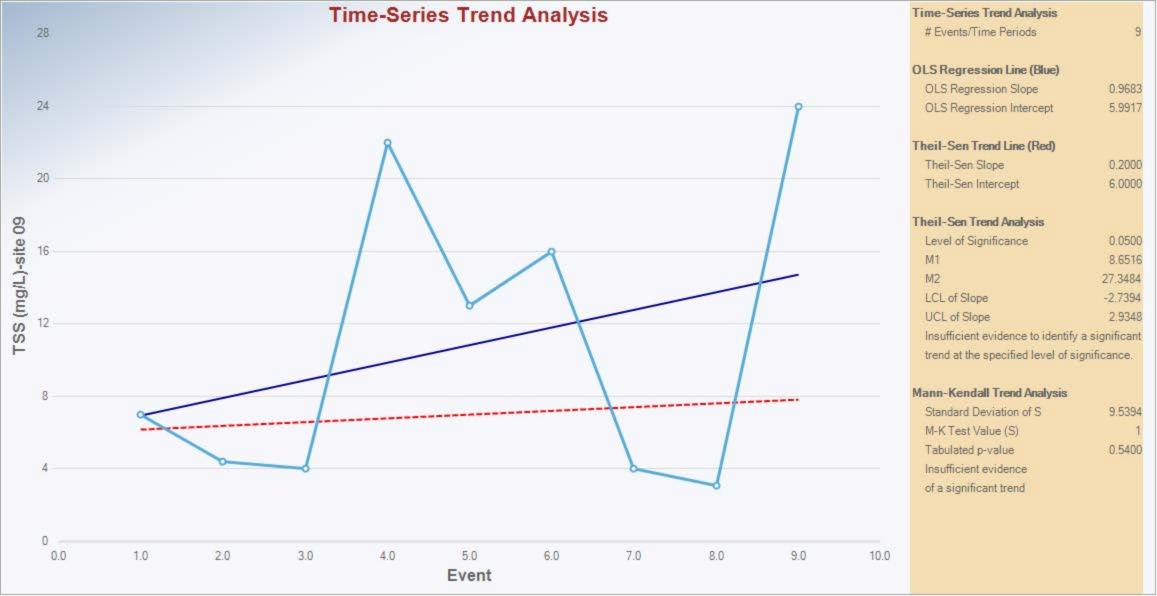










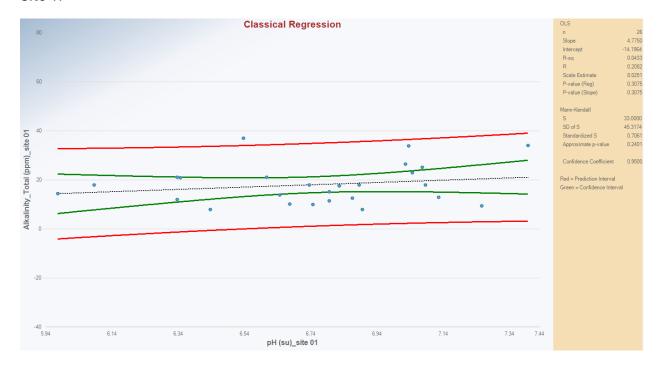


Appendix E

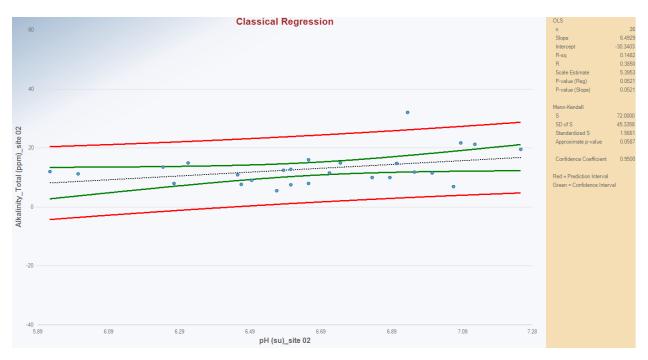
Miscellaneous Statistical Analysis Graphs and Calculations

Alkalinity vs. pH OLS Trend Analysis (Correlation) Graphs

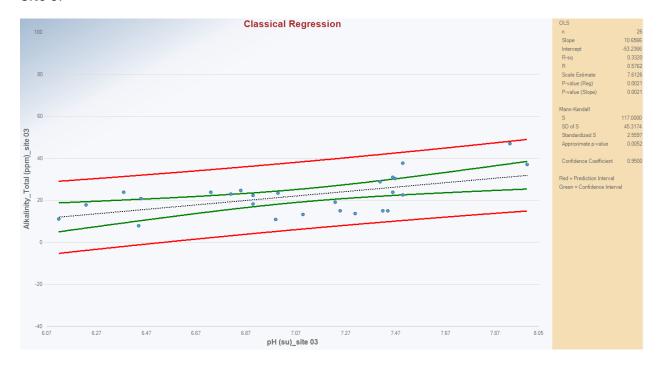
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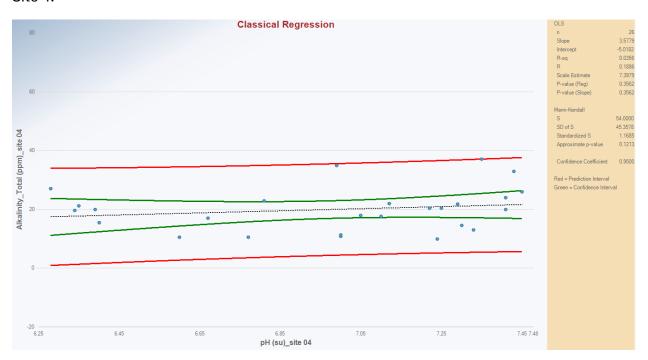
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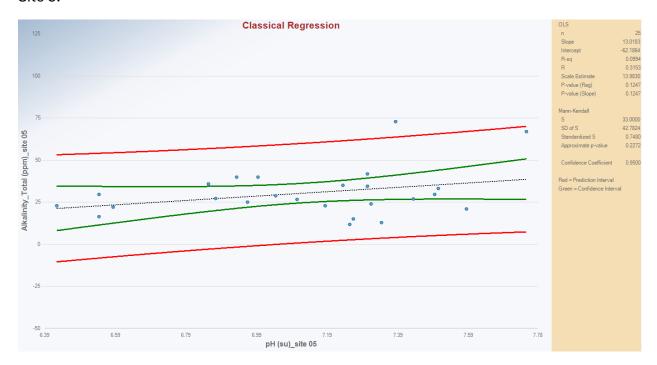
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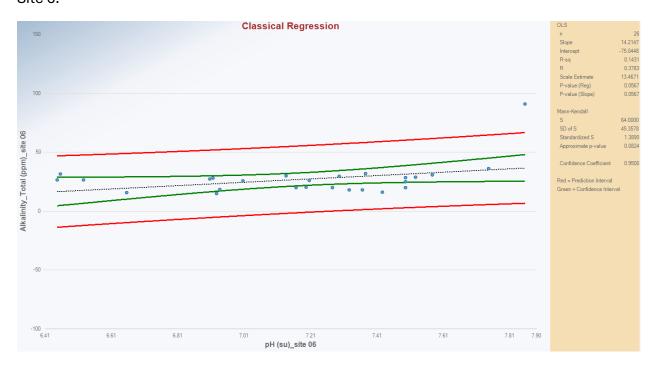
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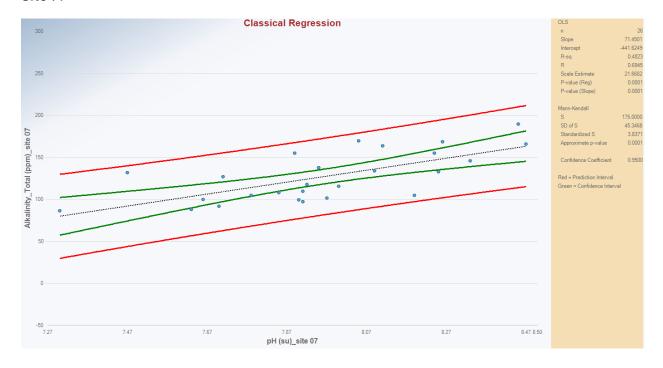
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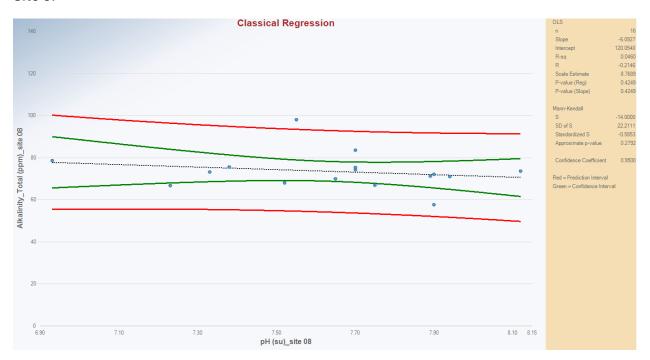
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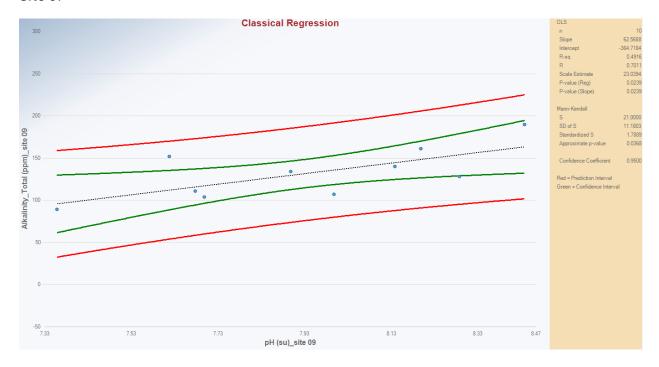
Site 7:



Site 8:



Site 9:

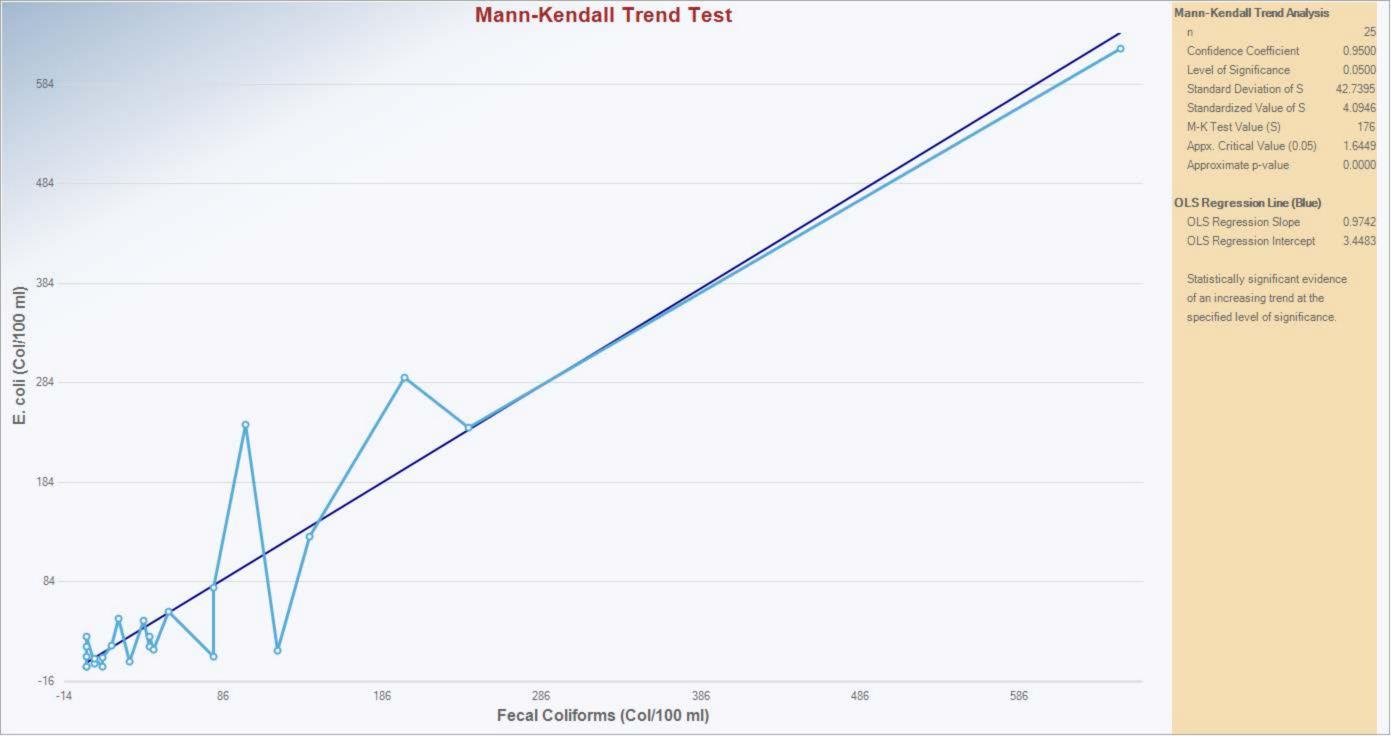


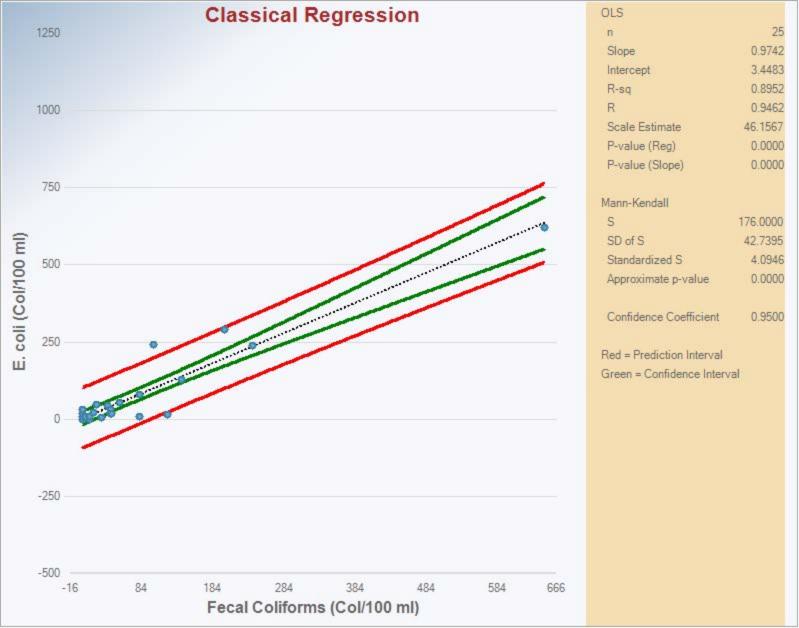
Alkalinity in Paulins Kill vs. Tributaries

	Α	В	С	D	E	F	G	Н	I	J	K	L	M
1													
2				t-Test S	Sample 1 vs	d Full Data	Sets without	NDs					
3													
4				elected Options	ProUCL 5.2								
5			Date/Time o	f Computation									
6				From File	All Data_Pro	UCL_11.12	.2024_a.xls						
7				Full Precision	OFF								
8			Confiden	ce Coefficient									
9				Difference (S)	0.000								
10		Selected Null Hypothesis Sample 1 Mean = Sample 2 Mean (Two Sided											
11			Alternati	ve Hypothesis	Sample 1 M	ean <> Sam	ple 2 Mean						
12													
13													
14		Sample 1 D	ata: Alkalini	ty, Total (mg/L)((paulins kill)								
15		Sample 2 D	ata: Alkalini	ty, Total (mg/L)((tributaries)								
16													
17													
18				Ra	w Statistics								
19							Sample 2						
20		Number of Valid Observation				36	171						
21		Number of Distinct Observation				32	123						
22		Minimum				86.8	5.52						
23					Maximum	190	98						
24					Mean	128.4	26.61						
25					Median	127.5	21.1						
26					SD	29.4	18.96						
27				S	E of Mean	4.901	1.45						
28													
29				Sample 1 vs Sar	mple 2 Two-	Sample t-Te	st						
30													
31		H0: Mean o	f Sample 1 :	= Mean of Samp	ole 2								
32						t-Test		Upper C.Val					
33		Method			DF	Value	t (0.025)	t (0.975)	P-Value				
34		Pooled (Equ	•		205	26.305	-1.972	1.972	0.000				
35				equal Variance)	41.3	19.925	-2.020	2.020	0.000				
36		Pooled SD:											
37			with Alpha =										
38		-		ect H0, Conclude									
39		Welch-Satt	terthwaite: R	eject H0, Conclu	ıde Sample [*]	ı <> Sample	2						
40					II	•							
41				Test of Ed	quality of Va	riances							
42						05:5	1						
43				Variance of		864.6							
44				Variance of	r Sample 2	359.4							
45			. 5-	T 5 ,	. 5-			D					
46			ator DF	Denomina			t Value	P-Value					
47			35 Al I	170)	2.4	406	0.000					
48			with Alpha =										
49		I wo variand	ces are not e	equal									
50													

	Α	В	С	D	E	F	G	Н	I	J	K	L	М		
1			\A/ilo	oven Menn	M/hitnov Son	nnio 1 vo So	mple 2 Com	noricen Teet	for Uncone	or Full Date	Sata without	NDo			
2			VVIIC	oxon-mann-	wnitney San	npie i vs Sa	mpie 2 Com	parison Test	tor Uncens	or Full Data	Sets Without	NUS			
3			Llear Salar	cted Ontions											
4		User Selected Options Date/Time of Computation From File From File All Data_ProUCL_5.2 12/13/2024 5:57:50 PM From File Confidence Coefficient Substantial Difference Selected Null Hypothesis Alternative Hypothesis Sample 1 Mean/Median = Sample 2 Mean/Median Sample 1 Data: Alkalinity, Total (mg/L)(paulins kill) Sample 2 Data: Alkalinity, Total (mg/L)(tributaries) Raw Statistics Sample 1 Sample 2 Sample 2 Sample 2 Sample 2 Sample 3													
5		Dat	e/ Time of Co												
6			Ful			JOOL_11.12	.2024_4.813								
7															
8															
9		Se													
10															
11 12				71											
13															
14		Sample 1 D	ata: Alkalini	ty, Total (mg	/L)(paulins k	cill)									
15		Sample 2 D	ata: Alkalini	ty, Total (mg	/L)(tributarie	es)									
16															
17		Raw Statistics													
18		Sample 1 Sample 2													
19			Number	r of Valid Ob	servations	36	171								
20			Number of	f Distinct Ob	servations	32	123								
21					Minimum	86.8	5.52								
22					Maximum	190	98								
23					Mean	128.4	26.61								
24					Median	127.5	21.1								
25					SD	29.4	18.96								
26				S	E of Mean	4.901	1.45								
27															
28				Wilcoxon-Ma	ann-Whitney	(WMW) Tes	st								
29		110-14(1)			///	£0l- 0									
30		HU: Mean/N	ledian of Sa	mpie i = Me	an/Median c	or Sample 2									
31			Sor	malo 1 Dank	Sum W-Stat	6814									
32			Sai		VMW U-Stat										
33			St		VMW U-Stat										
34					Mean (U)										
35				SD	(U) - Adj ties										
36		Lower Appr	roximate U-S		/alue (0.025)										
37					/alue (0.975)										
38					sted for Ties)										
40															
41		Conclusion	with Alpha =	= 0.05											
42		Reject HO), Conclude	Sample 1 <	> Sample 2										
43															
44		P-Value <	< alpha (0.05	5)											
45															
		ı									<u>I</u>				

Fecal Coliforms and E. coli





	Α	ВС	D E	F	G	Н		J	K	L	М
2			t-Test Sample 1 v	s Sample 2 (Comparison fo	or Uncensore	ed Full Data S	Sets without I	NDs		
3											
4		User Selecte	·								
5		Date/Time of Com		2 1/1/2025 1							
6				put_E.coli_F	C Data_a.xls						
7			recision OFF								
8		Confidence Co									
9		Substantial Differe	` '								
10		Selected Null Hyp	·		ple 2 Mean (T	wo Sided Alte	ernative)				
11		Alternative Hyp	oothesis Sample 1	Mean <> San							
12											
13											
14		Sample 1 Data: E. coli (Col									
15		Sample 2 Data: Fecal Colif	orms (Col/100 ml)								
16											
17											
18			Raw Statisti								
19				Sample 1	Sample 2						
20		Number o	f Valid Observations	25	25						
21		Number of D	istinct Observations	20	17						
22			Minimum	0	0						
23			Maximum	620	650						
24			Mean	77.99	76.52						
25			Median	20	36						
26			SD	139.6	135.6						
27			SE of Mean	27.92	27.11						
28					+						
29		Samp	ole 1 vs Sample 2 Tw	o-Sample t-T	est						
30											
31		H0: Mean of Sample 1 = M	ean of Sample 2								
32				t-Test	Lower C.Va	Upper C.Val					
33		Method	DF	Value	t (0.025)	t (0.975)	P-Value				
34		Pooled (Equal Variance)	48	0.038	-2.011	2.011	0.97				
35		Welch-Satterthwaite (Uneq	ual Variar 48	0.038	-2.011	2.011	0.97				
36		Pooled SD: 137.594	l .		_1						
37		Conclusion with Alpha = 0.0	050								
38		Student t (Pooled): Do No	t Reject H0, Conclud	e Sample 1 =	Sample 2						
39		Welch-Satterthwaite: Do N	Not Reject H0, Conclu	ıde Sample 1	= Sample 2						
40											
41			Test of Equality of \	/ariances							
42											
43		V	ariance of Sample 1	19484							
44		V	ariance of Sample 2	18380							
45							<u> </u>				
46		Numerator DF	Denominator DF	F-Tes	st Value	P-Value					
47		24	24	1	1.06	0.888					
48		Conclusion with Alpha = 0.0	05								
49		Two variances appear to b									
50											
JU											

	B C D E	F	G	H I	J	K	L	М
2	Wilcoxon-Mann-Whitney Sa	mple 1 vs Sa	ample 2 Com	parison Test for Un	censor Full Data	Sets without	NDs	
3								
4	User Selected Options							
5	Date/Time of Computation ProUCL 5.2	2 12/19/2024	4:52:50 PM					
6	From File ProUCL Inp	out_E.coli_F0						
7	Full Precision OFF							
8	Confidence Coefficient 95%							
9	Substantial Difference 0.000							
10	Selected Null Hypothesis Sample 1 M	Selected Null Hypothesis Sample 1 Mean/Median = Sample 2 Mean/N						
11	Alternative Hypothesis Sample 1 M	/lean/Median	<> Sample 2	2 Mean/Median				
12	•							
13								
14	Sample 1 Data: Bacterial quality (Col/100 ml)(fe	ecal coliform	s)					
15	Sample 2 Data: Bacterial quality (Col/100 ml)(e	. coli)						
16								
17	Raw Statistic	cs						
18								
19	Number of Valid Observations	25	25					
20	Number of Distinct Observations	17	20					
21	Minimum	0	0					
22	Maximum							
23	Mean							
24	Median							
25	SD	135.6	139.6					
26	SE of Mean	27.11	27.92					
27				1				
28	Wilcoxon-Mann-Whitney	/ (WMW) Te	st					
29								
30	H0: Mean/Median of Sample 1 = Mean/Median	of Sample 2						
31								
32	Sample 1 Rank Sum W-Sta	t 644						
33	WMW U-Star	t 319						
34	Standardized WMW U-Star	t 0.126						
35	Mean (U)	312.5						
36	SD(U) - Adj ties	51.51						
37	Lower Approximate U-Stat Critical Value (0.025)							
38	Upper Approximate U-Stat Critical Value (0.975)				<u> </u>			
39	P-Value (Adjusted for Ties)	0.899						
40		1	1					
41	Conclusion with Alpha = 0.05							
42	Do Not Reject H0, Conclude Sample 1 = San	nple 2						
43								
44	P-Value >= alpha (0.05)							
45								
40								

Phosphorus and Orthophosphate

	A B C	U			ı	G	П		J	I N	
1	Wilcoxon-Mann-	Whitney Sar	nple 1	vs Sa	mple 2 Com	parison Tes	t for Uncens	or Full Data	Sets without	NDs	
2		Γ									
3	User Selected Options										
4	-				12:50:16 PM						
5	From File	2024.12_P	Data_	a.xls							
6		OFF									
7		95%									
8		0.000									
9	Selected Null Hypothesis	Sample 1 M	ean/N	/ledian	= Sample 2 I	Mean/Media	n (Two Sided	Alternative	·)		
10	Alternative Hypothesis	Sample 1 M	ean/N	/ledian	<> Sample 2	Mean/Medi	an				
11											
12											
13	Sample 1 Data: Phosphate, Ortho (n	ng/L)									
14	Sample 2 Data: Phosphorous (mg/L))									
15											
16	F	Raw Statistic	s								
17			Samı		Sample 2						
18	Number of Valid Obs	servations	199		199						
19	Number of Distinct Obs	servations	37	7	45						
20		Minimum	0		0						
21		Maximum	0.	.1	0.54						
22		Mean	0.	0275	0.048						
23		Median 0.02									1
24		SD	0.	0263	0.077						
25	SE	E of Mean	0.0	0186	0.00546						
26					1						
27	Wilcoxon-Mann-Whitney (WMW) Test										
28											
29	H0: Mean/Median of Sample 1 = Me	an/Median c	f San	nple 2							
30											
31	Sample 1 Rank	Sum W-Stat	3750	08							
32	V	VMW U-Stat	1760	08							
33	Standardized V	VMW U-Stat	-1.	923							
34		Mean (U)	1980)1							
35	SD((U) - Adj ties	114	5							
36	Lower Approximate U-Stat Critical V	'alue (0.025)	-1	.96							
37	Upper Approximate U-Stat Critical Value (0.975) 1.96										
38	P-Value (Adjus	ted for Ties)	0.	0544							
39			1		1	1					
	Conclusion with Alpha = 0.05										
40	Do Not Reject H0, Conclude Sam	ple 1 = Sam	ple 2								
40 41	Do Not Nojoot No, Concidad Cam										1
41	Do Not Nojobi No, Gonolado Gam										
41 42	P-Value >= alpha (0.05)										
41	-										

Α

В

С

D

E

F

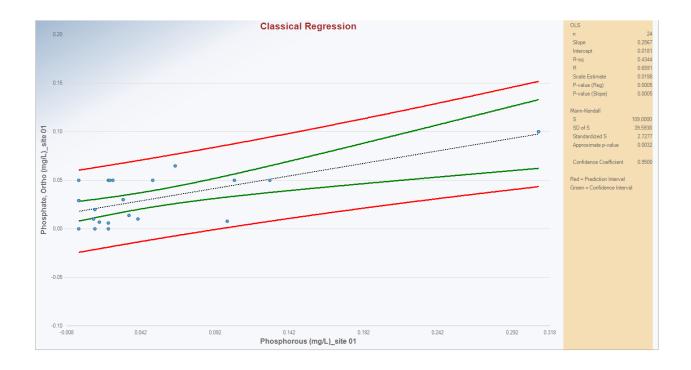
G

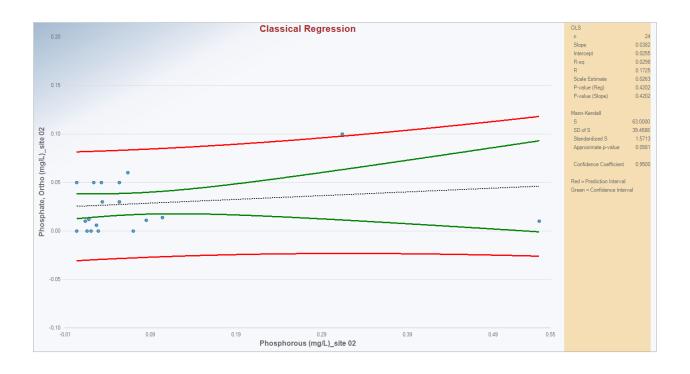
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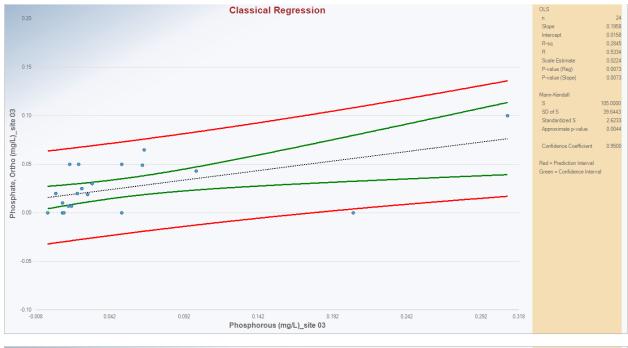
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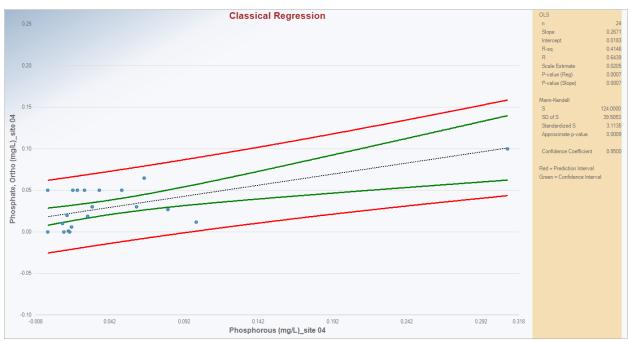
J

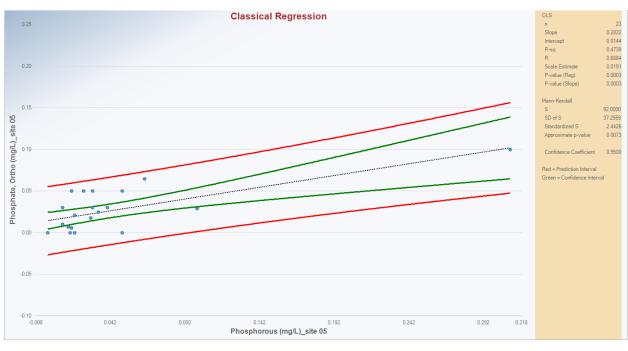
K

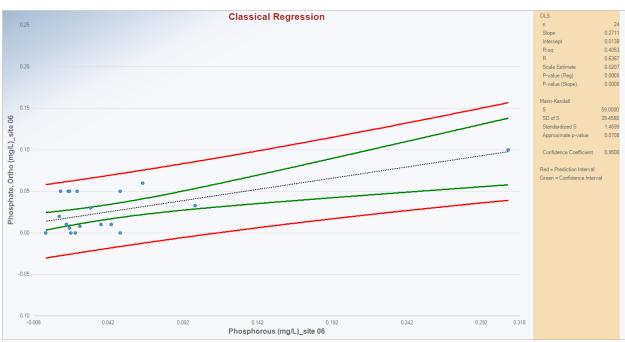


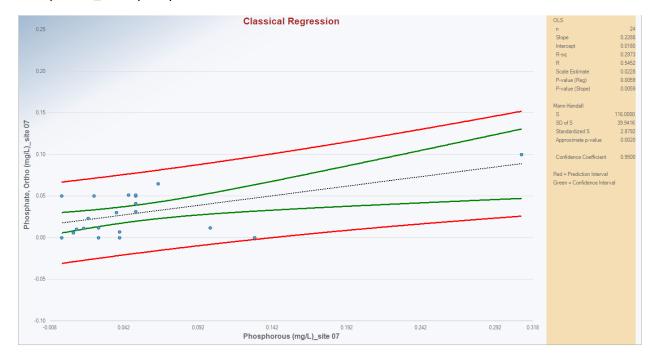


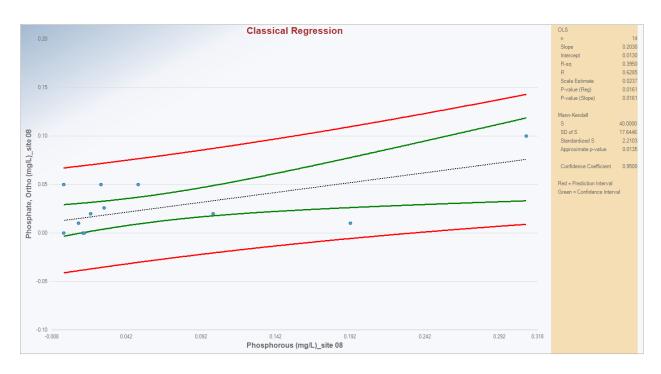


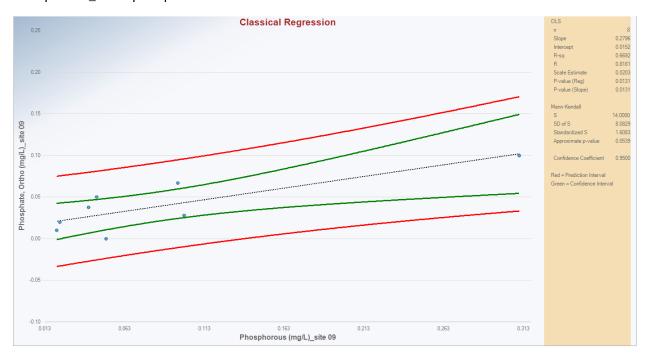












Specific Conductivity in Paulins Kill vs. Tributaries

t-Test Sample 1 vs Sample 2 Comparison for Uncensored Full Data Sets without NDs

User Selected Options

Date/Time of Computation ProUCL 5.2 9/1/2024 6:26:29 PM

From File All Data_ProUCL_04182024_n.xls

Full Precision OFF
Confidence Coefficient 95%

Substantial Difference (S) 0.000

Selected Null Hypothesis Sample 1 Mean = Sample 2 Mean (Two Sided Alternative)

Alternative Hypothesis Sample 1 Mean <> Sample 2 Mean

Sample 1 Data: Specific Conductivity (umhos/cm)(paulins kill) Sample 2 Data: Specific Conductivity (umhos/cm)(tributaries)

Raw Statistics

	Sample 1	Sample 2
Number of Valid Observations	34	165
Number of Distinct Observations	31	115
Minimum	300	63.4
Maximum	642	399
Mean	433.5	141.4
Median	437.5	132
SD	92.88	54.3
SE of Mean	15.93	4.227

Sample 1 vs Sample 2 Two-Sample t-Test

H0: Mean of Sample 1 = Mean of Sample 2

		t-Test	Lower C.Val		
Method	DF	Value	t (0.025)	t (0.975)	P-Value
Pooled (Equal Variance)	197	24.835	-1.972	1.972	0.000
Welch-Satterthwaite (Unequal Variance)	37.8	17.724	-2.024	2.024	0.000

Pooled SD: 62.450

Conclusion with Alpha = 0.050

Student t (Pooled): Reject H0, Conclude Sample 1 <> Sample 2
Welch-Satterthwaite: Reject H0, Conclude Sample 1 <> Sample 2

Test of Equality of Variances

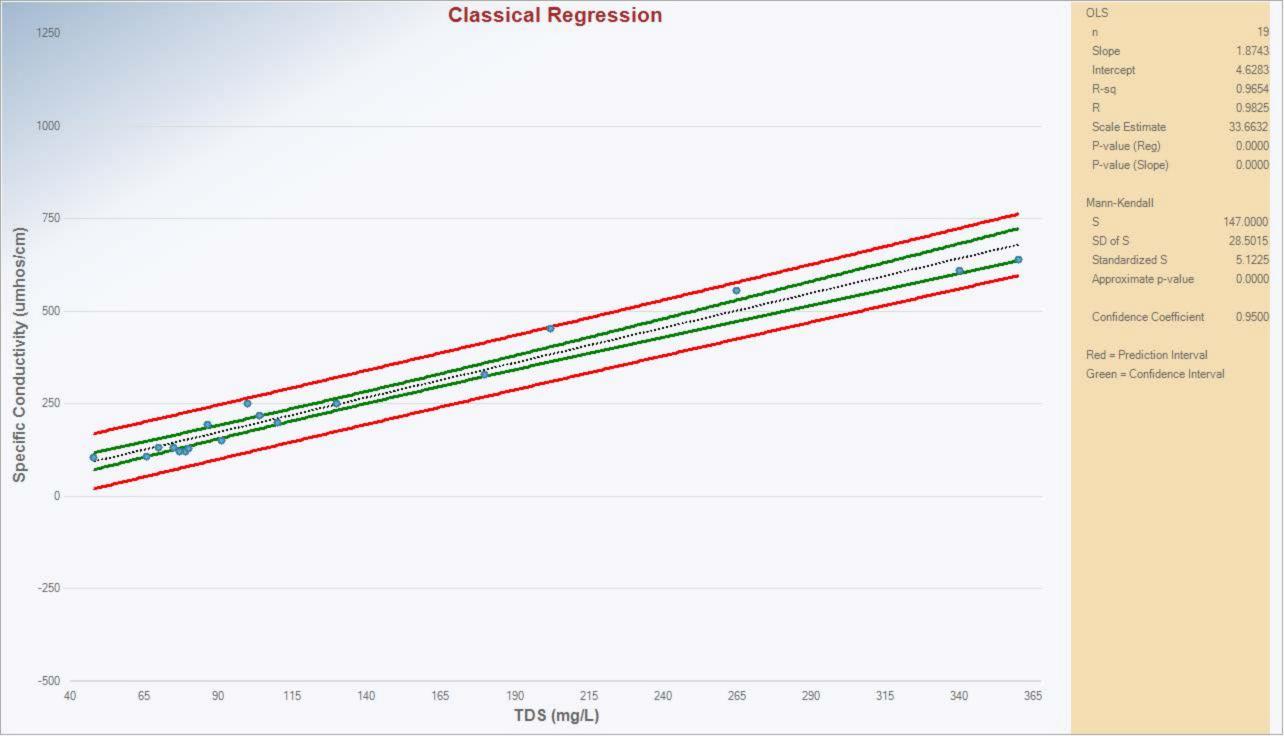
Variance of Sample 1 8627 Variance of Sample 2 2949

Numerator DF	Denominator DF	F-Test Value	P-Value
33	164	2.926	0.000

Conclusion with Alpha = 0.05

Two variances are not equal

Specific Conductivity and TDS Regression Analysis



TDS from Specific Conductivity and TDS Analysis

	Α	В	С	D	Е	F	G	Н	l	J	K	L	M			
1																
2				t-Test	Sample 1 vs	for Uncensor	red Full Data	3 Sets withou	ut NDs							
3																
4				cted Options												
5		Date	te/Time of Co	-	ProUCL 5.2											
6				From File		CL_Input_Sp	ec.Cond_TD	S_a.xls								
7				I Precision	OFF											
8			Confidence (95%											
9			bstantial Diffe		0.000											
10			elected Null F		-	Mean = Sample 2 Mean (Two Sided Alternative) Mean <> Sample 2 Mean										
11			Alternative F	lypothesis	Sample 1 M	ean <> Sam										
12								Τ	T	ı	1					
13		0 1 1 5														
14		-	ata: TDS (m		OE) ((I)											
15		Sample 2 D	ata: TDS fro	m SC(SC*0	.65) (mg/L)											
16																
17			Raw Statistics													
18						Sample 1	Sample 2									
19			Numbo	er of Valid Ob		19	19									
20				of Distinct Ob		18	16									
21			- Nullibel 0	- DISHITICE OD	Minimum	48	67.6									
22					Maximum	360	416									
23					Mean	136.5	169.3									
24					Median	100	130									
25					SD	92.15	114.3									
26				c	SE of Mean	21.14	26.21									
27					ic of wear	21.14	20.21									
28			Sa	mnle 1 vs S	ample 2 Two	-Sample t-T	·est									
29																
30		H0: Mean o	f Sample 1 =	= Mean of Sa	ample 2											
31						t-Test	Lower C.Val	Upper C.Val								
32		Method			DF	Value	t (0.025)	t (0.975)	P-Value							
33			ual Variance)		36	-0.974	-2.028	2.028	0.336							
34 35		` '	erthwaite (Un			-0.974	-2.032	2.032	0.337							
36		Pooled SD:		-												
36			with Alpha =	0.050												
38			•		0, Conclude	Sample 1 = :	Sample 2									
39		,			H0, Conclud	· ·	•									
40						<u> </u>										
41				Test of	Equality of V	ariances										
42																
43				Variance of	f Sample 1	8492										
44				Variance of	f Sample 2	13056										
45						<u>I</u>	<u>I</u>									
46		Numer	ator DF	Denomi	nator DF	F-Tes	t Value	P-Value								
47		1	18	1	18	1.5	538	0.370								
48		Conclusion	with Alpha =	0.05		-										
49		Two variand	ces appear to	o be equal												
50																
JU												1				

	Α	В	С	D	Е	F	G	Н		J	K	L	M				
1												<u> </u>					
2			Wilc	oxon-Mann-	Whitney San	nple 1 vs Sa	mple 2 Com	parison Test	for Uncense	or Full Data	Sets without	NDs					
3					T												
4				cted Options		10/00/000											
5		Dat	e/Time of Co	-	ProUCL 5.2												
6				From File		1_ProUCL_Input_Spec.Cond_TDS_a.xls											
7				Il Precision	OFF												
8			Confidence		95%												
9			Substantial		0.000	/B.A. I'	0 1 01	18 A 19	/ T 0:1								
10			elected Null H		-	Mean/Mediar		(Alternative									
11			Alternative I	Hypothesis	Sample 1 M	ean/Median	<> Sample 2	! Mean/Media	an —————								
12																	
13		0	-1 TDO /														
14		Sample 1 D			OE) (/I)												
15		Sample 2 Data: TDS from SC(SC*0.65) (mg/L)															
16		Raw Statistics															
17				ı			Commis 2										
18			Nimaka	r of Valid Ob		Sample 1	Sample 2										
19						19	19										
20			Number of	f Distinct Ob		18 48	16										
21					Minimum	360	67.6 416										
22					Maximum												
23					Mean	136.5 100	169.3 130										
24					Median SD	92.15	114.3										
25																	
26					E of Mean	21.14	26.21										
27				Wileeven Me	ann-Whitney	(A/MANAN Too											
28				VVIICOXOII-IVI	ariri-vvriiuriey	(VVIVIVV) TES	šί. 										
29																	
30		HO: Moon/M	ladian of Sa	male 1 = Me	an/Median o	f Comple 2											
31		no. Weathy	leulali oi Sa	inple i – Me	an/ivieulan o	or Sample 2											
32			Sar	mnlo 1 Dank	Sum W-Stat	335	1										
33			Jai	•	VMW U-Stat												
34				· ·	Mean (U)												
35				SD	(U) - Adj ties												
36			l ower II-S		/alue (0.025)												
37					'alue (0.975)												
38					VMW U-Stat												
39					nate P-Value												
40																	
41		Conclusion	with Alpha =	= 0.05													
42			=		ple 1 = Sam												
43			-,,														
44 45												1					
45		<u> </u>										1					

Appendix F

NJDEP Water Quality Data Exchange Selected Site Data

Near SWMP Station 6

Run At: 03/14/2024 05:39 pm

Organization	Location	Date	Depth	Ft/m	Media	Characteristic	Form	Fraction	Remark	Result	Units
NJDEP_BFBM	FIBI012	7/12/2010			Other	FIBI Rating				Good	None
NJDEP_BFBM	FIBI012	7/12/2010			Water	Dissolved oxygen (DO)				8.79	mg/l
NJDEP_BFBM	FIBI012	7/12/2010			Water	pН				7.01	None
NJDEP_BFBM	FIBI012	7/12/2010			Water	Specific conductance				198	mS/cm
NJDEP_BFBM	FIBI012	7/12/2010			Water	Temperature, water				20.04	deg C
NJDEP_BFBM	FIBI012	6/21/2017			Air	Barometric pressure				753	mmHg
NJDEP_BFBM	FIBI012	6/21/2017			Air	Temperature, air				23.7	deg C
NJDEP_BFBM	FIBI012	6/21/2017			Other	FIBI Rating				Fair	None
NJDEP_BFBM	FIBI012	6/21/2017			Other	Fish Index of Biotic Integrity				45.78	None
NJDEP_BFBM	FIBI012	6/21/2017			Other	Per. Abun. Cold & Nontolerant Coolwater Sp. (adj)				23.35	None
NJDEP_BFBM	FIBI012	6/21/2017			Other	Per. Abun. Dominant 3 Taxa (not Blacknose Dace)				27.01	None
NJDEP_BFBM	FIBI012	6/21/2017			Other	Percent Abundance Cyprinidae (adj)				19.81	None
NJDEP_BFBM	FIBI012	6/21/2017			Other	Percent Richness Benthic Insectivores				26.49	None
NJDEP_BFBM	FIBI012	6/21/2017			Other	Percent Richness Generalist Feeders				76.58	None
NJDEP_BFBM	FIBI012	6/21/2017			Other	Percent Richness of Rheophilic Species (adj)				60.07	None
NJDEP_BFBM	FIBI012	6/21/2017			Other	Per. Rich. of Lithophilic Spawners (minus w. suck)				57.95	None
NJDEP_BFBM	FIBI012	6/21/2017			Other	Tolerance Index					None
NJDEP_BFBM	FIBI012	6/21/2017			Water	Dissolved oxygen (DO)				8.48	mg/l
NJDEP_BFBM	FIBI012	6/21/2017			Water	Dissolved oxygen saturation				93.5	%
NJDEP_BFBM	FIBI012	6/21/2017			Water	Flow				13.88	cfs
NJDEP_BFBM	FIBI012	6/21/2017			Water	Inorganic nitrogen (nitrate and nitrite)	as N	Total		0.0816	mg/l
NJDEP_BFBM	FIBI012	6/21/2017			Water	Kjeldahl nitrogen		Total		0.277	_
NJDEP_BFBM	FIBI012	6/21/2017			Water	Phosphate-phosphorus	as P	Total		0.0317	_
NJDEP_BFBM	FIBI012	6/21/2017			Water	Specific conductance					uS/cm
NJDEP_BFBM	FIBI012	6/21/2017			Water	Temperature, water				20.1	deg C

D = Sample diluted, HT = Holding Time exceeded, J = Estimated, K= Less than, L = Greater than, NRP = No result possible, R = Less than Reporting Limit

Near SWMP Station 7

Run At: 03/14/2024 05:51 pm

Organization	Location	Date	Depth	Ft/m	Media	Characteristic	Form	Fraction	Remark	Result	Units
NJDEP_BFBM	BA50	8/29/2012			Air	Weather comments (text)				No rain day of sample, rain day before sample	None
NJDEP BFBM	BA50	8/29/2012			Water	Escherichia coli		Total		172.5	#/100ml
NJDEP BFBM	BA50	8/29/2012			Water	Fecal Coliform		Total			#/100ml
NJDEP BFBM	BA50	8/29/2012			Water	Temperature, water					deg C
NJDEP_BFBM	BA50	9/5/2012			Air	Weather comments (text)				Rain day before & day of Sample	None
NJDEP BFBM	BA50	9/5/2012			Water	Escherichia coli		Total		1732.9	#/100ml
NJDEP_BFBM	BA50	9/5/2012			Water	Fecal Coliform		Total			#/100ml
NJDEP_BFBM	BA50	9/5/2012			Water	Temperature, water				23.9	deg C
NJDEP_BFBM	BA50	9/12/2012			Air	Weather comments (text)				No rain day of sample or day before sample	None
NJDEP_BFBM	BA50	9/12/2012			Water	Escherichia coli		Total		52.9	#/100ml
NJDEP_BFBM	BA50	9/12/2012			Water	Fecal Coliform		Total		49	#/100ml
NJDEP_BFBM	BA50	9/12/2012			Water	Temperature, water				18.5	deg C
NJDEP_BFBM	BA50	9/19/2012			Air	Weather comments (text)				Rain day before & day of Sample	None
NJDEP_BFBM	BA50	9/19/2012			Water	Escherichia coli		Total		727	#/100ml
NJDEP_BFBM	BA50	9/19/2012			Water	Fecal Coliform		Total		920	#/100ml
NJDEP_BFBM	BA50	9/19/2012			Water	Temperature, water				20.1	deg C
NJDEP_BFBM	BA50	9/26/2012			Air	Weather comments (text)				Rain day of sample, no rain day before	None
NJDEP_BFBM	BA50	9/26/2012			Water	Escherichia coli		Total		48.8	#/100ml
NJDEP_BFBM	BA50	9/26/2012			Water	Fecal Coliform		Total		23	#/100ml
NJDEP_BFBM	BA50	9/26/2012			Water	Temperature, water				17.1	deg C

D = Sample diluted, HT = Holding Time exceeded, J = Estimated, K= Less than, L = Greater than, NRP = No result possible, R = Less than Reporting Limit

Near SWMP Station 8

Run At: 03/14/2024 06:03 pm

Organization	Location	Date	Depth	Ft/m	Media	Characteristic	Form	Fraction	Remark	Result	Units
NJDEP_BFBM	BA48	8/29/2012			Air	Weather comments (text)				No rain day of sample, rain day before sample	None
NJDEP_BFBM	BA48	8/29/2012			Water	Escherichia coli		Total		5.2	#/100ml
NJDEP_BFBM	BA48	8/29/2012			Water	Temperature, water				23.8	deg C
NJDEP_BFBM	BA48	9/5/2012			Air	Weather comments (text)				Rain day before & day of Sample	None
NJDEP_BFBM	BA48	9/5/2012			Water	Escherichia coli		Total		56.3	#/100ml
NJDEP_BFBM	BA48	9/5/2012			Water	Temperature, water				23.3	deg C
NJDEP_BFBM	BA48	9/12/2012			Air	Weather comments (text)				No rain day of sample or day before sample	
NJDEP_BFBM	BA48	9/12/2012			Water	Escherichia coli		Total			#/100ml
NJDEP_BFBM	BA48	9/12/2012			Water	Temperature, water				21.7	deg C
NJDEP_BFBM	BA48	9/19/2012			Air	Weather comments (text)				Rain day before & day of Sample	None
NJDEP_BFBM	BA48	9/19/2012			Water	Escherichia coli		Total		34.1	#/100ml
NJDEP_BFBM	BA48	9/19/2012			Water	Temperature, water				21.2	deg C
NJDEP_BFBM	BA48	9/26/2012			Air	Weather comments (text)				Rain day of sample, no rain day before	None
NJDEP_BFBM	BA48	9/26/2012			Water	Escherichia coli		Total		8.6	#/100ml
NJDEP BFBM	BA48	9/26/2012			Water	Temperature, water				19.7	deg C

D = Sample diluted, HT = Holding Time exceeded, J = Estimated, K= Less than, L = Greater than, NRP = No result possible, R = Less than Reporting Limit

Near SWMP Station 10

Run At: 03/15/2024 09:43 am

Organization	Location	Date	Depth	Ft/m	Media	Characteristic	Form	Fraction	Remark	Result	Units
NJDEP BFBM	FIBI081	7/20/2009	Бори	1 0111	Other	FIBI Rating	1 01111	114041011	Roman		None
NJDEP BFBM	FIBI081	7/20/2009			Water	Dissolved oxygen (DO)					mg/l
NJDEP BFBM	FIBI081	7/20/2009			Water	pH					None
NJDEP BFBM	FIBI081	7/20/2009			Water	Specific conductance					mS/cm
NJDEP_BFBM	FIBI081	7/20/2009			Water	Temperature, water					deg C
NJDEP_BFBM	FIBI081	7/2/2012			Air	Barometric pressure					mmHg
NJDEP BFBM	FIBI081	7/2/2012			Air	Temperature, air					deg C
NJDEP BFBM	FIBI081	7/2/2012			Water	Dissolved oxygen (DO)					mg/l
NJDEP_BFBM	FIBI081	7/2/2012			Water	Dissolved oxygen saturation				89.3	_
NJDEP BFBM	FIBI081	7/2/2012			Water	рН				6.88	None
NJDEP_BFBM	FIBI081	7/2/2012			Water	Specific conductance					uS/cm
NJDEP_BFBM	FIBI081	7/2/2012			Water	Temperature, water					deg C
NJDEP BFBM	FIBI081	6/21/2017			Water	Escherichia coli		Total			#/100ml
NJDEP_BFBM	FIBI081	6/21/2017			Water	Temperature, water					deg C
NJDEP_BFBM	FIBI081	6/26/2017			Water	Escherichia coli		Total			#/100ml
NJDEP_BFBM	FIBI081	6/26/2017			Water	Temperature, water				22.7	deg C
NJDEP_BFBM	FIBI081	7/5/2017			Water	Escherichia coli		Total			#/100ml
NJDEP_BFBM	FIBI081	7/5/2017			Water	Temperature, water					deg C
NJDEP_BFBM	FIBI081	7/10/2017			Water	Escherichia coli		Total			#/100ml
NJDEP_BFBM	FIBI081	7/10/2017			Water	Temperature, water				24	deg C
NJDEP_BFBM	FIBI081	7/19/2017			Water	Escherichia coli		Total			#/100ml
NJDEP_BFBM	FIBI081	7/19/2017			Water	Temperature, water				25.2	deg C
NJDEP_BFBM	FIBI081	8/21/2017			Air	Barometric pressure				761	mmHg
NJDEP_BFBM	FIBI081	8/21/2017			Air	Temperature, air				23	deg C
NJDEP_BFBM	FIBI081	8/21/2017			Other	FIBI Rating					None
NJDEP_BFBM	FIBI081	8/21/2017			Water	Dissolved oxygen (DO)				8.43	mg/l
NJDEP_BFBM	FIBI081	8/21/2017			Water	Dissolved oxygen saturation				96.1	%
NJDEP_BFBM	FIBI081	8/21/2017			Water	Flow				17.73	cfs
NJDEP_BFBM	FIBI081	8/21/2017			Water	Inorganic nitrogen (nitrate and nitrite)	as N	Total		0.165	mg/l
NJDEP_BFBM	FIBI081	8/21/2017			Water	Kjeldahl nitrogen		Total		0.215	mg/l
NJDEP_BFBM	FIBI081	8/21/2017			Water	рН				7.77	None
NJDEP_BFBM	FIBI081	8/21/2017			Water	Phosphate-phosphorus	as P	Total		0.0271	•
NJDEP_BFBM	FIBI081	8/21/2017			Water	Specific conductance				251.9	uS/cm
NJDEP_BFBM	FIBI081	8/21/2017			Water	Temperature, water				21.78	deg C
NJDEP_BFBM	FIBI081	8/14/2018			Water	Escherichia coli		Total			#/100ml
NJDEP_BFBM	FIBI081	8/14/2018			Water	Temperature, water				23.5	deg C
NJDEP_BFBM	FIBI081	8/21/2018			Water	Escherichia coli		Total		88	#/100ml
NJDEP_BFBM	FIBI081	8/21/2018			Water	Temperature, water					deg C
NJDEP_BFBM	FIBI081	8/28/2018			Water	Escherichia coli		Total			#/100ml
NJDEP_BFBM	FIBI081	8/28/2018			Water	Temperature, water					deg C
NJDEP_BFBM	FIBI081	9/4/2018			Water	Escherichia coli		Total			#/100ml
NJDEP_BFBM	FIBI081	9/4/2018			Water	Temperature, water					deg C
NJDEP_BFBM	FIBI081	9/11/2018			Water	Escherichia coli		Total			#/100ml
NJDEP_BFBM	FIBI081	9/11/2018			Water	Temperature, water				20.1	deg C

D = Sample diluted, HT = Holding Time exceeded, J = Estimated, K= Less than, L = Greater than, NRP = No result possible, R = Less than Reporting Limit